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Microbial Assessment and Proximate Composition of Pepper (Capsicum annum) and Tomato (Solanum lycopersicum) Displayed for Sales

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Authors' contributions

This work was carried out in collaboration among all authors. Author ON J,P' designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Author AB managed the analyses of the study. Authors ON and AB managed the literature searches.

All authors read and approved the final manuscript.

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ABSTRACT

Tomato (*Solanum lycopersicum*) and pepper (*Capsicum annuum*) are among the world's most important vegetable crops. A total of 20 samples made up of fresh tomatoes, spoilt tomatoes, fresh pepper and spoilt pepper were analysed in this study. Standard microbiological practices were carried out on the samples. Total Heterotrophic Bacterial (THB) Count for Spoilt tomatoes ranged from 1.98x10⁷cfu/g to 2.39x10⁷cfu/g. Fresh tomatoes samples had a THB counts ranging from 1.43x10⁶cfu/g to 2.14x10⁶cfu/g. Spoilt pepper total heterotrophic bacteria counts ranged from 1.46x10⁷cfu/g to 2.01x10⁷cfu/g. Fresh pepper had a THB count of 1.22x10⁶cfu/g to 1.66x10⁶cfu/g. Spoilt tomatoes samples had a higher fungal count that ranged from 3.5x10⁵cfu/g to 5.25x10⁵cfu/g while the fresh tomatoes samples had lower fungal counts ranging from 1.95x10⁴cfu/g to 3.45x10⁴cfu/g. The spoilt pepper had fungal counts ranging from 1.45x10⁵cfu/g to 2.65x10⁵cfu/g which was higher than that of fresh pepper with a low count of 1.6x10⁴cfu/g to 2.75x10⁴·cfu/g. The bacterial isolates identified during this study are *Staphylococcus* sp., *Escherichia coli*, *Bacillus* sp., *klebsiella* sp., *Pseudomonas* sp., *Shigella* sp., *Protues* sp., *Enterobacter* sp., *Citrobacter* sp., *Lactobacillus* sp. *Micrococcus* sp. *Listeria* sp. *Streptococcus* sp. and *Serratia* sp. Spoilt tomatoes had the highest coliform count 1.02x10⁵cfu/g to 9.0x10⁵cfu/g while fresh tomatoes had a lower count

of 4.65x10⁵cfu/g to 6.75x10⁵cfu/g. Spoilt pepper recorded coliform counts ranging from 3.8x10⁵cfu/g to 9.4x10⁵cfu/g. Most of the fungi isolated from both the tomatoes and pepper samples were molds and yeast which include *Aspergillus niger, Aspergillus flavus, Candida* sp., *Saccharomyces* sp., *Penicillium* sp. *Mucor* sp. and *Fusarium* sp. Adequate cooking is recommended before consumption.

Keywords: Tomatoes; pepper, fresh; spoilt; pathogen.

1. INTRODUCTION

All foods are great sources for nutrient that can contribute to growth. Food materials contain organic substances in plenty and also sufficient amount of water. They may be either neutral or slightly acidic in nature (Singh, 2013). They are subjected to natural contamination by many different kinds of microorganisms, including pathogens. Food spoilage refers to various changes in which the food becomes less palatable or even toxic to consumers these changes may be accompanied by alterations in taste, smell, appearance or texture. Numerous microbial defects of agricultural crops are characterized by the types of microorganisms responsible for their deterioration [1]. fruits and vegetables are usually kept on tables and in baskets for prospective customers in the open markets until it is bought, thereby making it easy for further microbial infections beside those associate with these whole fruit and vegetables surface and those from adjacent infected fruits [2].

Fruits and vegetables however, have serious challenges to their existence. These include changes in climatic condition, pests and microbial attack. Over the years, there has been an increase in the need to identify and isolate the microorganisms associated with the spoilage as a way of finding a means of controlling it [3].

Susceptibility of fruits to microbial deterioration is largely due to differential chemical composition such as pH and moisture contents are associated greater with predisposition microbial spoilage. The occurrence of fungal spoilage of fruits is also recognized as a source of potential health hazard to man and animal. This is due to their production of mycotoxins (naturally occurring toxic chemical often of aromatic structure) which are capable of producing aflatoxin in man, following ingestion or inhalation.

These fruits are usually displayed on benches and in baskets for prospective customers in the open markets until sold, thereby exposing them to further microbial infection beside those

associated with these whole fruit surface and those from adjacent infected fruits [2]. In developing countries, like Nigeria, post-harvest deterioration are often more severe due to inadequate storage and transportation facilities. Microbial fruits infection may occur during the growth season, harvesting, handling, transport post-harvest storage and marketing conditions, or after purchasing by the consumer. Fruits contain high levels of sugars and nutrient elements and their low pH values make them particularly desirable to fungal decay (Singh and Sharma, 2007). Studies by Li-Cohen and Bruhn [4] have shown that fungi can survive and/or grow on fresh produce and that the nutrient content (carbohydrate, protein and fat) of fresh produce support pathogens.

Fruits are affected by a wide array of microorganisms causing its decay. Spoilage microorganisms can be introduced to the crop on the seed itself, during crop growth in the field, during harvesting and post-harvest handling or during storage and distribution (loading and offloading) [5]. Those types of soil-borne spoilage microbes that occur on produce are the same spoilage microorganisms that are present on harvesting equipment, on handling equipment, in the packaging house, in the storage facility, and food contact surfaces throughout the distribution chain. Therefore, early intervention durina development crop harvesting through the use of good agricultural practices (GAP) will provide dramatic reductions in the yield loss due to deterioration at all subsequent steps in the food [5].

Tomato and pepper fruits are very rich in mineral, vitamins, and carbohydrate [6] In view of these, these fruits are often attacked by microorganisms especially after harvest, thus a fast and high rate of spoilage is often observed in storage [5].

Tomato and pepper fruits are very rich in mineral, vitamins, and carbohydrate and hence as may serve as good breeding sites for microbial proliferation. Growth of microorganisms on these fruits contributes to their spoilage and deterioration and render them unhealthy for

consumption as they may harbour microorganisms of potential public health importance.

This study sets out to determine the microbial profile of fresh and spoilt pepper (Capsicum annuum) and tomatoes (Solanum lycopersicum) sold in the market in Port Harcourt. Nigeria.Microbial contamination sources Tomato and pepper fruits include raw materials and contact with processing equipment. The microorganisms that exist on the surfaces of raw, whole produce appear to be the major source of microbial contamination and consequent spoilage of these fruit and vegetables. Sapers' et al., (2001) reported that, compared with the good practices, surface sanitization decontamination treatment or an ineffective antimicrobial treatment on whole tomatoes and pepper resulted in premature microbiological spoilage of tomato and pepper. Products can also be contaminated by microorganisms through contact by people or equipment during processing possibly by air processing and packaging steps, especially in market places

2. MATERIALS AND METHODS

Twenty samples each of fresh strong tomatoes and pepper and broken spoilt pepper and tomato (were sourced randomly from different vendors in Choba market within Obio-Akpor Local Government Area, of Rivers State and was transported in a sterilized bag to the Microbiology laboratory for analysis.

Commercially available nutrient media were used for isolation, identification and characterization of microorganisms. The media used include: Nutrient agar, Peptone water, Salmonella

Shigella agar, MacConkey agar, Mannitol salt agar, Potato dextrose agar.

All the media, diluents and glass wares used (Petri plates, bijou bottles, test tubes, pipette) were sterilized by autoclaving at 121°C for 15 minutes at 15 pounds per square inch (psi), unless stated otherwise and the work benches were disinfected with disinfectant, the wire loop was sterilized by passing through red hot flame from a Bunsen burner before use.

2.1 Isolation of Microorganisms

10g of the sample from different locations were added into 90 ml of peptone water, swirled and allowed to stay for few minutes after which a tenfold serial dilution was done by pipetting 1 ml from the stock solution into the next test tube (10 2), the process was done repeatedly up to (10 5).From the prepared diluents, 0.1 ml of each last two prepared dilutions were transferred into sterile Petri plates containing the different media used and was spread gently using sterile glass rod. The plates were incubated at 37°C for 18-24 hours for the bacteriological media used for bacteria growth and Potato dextrose agar for fungi isolation. The microbial count for each sample was obtained from the previously incubated Petri plates and was expressed as a colony forming unit (cfu/g).

2.2 Identification and Characterization of Isolates

Single colonies of bacteria growth were randomly selected from different media plates based on their morphology and were sub-cultured and incubated at 37°C for 24 hours to obtain pure colonies.

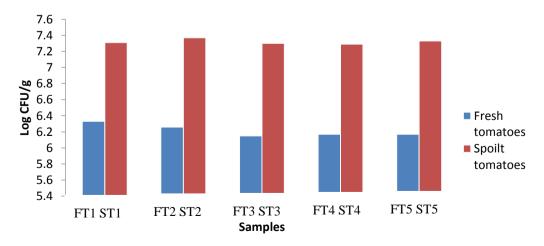


Fig. 1. Heterotrophic bacteria count from fresh and spoilt tomatoes samples

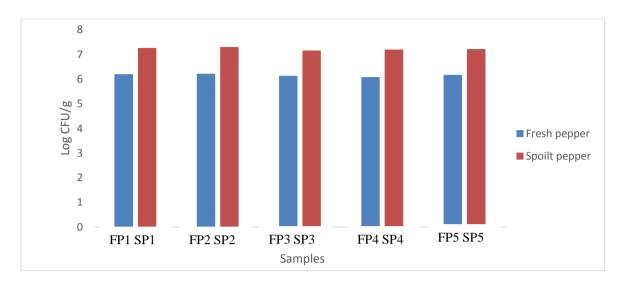


Fig. 2. The heterotrophic bacteria count from fresh and spoilt pepper samples

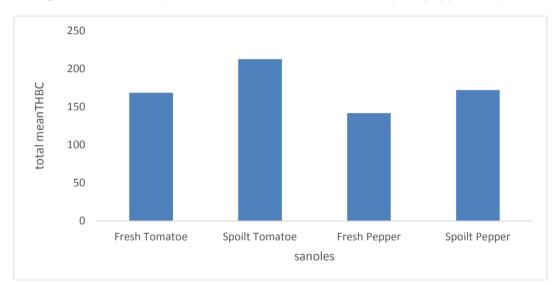


Fig. 3. Total mean of THBC in fresh tomatoes /pepper, and spoilt tomatoes and pepper

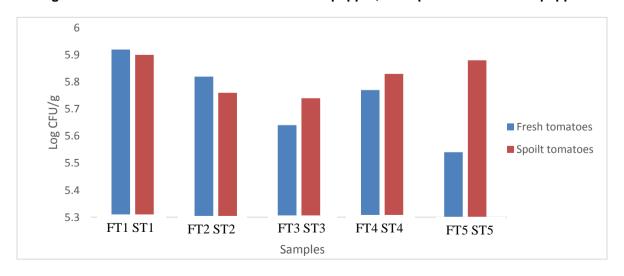


Fig 4. Staphylococcus count from fresh and Spoilt tomatoes samples

2.2.1 Examination of bacteria

Isolates were identified based on their morphological and cultural characteristics on growth media. Identification materials, reagents and protocols according to (Cheesebrough, 2000) were used to identify discrete colonies from the bacteriological media of sub-cultured isolates.

2.2.2 Examination of fungi

The cultural characteristics of each fungi isolates were identified according to their colour, shape and the cell morphology was done based on mycelia, hyphae, septate, spore formation using lactophenol blue. A piece of the mycelium from the Petri plates was mounted on a clean grease free slide using a sterile wire loop and covered with a cover slip, after which a drop of lactophenol cotton blue was added and examined with the microscope.

Proximate analysis was done as described by Garuba et al., [7].

2.3 Statistical Analysis

All analysis was done in duplicates for each of the samples, and data were reported for duplicate analyses. All statistical analyses were carried out using analysis of variance (ANOVA). Significance of the differences was ascribed at the 0.05 level for ANOVA

3. RESULTS AND DISCUSSION

3.1 Microorganisms Isolated from Pepper and Tomatoes Samples

A total of one hundred and fourteen (114) bacterial isolates were obtained from both the tomatoes and pepper samples; 23 from fresh pepper samples and 28 from the spoilt pepper samples, 26 from fresh tomatoes samples and 37 from spoilt tomatoes samples. The bacterial isolates identified during this study include Staphylococcus sp., Escherichia coli, Bacillus sp., klebsiella sp., Pseudomonas sp., Shigella sp., Proteus sp., Enterobacter sp., Citrobacter sp.

The percentage frequency of occurrence of bacteria isolated from the fresh tomatoes samples are in decreasing order *Klebsiella* sp. (19.23%), *Lactobacillus* sp. (15.38%), *Bacillus* sp. (15.38%), *Micrococcus* sp. (11.5%), *Citrobacter* sp. (11.5%) and *Staphylococcus* sp. (11.5%) and *Enterobacter* sp. (7.69%). For the spoilt tomatoes samples, *Lactobacillus* sp.

(16.2%), Klebsiella sp. (13.51%) and Bacillus sp. (13.51%), Enterobacter sp. (8.10%), Proteus sp. (8.10%), Listeria sp. (8.10%), Shigella sp. (5.40%) (5.40%). Pseudomonas sp. and Escherichia coli (5.40%). Bacteria isolated from the fresh pepper had a percentage frequency of occurrence with Enterobacter sp., Streptococcus sp. and *Micrococcus* sp. had a least occurrence of (13.04%), Staphylococcus sp. and Klebsiella sp. (17.39%), and *Bacillus* sp. (26.08%). Spoilt pepper samples had a frequency of occurrence for Serratia sp. (7.14%), Enterobacter sp. and Escherichia coli (10.71%), Proteus sp. and Pseudomonas sp. (14.28%), Klebsiella sp. (21.14%). These are represented in the figures.

Results obtained from this research shows that tomatoes and pepper harbor an array of microorganisms. The total heterotrophic counts for fresh tomatoes samples ranging from 1.43×10^6 cfu/g to 2.14×10^6 cfu/g is lower than that of spoilt tomatoes with counts ranging from 1.98×10^7 cfu/g to 2.39×10^7 cfu/g. This is higher than results reported by Ibrahim et al., 8]. According to Gosh [9], the high-water content of tomatoes makes them readily susceptible to The invasion microbial spoilage. microorganisms and their quick multiplication can cause spoilage. This is a possible reason for the increased counts in the spoilt tomatoes than in the fresh one. Also, advisory guidelines for microbiological quality have suggested that satisfactory food products should contain no more than 10⁵ cfu/g of starter organisms [8], hence, considering counts obtained for both fresh and spoilt tomatoes samples in this study, the allowable limit was exceeded.

The spoilt tomatoes had significantly higher Staphylococcus count ranging from 5.5x10⁵ cfu/g to 7.95×10^5 cfu/g(p<0.05)g while fresh tomatoes had counts of 3.5x10⁵ cfu/g to 6.5x10⁵ cfu/g. The Staphylococcus limit set by the Food and Drug Agency (FDA) for foods is <10⁵. Results obtained from this study fell within the acceptable Staphylococcus limits for foods, the coliform count of fresh tomatoes ranged from 4.56×10^5 cfu/g to 6.75×10^5 cfu/g, while spoilt tomatoes had a count of 1.02x10⁵ cfu/g to 9.0x10⁵ cfu/g. The work of Shenge et al., [10] tried to establish possible pathways for coliform contamination of tomatoes. Although, no one single pathway was conclusive enough, they suggested that lack of hygienic practices in handling the product, source of irrigation water, cross contamination during transportation and other unknown factors were responsible for contaminating tomatoes.

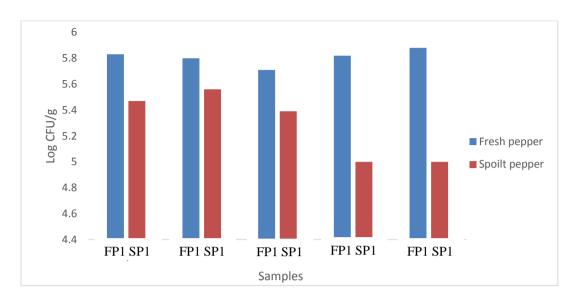


Fig. 5. The Staphylococcus count from fresh and Spoilt pepper samples

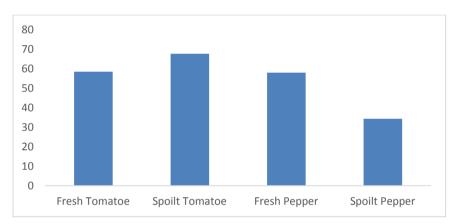


Fig. 6. Total mean *Staphylococcus* count in fresh tomatoes /pepper, and spoilt tomatoes and pepper

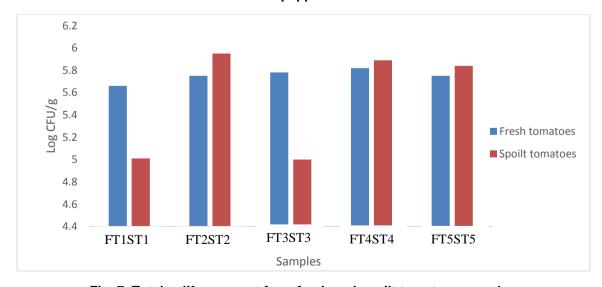


Fig. 7. Total coliform count from fresh and spoilt tomatoes samples

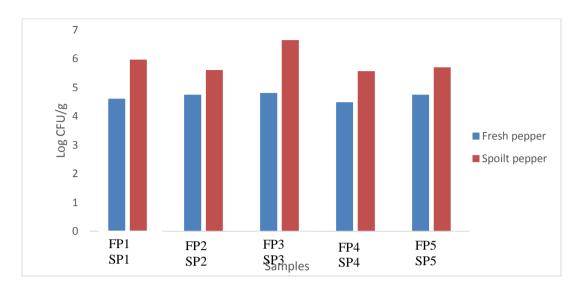


Fig. 8. Coliform count from fresh and spoilt pepper samples

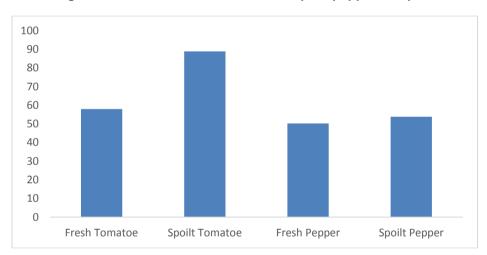


Fig. 9. Total mean of coliform count in fresh tomatoes/pepper and spoilt tomatoes and pepper

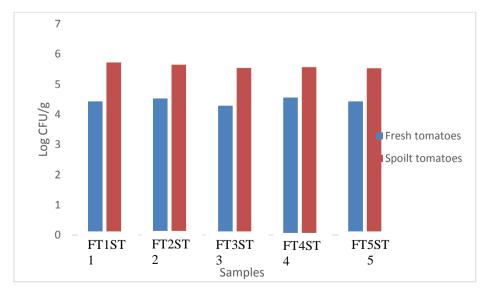


Fig. 10. Total heterotrophic fungal count from fresh and spoilt tomatoes samples

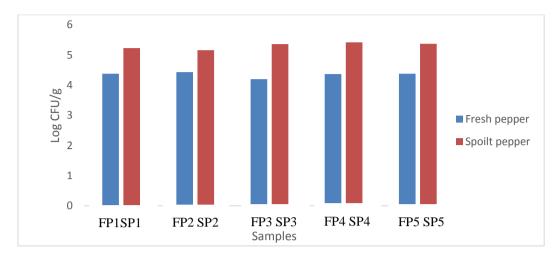


Fig. 11. Total heterotrophic fungal count from spoilt pepper samples

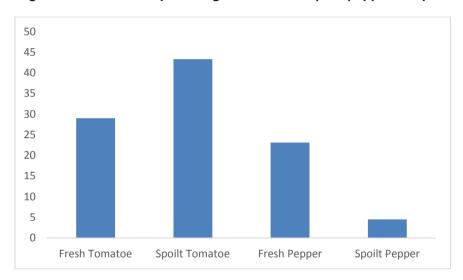


Fig. 12. Total mean of fungal count in fresh tomatoes/pepper and spoilt tomatoes and pepper

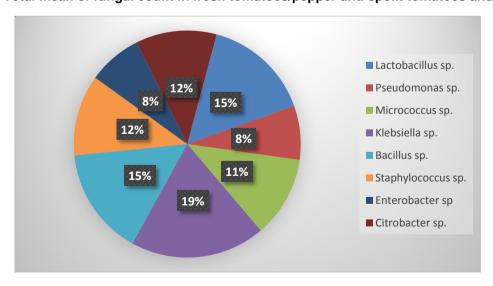


Fig. 13. Percentage frequency of bacterial isolates obtained from fresh tomatoes samples

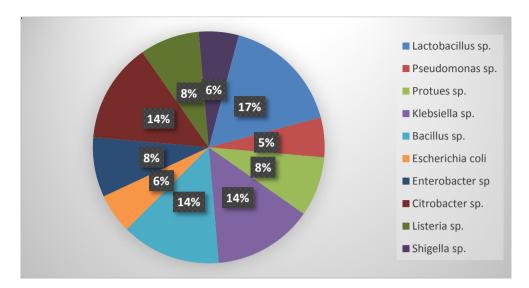


Fig. 14. Percentage frequency of bacterial isolates obtained from spoilt tomatoes sample

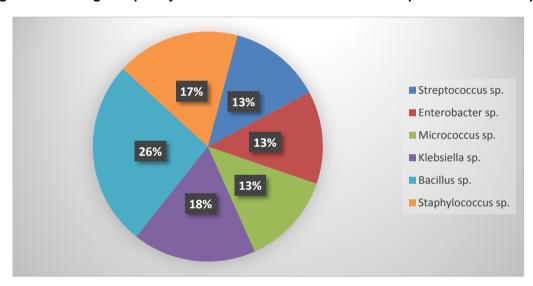


Fig. 15. Percentage frequency of bacterial isolates obtained from fresh pepper samples

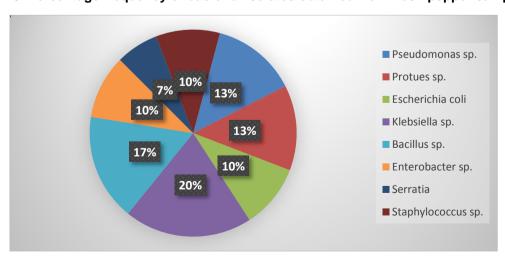


Fig. 16. Percentage frequency of bacterial isolates obtained from spoilt pepper samples

Table 1. Proximate analysis on tomatoes and pepper samples

S/no	Sample code	Moisture	Ash	СНО	Protein	Lipid	Fibre
1	Fresh tomatoes	91.50	0.98	5.00	3.25	0.067	<0.01
2	Fresh pepper	84.67	0.63	7.87	2.79	1.72	2.42
3	spoilt tomatoes	91.09	0.35	5.94	2.45	0.17	< 0.01
4	spoilt pepper	87.39	0.81	3.90	3.82	1.86	2.52

The fungal counts of fresh tomatoes ranged from 1.95x10⁴ cfu/g to 3.65x10⁴ cfu/g and spoilt tomatoes had a count of 3.5x10⁵ cfu/g to 5.25x10⁵ cfu/g.which was significantly high in the spoilt tomatoes when compared to the fresh sample(p<0.05) Obunukwu et al., [11] reported fungal counts for spoilt fresh tomatoes stored at ambient temperature similar to fungal counts obtained from fresh tomatoes in this study. In another study, Mwekaven et al., [12] reported counts similar to those obtained from spoilt tomatoes in this study.

Bacterial species isolated from fresh and spoilt tomatoes samples include Lactobacillus sp., Pseudomonas sp., Micrococcus sp., Klebsiella sp., Bacillus sp., Staphylococcus Enterobacter sp., Citrobacter sp. Escherichia coli was isolated only from the spoilt tomatoes samples. The organism isolated in this study is consistent with the works of Ibrahim et al., [8], Obunukwu et al., [11], and Mwekaven et al., [12] who isolated similar bacterial species from fresh and spoilt tomatoes respectively. Six (6) bacterial isolates- Lactobacillus fermenti, Pseudomonas stutzeri, Listeria monocytogenes, Leuconostoc sp., Rothia sp., were found in tomato. [13] isolated Leuconostoc sp. and Lactobacillus sp. as tomatoes natural flora which could participate spoilage of such fruit. Presence Micrococcus sp. in foods causes dental decay and Bacillus subtilis causes flat sour of fruits and denaturing of body (Nester et al., 1995). Most of the bacterial isolates are opportunistic infection agents and could lead to food borne bacterial diseases [12].

Eight (8) fungal isolates (filamentous fungi) were identified in both fresh and spoilt tomatoes samples. These fungal isolates are Aspergillus niger, Penicillium sp., Saccharomyces sp., Mucor sp., Candida sp., Aspergillus flavus, Aspergillus fumigates and Rhizopus stolonifer. The isolation of Aspergillus niger, Rhizopus stolonifer, Mucor species from rotten tomato confirmed the studies of [14]. According to Ghosh [9], fungi were the source of spoilage in most of tomatoes samples rather than bacteria. Also, Akinmusire [1] had reported that A. flavus and A. fumigatus caused

tomato spoilage. These fungal isolates are also potential sources of disease condition. Some like species of *Aspergillus* are known to produce mycotoxins which can be fetal when consumed [8].

Fresh pepper had a total heterotrophic bacterial count ranging from 1.22x10⁶ cfu/g to 1.66x10⁶ cfu/g. This was significantly lower than that of the spoilt pepper with counts ranging from 1.46x10' $2.01x10^7$ cfu/a to cfu/g, (p>0.05)Staphylococcus counts of fresh pepper ranged from 5.2x10⁵ cfu/g to 7.7x10⁵ cfu/g while that of spoilt pepper ranged from 2.5x105 cfu/g to 4.0x10⁵ cfu/g. Coliform counts of Spoilt pepper had a count ranging from 3.8x10⁵ cfu/g to 9.4x10⁵ cfu/g and fresh pepper had a count of 3.1×10^4 cfu/g to 6.5×10^4 cfu/g, the spoilt samples counts were significantly higher than the counts obtained for the fresh smples.. (p<0.05) Bacterial isolates obtained from fresh and spoilt respectively. pepper samples, These includ(p<0.05e Enterobacter sp., Staphylococcus Streptococcus Klebsiella sp., Micrococcus sp. while Pseudomonas Proteus sp., Escherichia coli, Klebsiella sp., Bacillus sp., Enterobacter sp., and Serratia sp.

While the fungal isolates obtained from the pepper samples which include Aspergillus niger, Penicillium sp., Rhizopus sp., Aspergillus Rhizopus stolonifer, Mucor sp., fumigates, Saccharomyces sp. These agree partly with the findings of [4] who discovered that species of fungi associated with the spoilage of some edible fruits including tomatoes include species of Aspergillus, fusarium, Penicillium and Rhizopus. The most frequent of the isolated molds from tomato and pepper belongs to Aspergillus sp. and Penicillium sp. and these confirms their prevalence in fruits and foods exposed to tropical humid climate, thus consisting potential health risks to consumers. In a similar study carried out on fungi associated with the spoilage of postharvest tomato fruits sold in major markets in Awka, Nigeria, Aspergillus niger, Rhizopus stolonifer, Fusarium oxysporum, Saccharomyces Alternaria alternata, Penicillium cerevisiae. digitatum and Geotrichum candidum were identified ([15]. Abel-Mallek et al., [16] also reported on the common occurrence of Aspergillus niger in healthy tomato fruits collected from markets in Assiut, Egypt. Several studies have also reported that Aspergillus sp. are associated with spoilage of tomatoes, apricot, orange, lemon, peach, apple, kiwi, mango etc. [17]. Onuorah and Orji, [15] showed that Aspergillus had the highest decay diameter among other fungi associated with tomatoes spoilage. Studies have shown that Asperaillus produce aflatoxins. Aflatoxins are associated with some diseases in live stocks and humans throughout the world. Aspergillus flavus is the main producer of the well-known carcinogenic aflatoxins and its presence in food is of huge concern in terms of food safety, and they are toxic at low concentrations [18]. The dominance of Aspergillus in rotten tomatoes could pose a serious health risk especially when the tomatoes are not well cooked. Healthy tomatoes fruit should be preferred as they seldom contain microbes [19].

Penicillium sp. were found next to Aspergillus in abundance. Fusarium are among the most important genera of mycotoxigenic fungi [20]. The mycotoxins are of greatest agro-economic importance. Some molds are capable of producing more than one mycotoxin and some mycotoxins are produced by more than one fungal species [20].

Fruits and vegetables are very important and have high dietary and nutritional qualities. The importance of these fruits with its nutritional and other dietary factors cannot be over emphasized. Its spoilage often results to wastage of economic resources as well as food poisoning, especially, when consumed. From the results obtained in this study, it was discovered that few organisms encountered are food borne pathogens. It is also revealed that some spoilage microorganism (mostly fungi) gained access into these fruits during the processes of cultivating, harvesting, and packing and environmental contaminant which have in one time or the other been involved in food poisoning. The prevalence frequency of occurrence of fungi was higher than that of bacteria in both fruits. The high amounts of fungi and bacteria demand that appropriate control measures against infection should be employed. Adequate microbiological knowledge and hygienic handling practices of these produce would help minimize wastes due to deterioration. It is therefore, important that both the farmers who harvest and package the fruits into bags for

transportation, the marketers, and consumers take necessary precautions to prevent contamination and eating of contaminated fruits. This will however, enhance reduction in the risk of microbial toxins that are deleterious to human health.

The result obtained from Proximate analysis indicates that the fresh tomatoes sample had higher Moisture contents, Ash, Lipid and Protein. The carbohydrate content for fresh tomatoes was less than those of spoilt tomatoes sample. Both fresh and spoilt tomatoes sample showed they have equal fiber content. The spoilt pepper sample had higher Moisture content, Ash, Protein, Lipid and Fiber content. The fresh pepper sample however showed a higher Carbohydrate content. The result however, contrasts with the works of Ikuomola et al., [21], Garuba et al., [7], Ismail et al., [22] who had varying values for Moisture, Ash, Carbohydrates, Protein, Lipid and Fibre.

4. CONCLUSION

It is concluded that most of the fungi isolated from both the tomatoes and pepper samples were molds and yeast which include *Aspergillus niger*, *Aspergillus flavus*, *Candida sp.*, *Saccharomyces sp.*, *Penicillium sp. Mucor sp.* and *Fusarium sp.* Adequate cooking is recommended before consumption.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Akinmusire OO. Fungal species associated with the spoilage of some edible fruits in Maiduguri, Northern Eastern Nigeria. Advances in Environmental Biology. 2011;5(1):157-161.
- Baiyewu NA. 2. RA, Amusa Ayoola OA, Babalola OO. Survey of the postharvest diseases and aflatoxin contamination of marketed Pawpaw fruit (Carica papaya L) in South Western Nigeria..African Journal of Agricultural Research. 2007;2(4):178-181. Cspringer science + Business Media, LLC. 2007;13-183
- 3. Akinyele BJ, Akinkunmi CO. Fungi associated with the spoilage of berry and

- their reaction to magnetic field. Journal of Yeast andFungal Research. 2012;3(4):49-57.
- 4. Li- Cohen E, Bruhn CM.Safety of consumer handling of fresh produce. Journal of food production. 2002; 65(8):1287-1297.
- Barth M, Hankinson TR, Zhuang H, Breidt F. Microbiological spoilage of fruits and vegetables in W.H.Sperber, M.P.Doyle (editions), Compendium of the Microbiological Spoilage of Foods and Beverages, Food Microbiology and safety; 2009.
- 6. Udoh JD, Ndoh AB, Asuquo EP, Nyandoh UN. Crop production Techniques for the Tropics. Concept publications. Limited, Lagos. 2005;261-265
- 7. Garuba T, Mustapha OT, Oyeyiola GP. (Shelf life and proximate composition of tomato (*Solanum lycopersicum* L.) fruits as influenced by storage methods. Ceylon Journal of Science. 2018;47(4):387-393
- 8. Ibrahim AD, Musa Sani KA, Aliero AA, Yusuf BS. Microorganisms associated with the production of volatile compounds in spoilt tomatoes. Research in Biotechnology. 2011;2(2):82-89.
- Gosh A. Identification of Microorganisms responsible for spoilage of tomato (*Lycopersicon* esculentum) fruit. Journal of Phytology. 2009;1(6):414-416.
- 10. Shenge KC, Clement MZW. RA, Yakubu LL, Omolehin Erbaugh JM, Miller SA, Lejeune JT. Contamination of **Tomatoes** with Coliforms and Escherichia Coli on Farms and in Markets of Northwest Nigeria. Journal of Food Protection. 2015;78(1):57-
- Obunukwu GM, Dike KS, Nwakwasi GE. Isolation and Identification of Microbial Deteriogens of Fresh Tomatoes Stored at Ambient Temperature. Microbiology Research Journal International. 2018; 26(1):1-8.
- 12. Mwekaven SS, Wanen I, Yange T, Alye CT, Aorkwagh MT. Identification and Enumeration of Microorganisms Associated with Post-Harvest Spoilage of Tomatoes Lycopersicum esculentum Sold in Gboko metropolis. Journal of Scientific

- and Engineering Research. 2019;6(6):153-158.
- Ajayi AO. Nature of Tomatoes Microflora under storage. American Journal of Experimental Agriculture. 2013;3(1):89-101.
- 14. Chuku EC. Ogbonna DN. Onuegbu BA, Adeleke MTV. Comparative studies on the fungi and biochemical characteristics of snake (Trichosanthes Curcumerina linn) and Tomato (Lycopersicon Esculentum Mill) in Rivers State, Nigeria. Journal of **Applied** sciences. 2008;8(1):168-172
- Onuorah S, Orji MU. Fungi Associated with the Spoilage of Post-harvest Tomato Fruits Sold in Major Markets in Awka, Nigeria. Universal Journal of Microbiology Research. 2015;3(2):11-16.
- Abdel-Mallek AY, Hemida SK. Bagy MMK. Studies on fungi associated with tomato fruits and effectiveness of some commercial fungicides against three pathogens. Mycopathologia. 1995;130:109-116.
- Rashad RA, Ahmed RA, Saleh AM. Isolation and identification of some fruit spoilage fungi:screening of plant cell wall degrading enzymes. African Journal of Microbiology. 2011;5(4):443-448.
- Rodriquez P, Soares C, Kozakiewicz Z, Paterson RRM, Lima N, Venancio A. Identification and Characterization of Aspergillus flavus and toxins. Communicating Current Research and Educational topics and trends in Applied Microbiology. 2017;527-534.
- Ugwu CO, Chukwuezi FO, Ozougwu VCO. Microbial Agents of tomato spoilage in Onitsha metropolis. Advances in Biological Research. 2014;8(2):87-93.
- Zain ME. Impact of mycotoxins on humans and animals. Journal of Saudi Chemical Society. 2011;15:129-144.
- Ikuomola DS, Eniola KIT. Assessment of Chemical composition, Microbiological and Sensory qualities of traditional sun-dried pepper (Capsicum annum L) in selected markets in South-western, Nigeria. Nigerian Journal of Microbiology. 2015;27(1):2619-2629.

22. Ismail IA, Nasiru A, Abdullahi Canned Tomato. Biosciences MA, Abdullahi SI. Proximate, Mineral and Vitamin Analysis of Fresh and 13(2):1163-1169.

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