



## Evaluation of Multiple Functions of *Polygonum* Genus Compounds

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### Authors' contributions

This work was carried out in collaboration between all authors. Authors AHLN, LM, HKW, PT and OLF designed and wrote the review. All authors read and approved the final manuscript.

### Article Information

DOI: 10.9734/EJMP/2015/15134

#### Editor(s):

(1) Marcello Iriti, Faculty of Plant Biology and Pathology, Department of Agricultural and Environmental Sciences, Milan State University, Italy.

#### Reviewers:

(1) Gyula Oros, PPI HAS, Budapest, Hungary.

(2) Isiaka A. Ogunwande, Natural Product Research Unit, Department of Chemistry, Faculty of Science, Lagos State University, Ojo, Nigeria.

Complete Peer review History: <http://www.sciencedomain.org/review-history.php?iid=793&id=13&aid=7467>

Review Article

Received 8<sup>th</sup> November 2014  
Accepted 25<sup>th</sup> November 2014  
Published 26<sup>th</sup> December 2014

### ABSTRACT

For thousands of years, traditional medicinal plants have been used to control several diseases, based on traditional knowledge and experience. Nevertheless, many potential medicinal plants have not attracted attention to their useful pharmacological properties and remain to be discovered. In recent years, a number of plants from various genera, species and families have been scientifically studied for their pharmacological potential. Among them, the genus *Polygonum* contains 300 species worldwide. Many document reported various studies of phytochemical and pharmacological potential of crude extracts and compounds isolated from several *Polygonum* species.

**Aims:** The present review describes some traditional uses from the *Polygonum* genus, the phytochemistry, the pharmacological effects, the pharmacokinetics, the toxicology and the known potential phytoconstituents of therapeutic importance that have been isolated.

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**Methodology:** A review of literature was carried out using several resources such as scientific papers, classical books, pubmed, Scifinder, Sirius, the web of Science and ethnobotanical information.

**Results:** Plants from *Polygonum* are widely distributed in the world and used as traditional medicine. Several compounds including phenolic compounds (flavonoids, chalcones, stibenes, coumarins and others) have been isolated and characterized from these plants and some of them are used as the effective pre-clinical to control various diseases in the world.

**Conclusion:** The present review covers many medicinal properties of some species from the *Polygonum* genus, setting out further mechanism of actions and toxicity yet to be established. Studies on *Polygonum* plant extracts could be targeted to develop novel anticancer, anti allergic agents, potential antiplasmodial and anti-inflammatory drugs, from the active compounds. Apart from this, a new approach could be developed for preparing this herbal product, as well as in combination with other plants. Other pharmacological properties for use in arteriosclerosis, neurological disorders, diabetes, hypertension and immunomodulatory effects should be evaluated.

**Keywords:** *Polygonum*; *Polygonaceae*; metabolites; phytoconstituents; pharmacological potential.

## 1. INTRODUCTION

Traditional medicinal plants have long been used in the cure of several human and animal diseases, based on knowledge and experience. Many of these plants have not been studied scientifically and therefore their active principles are still undiscovered. Even though herbal plants in crude form are available for treatment in folk medicine, their uses and activities have unproven track records. In recent years, a number of medicinal plants from various genera, species and families have been scientifically evaluated. Many active principles have been isolated and evaluated for their role in the prevention, control and treatment of many disease conditions.

The *Polygonum* genus contains 300 species found all over the world. They contain diverse pharmacologically active constituents with various properties [1]. These species are predominantly herbs, found in tropical and temperate regions [2]. Some species are important traditional medicines, as illustrated in Fig. 1. Many of these *Polygonum* species and their active principles have been studied for phytochemical screening, biological and pharmacological purposes. Some of them have shown efficacy in the prevention and treatment of diseases such as cancer, malaria, gastric ulcers, bacterial and fungal infections; their toxic effects have also been evaluated.

The present review focuses on some traditional plants from the *Polygonum* genus, their pharmacological and biological effects, and the

known phytochemicals of potential therapeutic importance that have been isolated.

## 2. BOTANY

*Polygonaceae* family contains several genus includes *Polygonum* which is one of most important. *Polygonum* name is from the Greek poly, "many" and gonu, "knee" referring to the swollen-jointed stem. Different species from *Polygonum* genus vary widely from prostrate herbaceous annual plants (4-5 cm high) to herbaceous perennial plants (3–4 m tall). Others are trees or perennial woody vines and grow to around 20–30 m high, or trees. Many species are aquatic and naturally grow in the swampy area as floating plants in the rivers or ponds. The leaves vary in shape between from lanceolate to oval forms. They range from 1 to 30 cm long. The stems are usually red, reddish and sometime red-speckled. Generally, flowers are formed in dense clusters from the leaf joints and the youngest are white, pink or greenish [2].

## 3. TRADITIONAL USES

In China, "Rèlínqīng Kēlì" is the name of *Polygonum* extract. It's used in the folk medicine to cure urinary tract infections [3]. According to the Chinese Pharmacopoeia, *P. cuspidatum* has been used to relieve joint pain, to treat jaundice caused by a bacterial infection or fungal problems and cough with amenorrhoeal expectoration. *Polygonum cuspidatum* is also called "Hu Zhang" in China, and is used as a

decoction to treat liver diseases or as a cream for topical application in burns, wounds and traumatic injuries [4]. It's currently also used in different forms such as powder, decoction or infusion to cure hepatitis and several other diseases [5]. In Korea, the rhizomes of *P. cuspidatum* are commonly used to maintain oral and dental hygiene [6]. In traditional Chinese medicine, *Polygonum multiflorum* has been used to treat mental and physical signs of aging, malaria, constipation and eczema. In Cameroonian traditional medicine, the raffia wine and aqueous extracts of the leaves of *Polygonum limbatum* are respectively used to cure gastrointestinal disorders, venereal diseases and skin infections [7]. In India, preparations of *Polygonum nepalense* are employed in colds, influenza, swelling, hemorrhoids, diarrhea and rheumatism [8]. The juice extract of the whole plant is taken orally for fetal mal-position and the decoction is also taken orally for juvenile pregnancy in Cameroon [9]. In Argentina, *Polygonum ferrugineum* is used to heal infected wounds in local medicine, it's also used to control several bacteria and fungi [10]. In Bangladesh *Polygonum lapathifolium* is used as an insecticide [11]. In Brazil, *Polygonum spectabile* is used by indigenous people for the treatment of diarrhea, ulcers, gingivitis, skin infections and rheumatism [12]. *Polygonum hydropiper* is used as spice to flavor foods by Chinese and Malaysian indigenous because this specie possesses strong peppery taste [13]. In Turkey, the roots of *Polygonum amphibium* are very important in folk medicine and are used as an astringent and cleanser for skin [14,15]; they are also eaten raw, or sometimes they are dried, pounded and the infusion is used in the treatment of chest colds [16]. These data show that the *Polygonum* genus is widely distributed around the world and is being used on all continents to control numerous illnesses.

#### 4. PHYTOCHEMISTRY

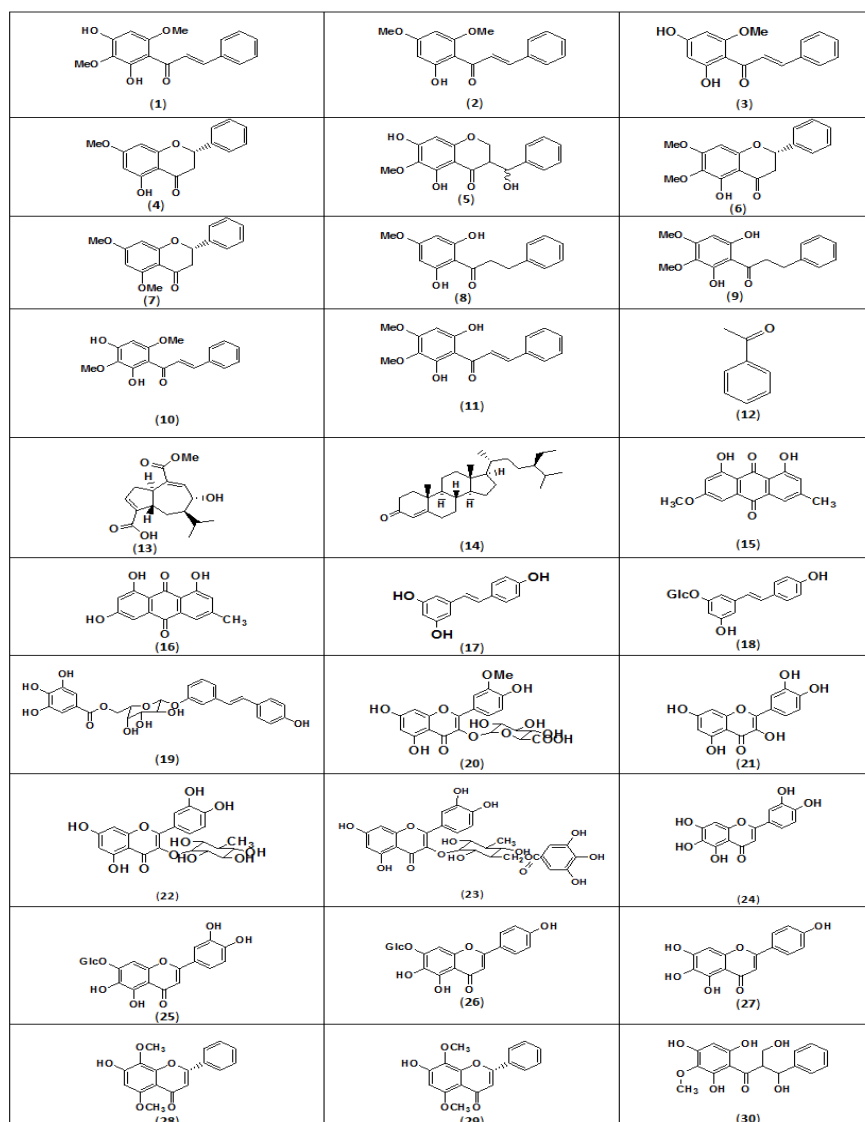
Several investigations were carried out to determine the possible chemical components from *Polygonum* plants. This genus is also well known for producing a wide variety of secondary metabolites including flavonoids [13], triterpenoids [17], anthraquinones [18], coumarins [19], phenylpropanoids [20], lignans [21], sesquiterpenoids [22], stilbenoids [23], tannins [1], proteins, amino acids and carbohydrates [8], and sucrose phenyl propanoid esters [19]. Amongst them, flavonoids are the most common components found in *Polygonum*

spp. and have been used as chemotaxonomic markers of the genus [22]. Fig. 1. presents some bioactive compounds obtained from the *Polygonum* genus and Table 1. shows the pharmacological effects of some extracts and compounds isolated from some *Polygonum* plants. In summary, the present review focuses on studies of a number of traditional medicinal plants from the *Polygonum* genus, their pharmacological and biological effects, and the known potential phytoconstituents of therapeutic importance isolated so far.

### 5. PHARMACOLOGICAL EFFECTS

#### 5.1 Anticancer Effects

Recently, Hu and coworkers studied the activities of aqueous *Polygonum cuspidatum* extract on hepatocarcinoma cells (Bel-7402 and Hepa 1-6) in suspension [4]. In this study, the sample (extract) significantly ( $P < 0.05$ ) inhibited the proliferation of these cells in suspension in a dose-dependent and time-dependent manner. This extract also respectively showed the significant inhibition and induction of hepatocarcinoma cells in soft agar and anoikis in human Bel -7402 cells. Previously, crude extracts and compounds such as emodin and resveratrol showed anticancer activities [24,25]. The growth of cancer cells specially Ehrlich's carcinoma was inhibited by aqueous extracts (20 g/kg/day) [25], alternatively the ethanol extract showed an antiproliferative activity against two different cancer cells (human Lung H1650 and A549), in a dose-dependent manner [26]. Resveratrol reduced by using 2.5 and 10 mg.kg<sup>-1</sup> for 5 days the volume and the weight of Lewis Lung tumor respectively (42%) and (44%). The tumor growth and metastasis in lungs are prevented by resveratrol (56%) [27]. Using resveratrol, several significant anticancer activities have been obtained by different research team. In an experiment on rats with intracerebral gliomas, the molecule (40 mg/kg/day, 150 days) exhibited anticancer activity such as higher and longer rat survival time and slower tumor growth [24]. Resveratrol in other experiments also showed cytotoxicity activities against lymph cancer [28], hepatic cancer [29], MCF-7 cells with IC<sub>50</sub> (58.4 µg.mL<sup>-1</sup>) and adriamycin-resistant MCF-7 cells with IC<sub>50</sub> (56.7 µg.mL<sup>-1</sup>) [30], ovarian cancer [31], human neuroblastomas and uveal melanoma tumors [32], atypical teratoid or rhabdoid tumors [33].



**Fig. 1.** Some bioactive compounds obtained from the *Polygonum* genus. (1) 2',4'-dihydroxy-3',6'-dimethoxychalcone; (2) 2'-hydroxy-4',6'-dimethoxychalcone; (3) cardamomin; (4) (S)-(-)-pinostrobin; (5) (±)-polygohomoisoflavanone; (6) (2S)-(-)-5-hydroxy-6,7-dimethoxyflavanone; (7) (2S)-(-)-5,7-dimethoxyflavanone; (8) 2',6'-dihydroxy-4'-methoxydihydrochalcone; (9) 2',6'-dihydroxy-3',4'-dimethoxydihydrochalcone; (10) 2',4'-dihydroxy-3',6'-dimethoxychalcone; (11) 2',6'-dihydroxy-3',4'-dimethoxychalcone; (12) acetophenone; (13) viscozulenenic acid; (14) sitosterone; (15) physcion; (16) emodin; (17) resveratrol; (18) polydatin; (19) resveratrol 4-O-D-(2'-galloyl)-glucopyranoside; (20) quercetin 3-O-β-D-glucuronide; (21) quercetin; (22) quercitrin; (23) galloyl kaempferol 3-glucoside; (24) kaempferol 3-glucoside; (25) 6-Hydroxyluteolin; (26) 6-Hydroxyluteolin 7-O'-D-glucopyranoside; (27) 6-Hydroxyapigenin; (28) scutillarein; (29) 5,8-dimethoxy-7-hydroxyflavanone; (30) homoferrugin-dihydrochalcone

Emodin also exerted cytotoxic effects against human prostate cancer cell LNCaP, glioma cells [33] and human chronic myelocytic leukemia K562 cells [34]. The anticancer activities of the crude extract, fractions and flavonoids isolated

from *Polygonum limbatum* were studied [7]. In experiment, the cytotoxicity of samples studied on several cancer cells such as breast carcinoma MCF-7, Leukemia THP-1, prostate carcinoma PC-3, Lung A549, cervical carcinoma HeLa. The

results showed that, more than 50% inhibition of the proliferation was observed on above cell lines except Lung A549. Cardamomin and 2',4'-dihydroxy-3',6'-dimethoxychalcone are very active against THP-1 cell with  $IC_{50}$  below  $4 \mu\text{g}\cdot\text{mL}^{-1}$  [7]. Furthermore, the cytotoxicity of crude extracts and compounds from *Polygonum spectabile* were also evaluated [22,35]. The cytotoxicity of crude extracts and compounds such as 2'-hydroxy-4',6'-dimethoxychalcone, 2',4'-dihydroxy-3',6'-dimethoxychalcone and 3-O- $\beta$ -D-glucosyl- $\beta$ -sitosterol ( $500\text{-}0.125 \mu\text{g}\cdot\text{mL}^{-1}$ ) to Vero and LLCMK2 cells were evaluated by using MTT assay. Each test was carried out in four replicates with at least four different concentrations. The results revealed that the three compounds isolated from *P. spectabile* showed moderate *in vitro* cytotoxicity activity against Vero and LLCMK2 cells with  $CC_{50} < 50 \mu\text{g}\cdot\text{mL}^{-1}$  [35].

## 5.2 Antioxidant and Free Radical Scavenging Effects

The antioxidant effect of *Polygonum chinense* extract was evaluated. In this experiment, the preventive effects of aqueous extract against gastric mucosal injury in rats were determined [36]. The result of study showed that this extract significantly protects gastric mucosal [36]. The anti-ulcer activity of aqueous extract of *Polygonum minus* against gastric ulcer was also investigated. The results revealed that the plant extract induced a marked reduction in edema and conferred gastric protection [37]. Moreover, the toxicity of *P. minus* was studied. The higher dose ( $5 \text{ g}\cdot\text{kg}^{-1}$ ) did not show any toxicological sign in rats [37]. The antioxidant effects of some *Polygonum* species have been examined by Hsu and coworkers. In this experiment, the ethanolic extract of the root of *P. cuspidatum* was used as sample and evaluated the antioxidant activities, free radical scavenging, superoxide radical scavenging, lipid peroxidation assays have been used and  $IC_{50}$  values have been respectively obtained;  $110 \text{ mg}\cdot\text{g}^{-1}$ ,  $3.2 \text{ mg}\cdot\text{g}^{-1}$  and  $8 \text{ mg}\cdot\text{g}^{-1}$ . Furthermore, the extract of this plant had a DNA-protective activity. The results indicated that *P. cuspidatum* extract had antioxidant activities [38]. In another experiment in 1993, in fact, Jin and coworkers evaluated the antioxidant activity of polydatin ( $3.2, 6.4, 12.8$  and  $25.6 \mu\text{mol}\cdot\text{l}^{-1}$ ), a molecule isolated from *P. cuspidatum* extract. Polydatin also showed a strong potential to scavenge oxygen free radicals with  $IC_{50} = 14.6 \mu\text{mol}\cdot\text{L}^{-1}$  for superoxide radicals ( $O_2^{\cdot-}$ ), hydroxyl

radicals ( $OH^{\cdot}$ ) ( $IC_{50} = 29.6 \mu\text{mol}\cdot\text{l}^{-1}$ ) and hydrogen peroxide ( $H_2O_2$ ) ( $IC_{50} = 13.0 \mu\text{mol}\cdot\text{L}^{-1}$ ) [39].

The *in vitro* antioxidant activity of *P. barbatum* extract was investigated by different strategies including hydroxyl radical scavenging assay, DPPH radical scavenging assay, superoxide scavenging assay, nitric oxide scavenging assay and total phenolic content. The  $IC_{50}$  ( $32.62 \mu\text{g}\cdot\text{ml}^{-1}$ ) value found in the DPPH radical scavenging method was higher than with other methods. *P. barbatum* extract contained phenolic compounds, which showed significant antioxidant effects [39]. The *in vitro* studies clearly showed that the ethanolic extract of *P. barbatum* is a new source of natural antioxidants that might prevent and control the process of oxidative stress [39].

## 5.3 Anti-inflammatory Effects

In 2008, one American research team studied the anti-inflammatory effects of *P. cuspidatum*. In this experiment, the plant material was extract with ethanolic solution and use as sample. The study evaluated the ability of sample to inhibit animal (mouse) ear inflammation. Sample was applied to both ears of animals at different doses  $0.075, 0.15, 0.3, 1.25$  and  $2.5 \text{ mg}\cdot\text{ear}^{-1}$  30 min after topical application of 12-O-tetradecanoylphorbol-13-acetate administration ( $2 \mu\text{g}\cdot\text{ear}^{-1}$ ). Comparing to the topical application of 12-O-tetradecanoylphorbol-13-acetate as control, sample treatment at different doses significantly decreased ear edema. Myeloperoxidase effect was inhibited at *P. cuspidatum* extract doses  $\geq 1.25 \text{ mg}\cdot\text{Kg}^{-1}$ . Trans-resveratrol also inhibit inflammation at different comparables doses. The development edema is inhibited by the extract of *P. cuspidatum* [40].

## 5.4 Wound Healing Effects

After wound treatment, a scar often forms, yielding an unpleasant spot. It is therefore necessary to erase the scar to regain the original luster. Based on multiple activities of plants of the genus *Polygonum*, the wound healing activity of *P. barbatum* was evaluated. Ethanol and water were used as solvent to extract the whole plant [41]. Excision and incision were two wound models used to study this effect. The extracts responded significantly ( $P < 0.001$ ) in both the wound models tested. Between the two extracts, the aqueous extract was more active than the ethanol extract [41].

### 5.5 Anti Allergic Effects

The anti allergic activity of *P. cuspidatum* radix extract was investigated by Lim and co-workers [42]. In this experiment, the extract was tested against two different mast cells and the sample showed very potent inhibitory effect with IC<sub>50</sub> values (62±2.1) µg.ml<sup>-1</sup> for RBL-2H3 and (46±3.2) µg.mL<sup>-1</sup> for bone marrow-derived mast cells by antigen stimulation. The extract also exhibited the ability to suppress the expression of interleukin-4 in RBL-2H3 cells and tumor necrosis factor-α. Concerning the *in vivo* animal allergy model, in dose dependent manner, *P.cuspidatum* radix inhibited passive cutaneous anaphylaxis, a local allergic reaction. The same sample also showed significant inhibition to activate phosphorylation of Syk and expressed the ability to suppress the mitogen-activated protein kinases ERK1/2 [42].

### 5.6 Anti Molluscidal and Antiplasmodial Effects

The anti molluscidal activity of crude aqueous and methanol extracts and compounds from *Polygonum senegalense* was evaluated [43] and the samples showed significant molluscidal effect against two different strains snails (*Biomphalaria pfeifferi* and *Biomphalaria sudanica*). One of compounds namely 2',4'-dihydroxy-3',6'-dimethoxychalcone was very active with 100% lethal to both snails in less than six hours. Two other fractions (hydrophobic and hydrophilic) of methanol extract of this plant also showed molluscidal activity on these snails [43]. Furthermore, the *in vitro* antiplasmodial activity of extract and compounds (polyphenol isoflavanone (19.2±0.6) µM for D6 and (18.2±1.8 µM), 2'- hydroxy- 4',6'- dimethoxychalcone (15.7±1.2 µM) for D6 and (11.3±0.2 µM) for W2 and 2',6'- dihydroxy- 3',4'- dimethoxychalcone (3.1±0.8 µM) for D6 and (2.4±0.3 µM) from *P. senegalense* was evaluated against two strains of *Plasmodium falciparum* (D6 and W2). The result showed that compounds isolated from *P. senegalense* are significantly active with IC<sub>50</sub> between (19.2 to 2.4 µM) against two strains (D6 and W2) malaria parasite [44].

### 5.7 Antibacterial and Antifungal Effects

Several experiments have documented the antibacterial and antifungal potential of extracts and compounds from the genus *Polygonum*. The extract and compounds isolated from *Polygonum*

*spectabile* were also evaluated for their effects on 15 bacteria. Hydrophilic extracts of *P. spectabile* were active against *Staphylococcus aureus*, *Bacillus subtilis*, *Micrococcus luteus*, *Micrococcus canis*, *Trichophyton mentagrophytes* and *Trichophyton rubrum*. The results indicated that the use of *P. spectabile* may act as an antibacterial and antifungal agent [35]. Another antimicrobial screening of ethanol leaf extract of *P. barbatum* was assessed on ten microorganisms which include four Gram-positive, four Gram-negative bacteria and two fungi. The ethanol leaf extract showed considerable activity on both the bacteria and fungi, with inhibition zone diameter ranging from 14.17 to 31.45 mm and the minimum inhibitory concentrations ranging from 12 to 16 µg.mL<sup>-1</sup>. These results showed that the ethanol extract of this plant is a broad-spectrum antimicrobial agent which can be used against Gram-positive and -negative bacteria and also against fungi [39].

The antimicrobial activities of *P. cuspidatum* were also investigated [45]. In this experiment, the aqueous extract of *P. cuspidatum* (1.0 g.mL<sup>-1</sup>) showed antibacterial activity against *Staphylococcus aureus*, *Staphylococcus albus*, *Escherichia coli*, *α-Streptococcus*, *β-Streptococcus*, *Bacterium typhosum* and *Pseudomonas aeruginosa*; the inhibition zones were 1.122, 1.113, 1.127, 0.800, 0.929, 0.903, 1.112 cm, respectively [46]. The methanol extracts of this plant also exhibited antibacterial activities against *Streptococcus sobrinus* and *Streptococcus mutans* at the minimum inhibitor concentration (MIC) (0.5–4 mg.mL<sup>-1</sup>). This result suggested that the methanol extract could be used to control dental plaque and dental caries formation [6,47]. In another experiment, the crude extract of *P. cuspidatum* was tested against five common foodborne bacteria. The sample exhibited antibacterial activities against *Bacillus cereus*, *S. aureus* and *Listeria monocytogenes* at the minimum inhibitor concentrations of 312.5, 312.5 and 156.3 µg.mL<sup>-1</sup> respectively, and the bactericidal minimum inhibitor concentrations were 625, 1250 and 312.2 µg.mL<sup>-1</sup> [48]. In 2010, the antibacterial activities of methanol extract and fractions of *P. cuspidatum* on the development of dental caries were re-investigated. The results of the experiment showed that the ethyl acetate fraction, constituted mainly of polydatin, resveratrol, emodin and anthraglycoside B, exhibited significant antibacterial activities against eight different strains of *Streptococcus*, including *Streptococcus sobrinus* KCTC 3388, *S.*

*mutans* from different strains (KCTC 3308, KCTC 3307, KCTC 3389, KCTC 3306, KCTC 3300, KCTC 3298) and *Streptococcus cricetus* KCTC 3292, with minimum inhibitor concentrations of 0.5, 1, 0.5, 1, 0.5, 0.5, 0.125 and 0.25 mg.mL<sup>-1</sup> respectively, and minimum bactericidal concentrations of 2, 4, 2, 2, 2, 1, 2, 0.5 mg.mL<sup>-1</sup> respectively [49].

### 5.8 Genotoxicity Effects

Genotoxicity is a destructive effect on a cell's genetic material (DNA, RNA) affecting its integrity. *Polygonum multiflorum* has been designed for its potential to control genotoxicity. In this context, Zhang and coworkers studied genotoxicity in mouse peripheral lymphocyte cells with the use of *Panax ginseng* and *P. multiflorum* and the activity of their combination [50]. They administered them individually or in combination, on mouse peripheral lymphocyte DNA. *P. multiflorum* and its combination with *Panax ginseng* were orally administered to mice at gradient doses (low, medium and high) for seven consecutive days. On the first, third and seventh days, two hours after drug administration, blood samples were collected from the vein cluster behind the eye. *Panax ginseng* (0.43, 1.3 and 3.9 g.kg<sup>-1</sup>) was found to have no harmful effects on peripheral lymphocyte DNA. *P. multiflorum* (3.9 and 11.7 g.kg<sup>-1</sup>), on the other hand, had harmful effects on peripheral lymphocyte DNA on the first, third and seventh days. The combination of both plants (5.2 and 15.6 g.kg<sup>-1</sup>) induced harmful effects on peripheral lymphocyte DNA on the first day, as showed in tail DNA tail length. Nevertheless, these harmful effects decreased on the third and seventh days. The results showed that the combination of these two plants may be used to control potential genotoxic effects and this activity is induced by *P. multiflorum* [50].

### 5.9 Hepatotoxicity Effects

Several experiments investigated the *in vitro* and *in vivo* hepatotoxicity of hot-water-extracts of *P. multiflorum* in mice [51]. After treating primary cultured hepatocytes with *P. multiflorum* extracts at three final concentrations (0.5, 1.0 and 5.0 mg.mL<sup>-1</sup>) with or without acetaminophen (10 mM) for 24 h, no cytotoxic effects was observed. Cell viability and lactate dehydrogenase leakage were significantly improved at the highest *P. multiflorum* extract dose compared with controls. Mice that received the *P. multiflorum* extracts (20 or 340 mg extracts/mouse x 2) orally

administered twice daily for 10 days indicated no unfavorable effect on the liver function. When the mice sub-chronically received the higher dose of *P. multiflorum* extract (340 mg) before the treatment with a single-bolus dose of acetaminophen (500 mg.kg<sup>-1</sup>), attenuation of acetaminophen-induced hepatotoxicity was significantly *in vivo* established [51]. The results showed that *P. multiflorum* does not induce any toxicological effect on the liver and may in fact elicit useful but limited beneficial effects on the liver *in vivo* [51].

### 5.10 Antiviral Effects

An antiviral activity, an important and specific effect of *P. cuspidatum* was extensively investigated. Several experiments have shown that crude extracts or compounds isolated from *P. cuspidatum* have ability to control HIV [26,52,53]. In 1998, Jiang and coworkers evaluated the antiviral effect of water extract of this plant (50 mg/mouse/day, for 1 month); the sample exhibited an antiviral activity in an HIV-infected murine model [52]. Several compounds such as resveratrol, catechin, 5,7-dimethoxyphthalide and emodin 8-O-β-D-glycopyranoside isolated from *P. cuspidatum* showed significant antiviral activity against HIV-1; EC<sub>50</sub> values were 4.37±1.96 µg.ml<sup>-1</sup>, 14.4±1.34 µg.ml<sup>-1</sup>, 19.97±5.09 µg.ml<sup>-1</sup> and 11.29±6.26 µg.ml<sup>-1</sup> respectively [26]. In addition, resveratrol was also considered as a new molecule for anti-HIV therapeutics [53]. In fact, resveratrol pretreatment dose-dependently increased sirtuins 1 protein expression and intracellular NAD<sup>+</sup> level. This compound and the extract of *P. cuspidatum* were used against Epstein-Barr virus (EBV) [54], hepatitis B virus (HPV) [55].

## 6. PHARMACOKINETICS

Plants from the *Polygonum* genus have many pharmacological effects, including anticancer, antioxidant and free radical scavenging, anti-inflammatory, wound healing, anti allergic, anti molluscidal, antiplasmodial, antibacterial, antifungal, genotoxic, hepatotoxic and antiviral effects. From these biological activities it can be noted that several extracts and pure compounds isolated from *Polygonum* genus plants have a promising future to control, prevent various pathogenics. The preliminary results suggest the viability of future investigations, such as studies involving the mechanisms of action, pharmacokinetics and pharmacodynamics of these pure compounds.

Table 1. Pharmacological effects of some extracts and compounds isolated from some *Polygonum*

Plants	Extracts. compounds	Pharmacological effects	Minimal active concentration.dose	In vivo. in vitro	Reference
<i>Polygonum spectabile</i>	Hexane extract	<i>Anti-Tricophyton mentagrophytes</i>	18.5±0.8 mg.disc <sup>-1</sup>	<i>In vitro</i>	[35]
		<i>Anti-Tricophyton rubrum</i>	10.5±1.1 mg.disc <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-Micrococcus canis</i>	9.5±1.4 mg.disc <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-Vero cells</i>	83.6±4.6 µg.ml <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti- LLCMK<sub>2</sub>Cells</i>	71.3±3.5µg.ml <sup>-1</sup>	<i>In vitro</i>	
	Dichloromethane extract	<i>Anti-Tricophyton mentagrophytes</i>	11.5±0.8 mg.disc <sup>-1</sup>	<i>In vitro</i>	[35]
		<i>Anti-Tricophyton rubrum</i>	10.5±0.5 mg.disc <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-Micrococcus canis</i>	8.5±0.9mg.disc <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-Vero cells</i>	128.2±6.9µg.ml <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti- LLCMK<sub>2</sub>Cells</i>	117.9±7.2µg.ml <sup>-1</sup>	<i>In vitro</i>	
	Ethyl acetate extract	<i>Anti-Staphylococcus aureus</i>	8.8±0.8 mg.disc <sup>-1</sup>	<i>In vitro</i>	[35]
		<i>Anti-Tricophyton mentagrophytes</i>	12.0±0.9 mg.disc <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-Tricophyton rubrum</i>	9.5±0.6 µg.ml <sup>-1</sup>	<i>In vitro</i>	
	Ethanol extract	<i>Anti-Staphylococcus aureus</i>	7.5±0.5 µg.ml <sup>-1</sup>	<i>In vitro</i>	[35]
		<i>Anti-Micrococcus luteus</i>	9.5±0.5 µg.ml <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-HHV-1<sup>a</sup></i>	21.9±1.8µg.ml <sup>-1</sup>	<i>In vitro</i>	
	2'-hydroxy-4',6'-dimethoxychalcone	<i>Anti-Staphylococcus aureus</i>	15.2±0.9 mm	<i>In vitro</i>	[35]
		<i>Anti-Staphylococcus epidermides</i>	11.3±0.5 mm	<i>In vitro</i>	
		<i>Anti-Bacillus subtilus</i>	12.7±0.8 mm	<i>In vitro</i>	
<i>Anti-Micrococcus luteus</i>		11.3±0.5 mm	<i>In vitro</i>		
<i>Anti-Micrococcus canis</i>		14.6 mm	<i>In vitro</i>		
<i>Anti-Tricophyton mentagrophytes</i>		24.7 mm	<i>In vitro</i>		
<i>Anti-Tricophyton rubrum</i>		24.3 mm	<i>In vitro</i>		
2',4'-dihydroxy-3',6'-dimethoxychalcone	<i>Anti-Vero cells</i>	31.5 µg.mL <sup>-1</sup>	<i>In vitro</i>	[35]	
	<i>Anti- LLCMK<sub>2</sub>Cells</i>	29.6 µg.mL <sup>-1</sup>	<i>In vitro</i>		
	<i>Anti-Vero cells</i>	27.2 µg.mL <sup>-1</sup>	<i>In vitro</i>		
3-O-β-D-glucosyl-β-sitosterol	<i>Anti- LLCMK<sub>2</sub>Cells</i>	7.8 µg.mL <sup>-1</sup>	<i>In vitro</i>	[35]	
<i>Polygonum limbatum</i>	cardamomin	<i>Anti-THP-1 (Leukemia)</i>	1.8 µg.mL <sup>-1</sup>	<i>In vitro</i>	[7]
		<i>Anti-Hela (cervix)</i>	17 µg.mL <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-PC-3 (prostate)</i>	49 µg.mL <sup>-1</sup>	<i>In vitro</i>	
		<i>Anti-MCF-7 (breast)</i>	32 µg.mL <sup>-1</sup>	<i>In vitro</i>	



Plants	Extracts. compounds	Pharmacological effects	Minimal active concentration.dose	In vivo. in vitro	Reference
	(±) Polygohomoisoflavanone	Anti-THP-1 (Leukemia)	32.5 µg.mL <sup>-1</sup>	In vitro	[7]
		Anti-PC-3 (prostate)	40 µg.mL <sup>-1</sup>	In vitro	[7]
		Anti-MCF-7 (breast)	36 µg.mL <sup>-1</sup>	In vitro	[7]
	2',4'-dihydroxy-3',6'-dimethoxychalcone	Anti-THP-1 (Leukemia)	3.5µg.mL <sup>-1</sup>	In vitro	[7]
		Anti-Hela (cervix)	22 µg.mL <sup>-1</sup>	In vitro	[7]
	(S)(-)-pinostrobin	Anti-THP-1 (Leukemia)	9 µg.mL <sup>-1</sup>	In vitro	[7]
		Anti-PC-3 (prostate)	40 µg.ml <sup>-1</sup>	In vitro	[7]
		Anti-MCF-7 (breast)	36 µg.mL <sup>-1</sup>	In vitro	[7]
	(2S)(-)-5-hydroxy-6,7-dimethoxyflavanone	Anti-THP-1 (Leukemia)	25.5 µg.m <sup>-1</sup> L	In vitro	[7]
		Anti-MCF-7 (breast)	47 µg.mL <sup>-1</sup>	In vitro	[7]
	(2S)(-)-5,7-dimethoxyflavanone	Anti-THP-1 (Leukemia)	10 µg.mL <sup>-1</sup>	In vitro	[7]
		Anti-MCF-7 (breast)	37 µg.mL <sup>-1</sup>	In vitro	[7]
	Methanol extract	Anti-THP-1 (Leukemia)	10 µg.mL <sup>-1</sup>	In vitro	[7]
		Anti-PC-3 (prostate)	28 µg.mL <sup>-1</sup>	In vitro	[7]
		Anti-MCF-7 (breast)	20 µg.mL <sup>-1</sup>	In vitro	[7]
n-Butanol extract	Anti-THP-1 (Leukemia)	9 µg.mL <sup>-1</sup>	In vitro	[7]	
	Anti-PC-3 (prostate)	9 µg.mL <sup>-1</sup>	In vitro	[7]	
	Anti-MCF-7 (breast)	18 µg.mL <sup>-1</sup>	In vitro	[7]	
Ethyl acetate extract	Anti-THP-1 (Leukemia)	8.5 µg.mL <sup>-1</sup>	In vitro	[7]	
	Anti-MCF-7 (breast)	23 µg.mL <sup>-1</sup>	In vitro	[7]	
<i>Polygonum cuspidatum</i>	Ethanol extract	Anti-inflammatory(reduced ear edema induced by TPA)	0.075 mg.ear <sup>-1</sup>	In vivo	[40]
		Anti-inflammatory(reduced myeloperoxidase activity)	2.5 mg.ear <sup>-1</sup>	In vivo	[40]
	polydatin	Inhibits the vasoconstrictive effect	1.71 mmol.L <sup>-1</sup>	In vivo	[60]
		Reduce lipogenesis of rats	50 mg.kg <sup>-1</sup>	In vivo	[61]
		Enhanced MCF-7 proliferation	20 µmol.L <sup>-1</sup>	In vitro	[62]
	physcion	Inhibition of melanogenesis effect	10 µmol.L <sup>-1</sup>	In vitro	[63]
	emodin	Inhibition of melanogenesis effect	10 µmol.L <sup>-1</sup>	In vitro	[63]
		Anti-HSV-1	4 mg.mL <sup>-1</sup>	In vivo	[64]
		Anti-human chronic myelocytic leukemia K562 cells	25 µmol.L <sup>-1</sup>	In vitro	[34]
		Anti-human prostate cancer cells	10 µmol.L <sup>-1</sup>	In vitro	[65]

Plants	Extracts. compounds	Pharmacological effects	Minimal active concentration.dose	In vivo. in vitro	Reference
		<i>Anti-glioma cells</i>	100 $\mu\text{mol.L}^{-1}$	<i>In vitro</i>	[66]
	cytreosein	<i>Inhibition of melanogenesis effect</i>	10 $\mu\text{mol.L}^{-1}$	<i>In vitro</i>	[63]
	anthraglycoside	<i>Inhibition of melanogenesis effect</i>	10 $\mu\text{mol.L}^{-1}$	<i>In vitro</i>	[63]
	Water extract	<i>Anti-Escherichia coli</i>	1 $\text{g.mL}^{-1}$	<i>In vitro</i>	[46]
		<i>Anti-Bacterium typhosum</i>	1 $\text{g.mL}^{-1}$ ,	<i>In vitro</i>	[46]
		<i>Anti-pseudomonas aeruginosa</i>	1 $\text{g.mL}^{-1}$ ,	<i>In vitro</i>	[46]
		<i>Anti-Staphylococcus aureus</i>	1 $\text{g.mL}^{-1}$ ,	<i>In vitro</i>	[46]
		<i>Anti-Staphylococcus albus</i>	1 $\text{g.mL}^{-1}$ ,	<i>In vitro</i>	[46]
		<i>Anti-<math>\alpha</math>-Streptococcus aureus</i>	1 $\text{g.mL}^{-1}$	<i>In vitro</i>	[46]
		<i>Anti-<math>\beta</math>-Streptococcus</i>	1 $\text{g.mL}^{-1}$	<i>In vitro</i>	[46]
	resveratrol	<i>Anti-tumor</i>	40 $\text{mg.kg}^{-1}$	<i>In vivo</i>	[24]
		<i>Anti-uveal melanoma tumors</i>	20 $\text{mg.tumor.times}^{-1}$	<i>In vivo</i>	[32]
		<i>Anti-human neuroblastomas</i>	5 $\text{mg.tumor.times}^{-1}$	<i>In vivo</i>	[32]
		<i>Anti-lymph cancer</i>	100 $\mu\text{M}$	<i>In vitro</i>	[28]
		<i>Anti-atypical teratoid tumors</i>	150 $\mu\text{M}$	<i>In vitro</i>	[33]
	resveratrol	<i>Anti-hepatic cancer</i>	12.5 $\mu\text{M}$	<i>In vitro</i>	[38]
	resveratrol	<i>Anti-EBV</i>	13.8 $\mu\text{M}$	<i>In vitro</i>	[54]
<i>Polygonum senegalense</i>	2',6'-dihydroxy-4'-methoxydihydrochalcone	<i>Anti-D6 plasmodium falciparum</i>	23.5 $\mu\text{M}$	<i>In vitro</i>	[44]
		<i>Anti-W2 plasmodium falciparum</i>	22.8 $\mu\text{M}$	<i>In vitro</i>	[44]
	2',6'-dihydroxy-3',4'-dimethoxydihydrochalcone	<i>Anti-D6 plasmodium falciparum</i>	11.8 $\mu\text{M}$	<i>In vitro</i>	[44]
		<i>Anti-W2 plasmodium falciparum</i>	12. $\mu\text{M}$	<i>In vitro</i>	[44]
	2',4'-dihydroxy-3',6'-dimethoxychalcone	<i>Anti-W2 plasmodium falciparum</i>	17.8 $\mu\text{M}$	<i>In vitro</i>	[44]
		<i>Anti-D6 plasmodium falciparum</i>	16.6 $\mu\text{M}$	<i>In vitro</i>	[44]
<i>Polygonum Barbatum</i>	Acetophenone	<i>Anti-DPPH</i>	1.8 x 10 <sup>-1</sup> $\text{mg.mL}^{-1}$	<i>in vitro</i>	[67]
	Viscozulenic acid	<i>Anti-DPPH</i>	1.2X10 <sup>-1</sup> $\text{mg.mL}^{-1}$	<i>In vitro</i>	[67]
	Sitosterone	<i>Anti-DPPH</i>	2.1X10 <sup>-1</sup> $\text{mg.mL}^{-1}$	<i>In vitro</i>	[67]
<i>Polygonum hydropiper</i>	Quercetin 3-O- $\beta$ -D-glucuronide	<i>Antioxidant</i>	5.08 (TEAC value)	<i>In vitro</i>	[13]
	Quercetin	<i>Antioxidant</i>	4.65(TEAC value)	<i>In vitro</i>	[13]
	Quercitrin	<i>Antioxidant</i>	3.46 (TEAC value)	<i>In vitro</i>	[13]
	Galloyl kaempferol 3-glucoside	<i>Antioxidant</i>	2.90 (TEAC value) <sup>1</sup>	<i>In vitro</i>	[13]
	Kaempferol 3-glucoside	<i>Antioxidant</i>	1.39 (TEAC value)	<i>In vitro</i>	[13]
	6-Hydroxyluteolin	<i>Antioxidant</i>	2.33 (TEAC value)	<i>In vitro</i>	[13]
	6-Hydroxyluteolin 7-O'-D-	<i>Antioxidant</i>	2.87 (TEAC value)	<i>In vitro</i>	[13]

Plants	Extracts. compounds	Pharmacological effects	Minimal active concentration.dose	In vivo. in vitro	Reference
<i>Polygonum ferrugineum</i>	glucopyranoside				
	5.8-dimethoxy-7-hydroxyflavanone	<i>Anti-Trichophyton mentagrophytes</i> ATCC9972	250 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]
		<i>Anti-Trichophyton rubrum</i> CCC113	250 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]
		<i>Anti-Epidermophyton floccosum</i> CCC114	250 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]
	Homoferrugen-dihydrochalcone	<i>Anti-Trichophyton mentagrophytes</i> ATCC9972	200 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]
		<i>Anti-Trichophyton rubrum</i> CCC113	250 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]
	Methanol extract	<i>Microsporum gypseum</i> CCC 115	125 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]
		<i>Anti-Trichophyton rubrum</i> CCC113	50 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]
<i>Anti-Trichophyton mentagrophytes</i> ATCC9972		125 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]	
<i>Anti-Epidermophyton floccosum</i> CCC114		50 µg.mL <sup>-1</sup>	<i>In vitro</i>	[68]	

Furthermore, a few pharmacokinetic investigations have been done on *Polygonum* plants. In 2008, one pharmacokinetic experiment with rats received the extract of *P. cuspidatum*. After this experiment, methanol was used as polar organic solvent to extract tissues and solid-phase extraction was used to clean urinary and biliary samples. The secondary metabolites were analyzed and identified in mass spectrometry. The major of the chemical constituents of this extract was resveratrol, which was distributed in the liver, kidney, duodenum and stomach [56]. With this result, other scientists then evaluated different active compounds of *P. cuspidatum*. In particular, emodin was investigated in rats using selective HPLC method. The result of this experiment clearly showed that emodin is quickly absorbed into the blood stream and rapidly transferred into the liver [57]. In 2012, the results of experiments using a HPLC method showed that after oral administration of *Polygonum cuspidatum* extract, glucuronides or sulfates of resveratrol and emodin were the best forms in distribution and organs [58,59].

## 7. TOXICOLOGY

Plants from the *Polygonum* genus have long been used as an important medicinal plant in the world. Some of these plants are traditionally considered to be toxic, such as *P. cuspidatum*, which according to Chinese folk medicine, the plant induces abortion and for this reason, it's prohibited for pregnant women [5]. The aqueous extract of *Polygonum minus* was administered orally in rats with 5 g.kg<sup>-1</sup> (higher dose), the acute toxicity was studied and animals model did not present toxicological signs [37]. It was also reported that LD<sub>50</sub> of emodin (249.5±34.3 mg.kg<sup>-1</sup>) and polydatin (1000±57.3 mg.kg<sup>-1</sup>) were (249.5±34.3 mg.kg<sup>-1</sup>) administered orally did not cause death in mice. In one acute toxicity study, rats were treated with the extract of *P. chinense* at a dose of 2 or 5 g.kg<sup>-1</sup>. Despite the bad health of rats, no signs of toxicity have been observed at these doses. No abnormalities have been observed after biochemical serology and histological indicators of liver and kidney analysis [36].

## 8. CONCLUSION AND PROSPECTS

The present review covers many medicinal properties of some plants from the *Polygonum* genus, with further mechanism of actions and toxicity yet to be established. Studies on

*Polygonum* plant extracts could be targeted to develop novel anticancer and anti allergic agents and potential antiplasmodial and anti-inflammatory drugs, from the active compounds [69]. These active principles could be clinically evaluated to develop therapeutic drugs with more safety, efficacy and tolerability. Apart from this, a new approach could be developed for preparing this herbal product, as well as in combination with other plants. Other pharmacological properties for use in atherosclerosis, neurological disorders, diabetes, hypertension and immunomodulatory effects should be evaluated. Moreover, in agriculture, the anti-nematode and anti-insecticidal activities of *Polygonum* plants extracts against potato pathogens will also be evaluated. In summary, data reviewed here clearly show the enormous potential of *Polygonum* plants for the development of new pharmaceuticals, adding this genus to the list of highly promising native compounds to undergo further research.

## CONSENT

Not applicable.

## ETHICAL APPROVAL

Not applicable.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Wang KJ, Zhang YJ, Yang CR. Antioxidant phenolic compounds from rhizomes of *Polygonum paleaceum*. Journal of Ethnopharmacology. 2005;96(3):483-487.
2. Hutchinson J, Dalziel JM. Flora of west tropical Africa revised by Keay RWJ. Whitefriars, London. 2<sup>nd</sup> edition. Crown Agents. London; 1954.
3. Luczaj L. Archival data on wild food plants used in Poland in 1948. Journal of Ethnobiology and Ethnomedicine. 2008;4:4.
4. Hu B, An HM, Shen KP, Song HY, Deng S. *Polygonum cuspidatum* extract induces anoikis in hepatocarcinoma cells associated with generation of reactive oxygen species and downregulation of focal adhesion kinase. Evidence-Based

- Complementary and Alternative Medicine, Ecam; 2012. DOI: 1155/2012/607675.
5. State administration of traditional Chinese medicine. 1999;1(1).
  6. Song JH, Kim SK, Chang KW, Han SK, Yi HK, Jeon JG. *In vitro* inhibitory effects of *Polygonum cuspidatum* on bacterial viability and virulence factors of *Streptococcus mutans* and *Streptococcus sobrinus*. Archives of Oral Biology. 2006;51(12):1131-1140.
  7. Dzoyem JP, Nkuete AH, Kuete V, Tala MF, Wabo HK, Guru SK, Rajput VS, Sharma A, Tane P, Khan IA, Saxena AK, Laatsch H, Tan NH. Cytotoxicity and antimicrobial activity of the methanol extract and compounds from *Polygonum limbatum*. Planta medica. 2012;78(8):787-792.
  8. Rakesh KG, Kalra S, Mahadevan NS, Dhar VJ. Pharmacognostic investigation of *Polygonum nepalense*. International Journal of Recent Advances in Pharmaceutical Research. 2011;1:40-44.
  9. Focho DA, Newu MC, Anjah MG, Nwana FA, Ambo FB. Ethnobotanical survey of trees in Fundong, Northwest Region, Cameroon. Journal of Ethnobiology and Ethnomedicine. 2009;5:17.
  10. Del Vitto LA, Petenatti EM, Petenatti ME, Recursos herbolarios de San Luis (República Argentina). Primeira parte: plantas nativas. Mendoza. 1997;6:49-66
  11. Ahmed M, Khaleduzzaman M, Saiful Islam M. Isoflavan-4-ol, dihydrochalcone and chalcone derivatives from *Polygonum lapathifolium*. Phytochemistry. 1990;29:2009-2011.
  12. Pio Corrêa M, Dicionário das plantas uteis do Brasil e das exóticas cultivadas. IBDF - Rio de Janeiro: Imprensa Nacional. 1978;6.
  13. Peng ZF, Strack D, Baumert A, Subramaniam R, Goh NK, Chia TF, Tan SN, Chia LS, Antioxidant flavonoids from leaves of *Polygonum hydropiper* L. Phytochemistry. 2003;62(2):219-228.
  14. Singh G, Kachroo P. Forest Flora of Srinagar and plants of Neighborhood. Bishen Singh Mahendra Pal Singh: Delhi, India. 1976;27-218.
  15. Coffey T. The history and folklore of north American wild flowers. Houghton Mifflin: Boston, USA. 1994;36-189.
  16. Moerman D. Native American ethnobotany. Timber Press: Portland, Oregon, USA. 1998;42:691.
  17. Duwiejua M, Zeitlin IJ, Gray AI, Waterman PG. The anti-inflammatory compounds of *Polygonum bistorta*: Isolation and characterisation. Planta Medica. 1999;65(4):371-374.
  18. Matsuda H, Shimoda H, Morikawa T, Yoshikawa M, Phytoestrogens from the roots of *Polygonum cuspidatum* (*Polygonaceae*): Structure-requirement of hydroxyanthraquinones for estrogenic activity. Bioorganic & Medicinal Chemistry Letters. 2001;11(14):1839-1842.
  19. Sun X, Sneden AT. Neoflavonoids from *Polygonum perfoliatum*. Planta Medica. 1999;65(7):671-673.
  20. Murai Y, Kashimura S, Tamezawa S, Hashimoto T, Takaoka S, Asakawa Y, Kiguchi K, Murai F, Tagawa M. Absolute configuration of (6S,9S)-roseoside from *Polygonum hydropiper*. Planta Medica. 2001;67(5):480-481.
  21. Kim HJ, Woo ER, Park H. A novel lignan and flavonoids from *Polygonum aviculare*. Journal of Natural Products. 1994;57:587-596.
  22. Datta BK, Datta SK, Rashid MA, Nash RJ, Sarker SD. A sesquiterpene acid and flavonoids from *Polygonum viscosum*. Phytochemistry. 2000;54(2):201-205.
  23. Nonaka G, Miwa N, Nishioka I. Stilbene glycoside gallates and proanthocyanidins from *Polygonum multiflorum*. Phytochemistry. 1982;21:429-432.
  24. Tseng SH, Lin SM, Chen JC, Su YH, Huang HY, Chen CK, Lin PY, Chen Y. Resveratrol suppresses the angiogenesis and tumor growth of gliomas in rats. Clinical cancer research: An Official journal of the American Association for Cancer Research. 2004;10(6):2190-2202.
  25. Zhou LD, Zhou XH, Zhang SC. Inhibitive effect of water extracts of *Polygonum cuspidatum* on Ehrlich's carcinoma. Chinese Journal of Integrated Traditional Western Medicine. 1989;9:111.
  26. Lin HW, Sun MX, Wang YH, Yang LM, Yang YR, Huang N, Xuan LJ, Xu YM, Bai DL, Zheng YT, Xiao K. Anti-HIV activities of the compounds isolated from *Polygonum cuspidatum* and *Polygonum multiflorum*. Planta medica. 2010;76(9):889-892.
  27. Kimura Y, Okuda H. Resveratrol isolated from *Polygonum cuspidatum* root prevents tumor growth and metastasis to lung and tumor-induced neovascularization in Lewis

- lung carcinoma-bearing mice. The Journal of Nutrition. 2001;131(6):1844-1849.
28. Yan Y, Gao YY, Liu BQ, Niu XF, Zhuang Y, Wang HQ. Resveratrol-induced cytotoxicity in human Burkitt's lymphoma cells is coupled to the unfolded protein response. BMC Cancer. 2010;10:445.
  29. Hsu CM, Hsu YA, Tsai Y, Shieh FK, Huang SH, Wan L, Tsai FJ. Emodin inhibits the growth of hepatoma cells: Finding the common anti-cancer pathway using Huh7, Hep3B, and HepG2 cells. Biochemical and Biophysical Research Communications. 2010;392(4):473-478.
  30. Feng L, Zhang LF, Yan T, Jin J, Tao WY. Studies on active substance of anticancer effect in *Polygonum cuspidatum*. Zhong yao cai = Zhongyao cai = Journal of Chinese Medicinal Materials. 2006;29(7):689-691.
  31. Guo L, Peng Y, Yao J, Sui L, Gu A, Wang J. Anticancer activity and molecular mechanism of resveratrol-bovine serum albumin nanoparticles on subcutaneously implanted human primary ovarian carcinoma cells in nude mice. Cancer Biotherapy & Radiopharmaceuticals. 2010;25(4):471-477.
  32. van Ginkel PR, Sareen D, Subramanian L, Walker Q, Darjatmoko SR, Lindstrom MJ, Kulkarni A, Albert DM, Polans AS. Resveratrol inhibits tumor growth of human neuroblastoma and mediates apoptosis by directly targeting mitochondria. Clinical Cancer Research: An Official Journal of the American Association for Cancer Research. 2007;13(17):5162-5169.
  33. Kao CL, Huang PI, Tsai PH, Tsai ML, Lo JF, Lee YY, Chen YJ, Chen YW, Chiou SH. Resveratrol-induced apoptosis and increased radiosensitivity in CD133-positive cells derived from atypical teratoid/rhabdoid tumor. International Journal of Radiation Oncology, Biology, Physics. 2009;74(1):219-228.
  34. Chun-Guang W, Jun-Qing Y, Bei-Zhong L, Dan-Ting J, Chong W, Liang Z, Dan Z, Yan W. Anti-tumor activity of emodin against human chronic myelocytic leukemia K562 cell lines *in vitro* and *in vivo*. European Journal of Pharmacology. 2010;627(1-3):33-41.
  35. Brandao GC, Kroon EG, Duarte MG, Braga FC, de Souza Filho JD, de Oliveira AB. Antimicrobial, antiviral and cytotoxic activity of extracts and constituents from *Polygonum spectabile* mart. Phytomedicine: International Journal of Phytotherapy and Phytopharmacology. 2010;17(12):926-929.
  36. Ismail IF, Golbabapour S, Hassandarvish P, Hajrezaie M, Abdul Majid N, Kadir FA, Al-Bayaty F, Awang K, Hazni H, Abdulla MA. Gastroprotective activity of *Polygonum chinense* aqueous leaf extract on ethanol-induced hemorrhagic mucosal lesions in rats. Evidence-Based Complementary and Alternative Medicine: eCAM; 2012.
  37. Wasman SQ, Mahmood AA, Zahra AA, Salmah I. Cytoprotective activities of *Polygonum minus* aqueous leaf extract on ethanol-induced gastric ulcer in rats. Journal of Medicinal Plants Research. 2010;4:2658-2665.
  38. Hsu CY, Chan YP, Chang J. Antioxidant activity of extract from *Polygonum cuspidatum*. Biological research. 2007;40(1):13-21.
  39. Sheela QR, Ramani A. *In vitro* antioxidant activity of *Polygonum barbatum* leaf extract. Asian Journal of Pharmaceutical and Clinical Research. 2011;4:113-115.
  40. Bralley EE, Greenspan P, Hargrove JL, Wicker L, Hartle DK. Topical anti-inflammatory activity of *Polygonum cuspidatum* extract in the TPA model of mouse ear inflammation. Journal of Inflammation (Lond). 2008;5:1-7.
  41. Kinger KH, Gupta KM. Wound healing activity of *Polygonum barbatum* Linn. (whole plant). Journal of Pharmacy and Pharmaceutical Sciences. 2012;1:1084-1091.
  42. Lim BO, Lee JH, Ko NY, Mun SH, Kim JW, Kim do K, Kim JD, Kim BK, Kim HS, Her E, Lee HY, Choi WS. *Polygoni cuspidati* radix inhibits the activation of syk kinase in mast cells for anti allergic activity. Experimental Biology and Medicine (Maywood). 2007;232(11):1425-1431.
  43. Maradufu A, Ouma HJ. A new chalcone as a natural molluscicide from *Polygonum senegalense*. Phytochemistry. 1978;17.
  44. Midiwo JO, Omoto FM, Yenesew A, Akala MH, Wangui J, Liyala P, Wasunna C, Waters CN. The first 9-hydroxy homoisoflavanone, and antiplasmodial chalcones, from the aerial exudates of *Polygonum senegalense*. ARKIVOC. 2007;9:21-27.
  45. Wang H, Dong Y, Xiu ZL. Microwave-assisted aqueous two-phase extraction of piceid, resveratrol and emodin from *Polygonum cuspidatum* by

- ethanol/ammonium sulphate systems. *Biotechnology Letters*. 2008;30(12):2079-2084.
46. Wang QL, Li BY, Qiu SC, Li YL, Mi W, Song HY. Study on antibacteria effect *in vitro* of *Polygonum cuspidatum* sieb. *Lishizhen Medicine and Material Medica Research*. 2006;(17):762-763.
  47. Song JH, Yang TC, Chang KW, Han SK, Yi HK, Jeon JG. *In vitro* effects of a fraction separated from *Polygonum cuspidatum* root on the viability, in suspension and biofilms, and biofilm formation of mutans streptococci. *Journal of Ethnopharmacology*. 2007;112(3):419-425.
  48. Shan B, Cai YZ, Brooks JD, Corke H. Antibacterial properties of *Polygonum cuspidatum* roots and their major bioactive constituents. *Food Chemistry*. 2008;109:530-537.
  49. Ban SH, Kwon YR, Pandit S, Lee YS, Yi HK, Jeon JG. Effects of a bio-assay guided fraction from *Polygonum cuspidatum* root on the viability, acid production and glucosyl tranferase of mutans streptococci. *Fitoterapia*. 2010;81(1):30-34.
  50. Zhang Q, Wu C, Duan L, Yang J. Genotoxic studies on panax ginseng and *Polygonum multiflorum* and their combination in mouse peripheral lymphocyte cells. *Asian Journal of Traditional Medicine*. 2007;(2):217-134.
  51. Noda T, Yamada T, Ohkubo T, Omura T, Ono T, Adachi T, Awaya T, Tasaki Y, Shimizu K, Matsubara K. Hot-water-extracts of *Polygonum multiflorum* do not induce any toxicity but elicit limited beneficial effects on the liver in mice. *Journal of Health Science*. 2009;55:720-725.
  52. Jiang S, Lin K, Lu M. A conformation-specific monoclonal antibody reacting with fusion-active gp41 from the human immunodeficiency virus type 1 envelope glycoprotein. *Journal of Virology*. 1998;72(12):10213-10217.
  53. Zhang HS, Zhou Y, Wu MR, Zhou HS, Xu F. Resveratrol inhibited Tat-induced HIV-1 LTR transactivation via NAD(+)-dependent SIRT1 activity. *Life sciences*. 2009;85(13-14):484-489.
  54. Yiu CY, Chen SY, Chang LK, Chiu YF, Lin TP. Inhibitory effects of resveratrol on the Epstein-Barr virus lytic cycle. *Molecules*, 2010;15(10):7115-7124.
  55. Chang JS, Liu HW, Wang KC, Chen MC, Chiang LC, Hua YC, Lin CC. Ethanol extract of *Polygonum cuspidatum* inhibits hepatitis B virus in a stable HBV-producing cell line. *Antiviral Research*. 2005;66(1):29-34.
  56. Wang DG, Xu YR, Liu BB. Tissue distribution and excretion of resveratrol in rat after oral administration of *Polygonum cuspidatum* extract (PCE). *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2011;15:859-866.
  57. Peng J, Song ZF, Ma C. Emodin studies on pharmacokinetics and distribution in rat liver after *Polygonum cuspidatum* extract administration. *World science and technology medernization of traditional Chinese medicine and Materia Medica*. 2008;10:64-67.
  58. Lin SP, Chu PM, Tsai SY, Wu MH, Hou YC. Pharmacokinetics and tissue distribution of resveratrol, emodin and their metabolites after intake of *Polygonum cuspidatum* in rats. *Journal of Ethnopharmacology*. 2012;144(3):671-676.
  59. Jin WJ, Chen SY, Qian ZX, Shi XH. Effects of polydatin IV on inhibiting respiratory burst of PMNs and scavenging oxygen free radicals. *Chinese Pharmacological Bulletin*. 1993;355-357.
  60. Luo SZ, Zhang PW, Li RS. Dilating action of 3,4,5-trihydroxystibene-3-p-mono-D-glucoside on rabbit's blood vessels. *Journal of First Military Medical University*. 1992;12:10-13.
  61. Arichi H, Kimura Y, Okuda H, Baba K, Kozawa M, Arichi S. Effects of stilbene components of the root of *Polygonum cuspidatum* sieb. Et zucc. On lipid metabolism. *Chemical & Pharmaceutical Bulletin*. 1980;30:1766-1770.
  62. Jeong ET, Jin MY, Kim MS, Chang YH, Park SG. Inhibition of melagenesis by piceid isolated from *Polygonum cuspidatum*. *Archives of Phamacology Research*. 2010;33:1331-1338.
  63. Leu YL, Hwang TL, Hu JW. Anthraquinones from *Polygonum cuspidatum* as tyrosinase inhibitors for dermal use. *Phytotherapy Research*. 2008;22:552-556.
  64. Wang ZH, Huang TN, Guo SF, Wang RH. Effects of emodin extracted from *Rhizoma Polygoni cuspidati* in treating HSV-1 cutaneous infection in guinea pigs. *Journal of Anhui Traditional Chinese Medical College*. 2003;22:36-39.
  65. Yu CX, Zhang XQ, Kang LD, Zhang PJ, Chen WW, Liu WW, Liu QW, Zhang JY.

- Emodin induces apoptosis in human prostate cancer cell LNCaP. Asian Journal of Andrology. 2008;10:625-634.
66. Kuo TC, Yang JS, Lin MW, Hsu SC, Lin JJ, Lin HJ, Hsia TC, Liao CL, Yang MD, Fan MJ, Wood WG. Emodin has cytotoxic and protective effects in rat C6 glioma cells: Roles of Mdr1a and nuclear factor Kappa B in cell survival. Journal of Pharmacology and Experimental Therapeutics. 2009;330:736-744.
67. Mazid AM, Datta KB, Nahar L, Bashar KMAS, Bachar CS, Saker SD. Phytochemical studies on *Polygonum barbatum* (L) hara var. barbatua (*Polygonaceae*). Records of Natural Products. 2011;5(2):143-146.
68. Lopez SN, Manuel GS, Susana JG, Ricardo LF, Susana AZ. An unusual homoisoflavonone and a structurally-related dihydrochalcone from *Polygonum ferrugineum* (*Polygonaceae*). Phytochemistry. 2006;67:2152-2158.
69. Bulbul L, Uddin MJ, Sushanta SM, Roy J. Phytochemical screening, anthelmintic and antiemetic activities of *Polygonum lapathifolium* flower extract. European Journal of Medicinal Plants. 2013;3:333-344.

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