

Review **Endotyping Chronic Respiratory Diseases: T2 Inflammation in the United Airways Model**

Pasquale Ambrosino 1,* [,](https://orcid.org/0000-0002-9398-0428) Giuseppina Marcuccio ² [,](https://orcid.org/0000-0001-9211-5523) Giuseppina Raffio ² , Roberto Formisano ³ , Claudio Candia ⁴ [,](https://orcid.org/0000-0002-5888-8781) Fabio Manzo ⁵ [,](https://orcid.org/0009-0005-7493-5954) Germano Guerra ⁶ [,](https://orcid.org/0000-0002-4342-962X) Ennio Lubrano 3,6 [,](https://orcid.org/0000-0001-6189-5328) Costantino Mancusi [7](https://orcid.org/0000-0001-6690-1408) and Mauro Maniscalco 2,4,[*](https://orcid.org/0000-0001-6751-9921)

- 1 Istituti Clinici Scientifici Maugeri IRCCS, Scientific Directorate of Telese Terme Institute, 82037 Telese Terme, Italy
- $\overline{2}$ Istituti Clinici Scientifici Maugeri IRCCS, Pulmonary Rehabilitation Unit of Telese Terme Institute, 82037 Telese Terme, Italy; giuseppina.marcuccio@icsmaugeri.it (G.M.); giuseppina.raffio@icsmaugeri.it (G.R.)
- 3 Istituti Clinici Scientifici Maugeri IRCCS, Cardiac Rehabilitation Unit of Telese Terme Institute, 82037 Telese Terme, Italy; roberto.formisano@icsmaugeri.it (R.F.); ennio.lubrano@unimol.it (E.L.)
- ⁴ Department of Clinical Medicine and Surgery, Federico II University, 80131 Naples, Italy; claudio.candia@unina.it
- ⁵ Fleming Clinical Laboratory, 81020 Casapulla, Italy; fabioce26@gmail.com
- ⁶ Department of Medicine and Health Sciences, University of Molise, 86100 Campobasso, Italy; germano.guerra@unimol.it
- ⁷ Department of Advanced Biomedical Science, Federico II University, 80131 Naples, Italy; costantino.mancusi@unina.it
- ***** Correspondence: pasquale.ambrosino@icsmaugeri.it (P.A.); mauro.maniscalco@icsmaugeri.it (M.M.)

Abstract: Over the past 15 years, the paradigm of viewing the upper and lower airways as a unified system has progressively shifted the approach to chronic respiratory diseases (CRDs). As the global prevalence of CRDs continues to increase, it becomes evident that acknowledging the presence of airway pathology as an integrated entity could profoundly impact healthcare resource allocation and guide the implementation of pharmacological and rehabilitation strategies. In the era of precision medicine, endotyping has emerged as another novel approach to CRDs, whereby pathologies are categorized into distinct subtypes based on specific molecular mechanisms. This has contributed to the growing acknowledgment of a group of conditions that, in both the upper and lower airways, share a common type 2 (T2) inflammatory signature. These diverse pathologies, ranging from allergic rhinitis to severe asthma, frequently coexist and share diagnostic and prognostic biomarkers, as well as therapeutic strategies targeting common molecular pathways. Thus, T2 inflammation may serve as a unifying endotypic trait for the upper and lower airways, reinforcing the practical significance of the united airways model. This review aims to summarize the literature on the role of T2 inflammation in major CRDs, emphasizing the value of common biomarkers and integrated treatment strategies targeting shared molecular mechanisms.

Keywords: chronic obstructive pulmonary disease; asthma; chronic disease; disability; exercise; rehabilitation; outcome

1. Introduction

With an increasing worldwide incidence and over 4.0 million annual deaths globally [\[1\]](#page-14-0), chronic respiratory diseases (CRDs) are a leading cause of physical, psychological, and occupational disability $[2-4]$ $[2-4]$, resulting in increased healthcare costs and specific rehabilitation needs [\[5](#page-14-3)[–7\]](#page-14-4). Although the concept of CRDs might seem contradictory or outdated in the context of precision medicine, it remains highly relevant in terms of global health policies [\[8](#page-14-5)[,9\]](#page-14-6). Consequently, the World Health Organization (WHO) has established the WHO Chronic Respiratory Diseases Programme to support governments in their efforts to reduce the burden of morbidity, disability, and premature mortality associated with CRDs [\[10\]](#page-14-7). While the term CRDs is traditionally associated with more common conditions

Citation: Ambrosino, P.; Marcuccio, G.; Raffio, G.; Formisano, R.; Candia, C.; Manzo, F.; Guerra, G.; Lubrano, E.; Mancusi, C.; Maniscalco, M. Endotyping Chronic Respiratory Diseases: T2 Inflammation in the United Airways Model. *Life* **2024**, *14*, 899. [https://doi.org/10.3390/](https://doi.org/10.3390/life14070899) [life14070899](https://doi.org/10.3390/life14070899) **Example 12.5 Marson Control C**

Academic Editor: Chin Kook Rhee

Received: 25 June 2024 Revised: 12 July 2024 Accepted: 17 July 2024 Published: 19 July 2024

Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

like asthma, chronic obstructive pulmonary disease (COPD), or interstitial lung disease, the Global Burden of Disease Dictionary extends it to all pathological conditions affecting any part of the airways, from the nose to the alveoli [\[9\]](#page-14-6). In this scenario, the perspective of considering the upper and lower airways as a unified system has emerged as a promising and contentious topic, with the potential to revolutionize the management of CRDs [\[11\]](#page-14-8). Based on several shared characteristics, this model posits that the upper and lower respiratory tracts are part of a single, integrated system rather than two distinct entities [\[12\]](#page-15-0). Hence, it is reasonable to assume that recognizing airway pathology as an integrated entity could profoundly impact healthcare resource allocation and guide the implementation of pharmacological and rehabilitation strategies [\[13\]](#page-15-1).

In the era of precision medicine, endotyping has emerged as another novel approach to CRDs, whereby pathologies are categorized into distinct subtypes based on specific molecular mechanisms, gene expression patterns, and immune fingerprints [\[14\]](#page-15-2). This approach holds the potential to refine our understanding of the biological basis of CRDs, thereby enabling precise and targeted treatment strategies tailored to the specific endotype rather than solely focusing on clinical manifestations [\[15\]](#page-15-3). In this regard, endotyping CRDs has contributed to the growing acknowledgment of a group of conditions that, in both the upper and lower airways, share a common type 2 (T2) inflammatory signature [\[16\]](#page-15-4). These phenotypically diverse pathologies, spanning from allergic rhinitis to severe asthma, may often coexist, while also sharing a number of diagnostic and prognostic biomarkers as well as therapeutic approaches tailored to common molecular pathways [\[17\]](#page-15-5). Therefore, T2 inflammation may represent a unifying endotypic trait of the upper and lower airways, reinforcing the practical significance of the united airways model [\[13\]](#page-15-1).

This comprehensive review aims to summarize the literature's findings on the role of T2 inflammation in major CRDs of the upper and lower airways, emphasizing the usefulness of common biomarkers and unified treatment strategies directed at shared molecular mechanisms.

2. The Unified Airways Theory

From the nose to the alveoli, the airways share most of their anatomical and functional features in both physiological and pathological conditions, thus hypothetically determining a single morpho-functional unit [\[18\]](#page-15-6). As a matter of fact, the perspective of viewing the upper and lower airways as a unified system is traditionally based on two fundamental observations, namely the frequent association of their pathological conditions and the evidence of joint histopathological features with common innate and adaptive immune mechanisms [\[12\]](#page-15-0). This concept, now widely studied as the theory of the unified airways, dates back to more than 2000 years ago when Galenus (129–201 AD) first hypothesized that asthma and chronic rhinosinusitis (CRS) were comorbidities generated by the same pathogenetic mechanism, namely "a secretion that descends from the brain through the nose into the lungs" [\[19\]](#page-15-7). This ancient intuition is now supported by the evidence that over 80% of patients with asthma have allergic rhinitis and about 15%-38% of patients with rhinitis have asthma, with nasal symptoms being reported in up to 85% of asthma patients [\[20\]](#page-15-8). Although less frequent, the co-occurrence of rhinitis and asthma appears to extend to nonallergic forms of rhinitis as well [\[21\]](#page-15-9). This direct association between inflammation in the nasal mucosa and the lower airways is also supported by a significant correlation between the inflammatory profiles of nasal and bronchial biopsies in patients with CRS with nasal polyps (CRSwNP) [\[22\]](#page-15-10). Accordingly, it has been observed that the presence of nasal polyps may be linked to a greater decline in lung function and worse clinical outcomes in asthma patients [\[23\]](#page-15-11). Aspirin-exacerbated respiratory disease (AERD) is another condition characterized by the involvement of both the upper and lower airways [\[24\]](#page-15-12). In 1968, Samter and Beers described it as a triad characterized by the presence of nasal polyps, asthma, and aspirin sensitivity [\[25\]](#page-15-13). Later, it was reported that the hallmark of this condition is represented by exceptionally high levels of tissue and blood eosinophils [\[26\]](#page-15-14), thus leading

to the acknowledgment that this condition is characterized by T2 inflammation along with an overproduction of cysteinyl leukotrienes and inflammatory prostaglandins [\[27\]](#page-15-15).

Collectively, the observation that some phenotypically diverse pathologies frequently coexist has prompted a more thorough investigation into the hypothesis that they may be linked conditions manifesting in different locations, with inflammation serving as the linking factor [\[28\]](#page-15-16). This has even led the Global Asthma Association to take a clear position on united airway diseases, advocating for specific diagnostic and therapeutic strategies, as well as targeted education for medical students, physicians, health professionals, patients, and caregivers [\[29\]](#page-15-17). Translational research has further supported this hypothesis, confirming that the cascade of events resulting from mucosal insult and barrier disruption may lead in both the upper and lower airways to the production and release of specific proinflammatory cytokines with a common inflammatory signature (or endotype) [\[30\]](#page-15-18).

3. Inflammatory Endotypes in CRDs: Focus on T2 Inflammation

In recent times, there has been a surge in the popularity of the endotype-driven approach to CRDs in both scientific research and clinical practice [\[13\]](#page-15-1). This has led to the identification of three distinct immunological and inflammatory profiles, based on the prevalent activity of different types of T-helper (Th) cells, which modulate diverse molecular pathways and cytokine production [\[31\]](#page-15-19). Thus, the Th1/Th2 paradigm has been updated and extended in recent years with the identification of a third subset of effector T cells, known as Th17 cells [\[32\]](#page-15-20). In this regard, asthma has served as a sort of prototype in endotyping CRDs, leading to the identification of two major disease categories, namely those predominantly driven by T2 inflammation and those that are not [\[33\]](#page-15-21).

Interferon-gamma (IFN- γ) and tumor necrosis factor-alpha (TNF- α) are the main proinflammatory cytokines contributing to Th1-driven inflammation, which often results in neutrophilic infiltration with mucus hypersecretion and tissue damage, eventually leading to chronic bronchitis and emphysema [\[31\]](#page-15-19). Accordingly, Th1 responses are commonly observed in COPD and non-eosinophilic asthma, where neutrophilic inflammation predominates [\[34\]](#page-15-22). Th17 inflammation is also characterized by prevalent neutrophilic infiltration, driven by the production of cytokines such as interleukin (IL)-17 and IL-22 [\[35\]](#page-15-23). Mounting evidence indicates that Th17 immune response may coexist with both Th1 and Th2, thus exhibiting a closer interconnection than previously assumed [\[31\]](#page-15-19). For example, IFN- γ produced by Th1 cells has the capacity to stimulate the differentiation of Th17 cells, whereas IL-17 released by Th17 can in turn promote the generation of proinflammatory cytokines by Th1 cells. Moreover, contrary to the traditional belief that IL-17 only originates from Th17 cells, it is now recognized that IL-17 is also generated by IL-17-producing Th2 cells [\[36\]](#page-15-24), with a high prevalence of these dual-function cells being detected in both the blood and bronchoalveolar lavage fluid of individuals with severe steroid-resistant asthma [\[37\]](#page-16-0). Collectively, the Th17 inflammatory endotype and its complex crosstalk with other inflammatory patterns are frequently associated with severe asthma and CRS without nasal polyps (CRSsNP) [\[31\]](#page-15-19).

In allergic conditions, exposure to allergens triggers Th2-mediated (or T2) inflammation. In brief, inhaled allergens are captured by antigen-presenting cells (APCs), like dendritic cells, which promote the differentiation of naïve T cells in Th2 cells [\[38\]](#page-16-1). Th2 cells produce a particular set of cytokines, including IL-4, IL-5, and IL-13, driving airway eosinophilia, mucus hypersecretion through goblet cells, and parenchymal remodeling [\[38\]](#page-16-1). IL-4 and IL-13 stimulate B cells to switch from producing IgG to IgE, which in turn bind their receptors on mast cells and basophils and sensitize them to the allergen [\[39\]](#page-16-2). The key effectors of T2 response remain eosinophils, whose differentiation, activation, and recruitment from the bone marrow are orchestrated by IL-5 [\[40\]](#page-16-3). Eosinophils are structurally highly complex cells with a number of molecules and surface receptors, including IL-4 (IL-4R α) and IL-5 receptors alpha (IL-5R α) [\[40\]](#page-16-3). Additionally, eosinophils express surface receptors for various chemotactic agents, such as the type 3 C-C chemokine (CCR3) receptor, which binds eotaxins, as well as adhesion molecules that allow interactions with

airway epithelial cells and endothelial cells [\[41\]](#page-16-4). The biological characteristic of eosinophils lies in their intra-cytoplasmic granules, which can be released through a fragmentary de-granulation mechanism or following cytolysis [\[42](#page-16-5)[,43\]](#page-16-6). Four cationic proteins are contained in these granules, along with growth factors such as transforming growth factor β (TGF-β), metalloproteinases (MMPs), and tissue inhibitors of metalloproteinases (TIMPs), playing significant roles in fibrosis development and parenchymal remodeling [\[40\]](#page-16-3).

While allergen-specific Th2 cells are the key effectors of adaptive immunity, nonspecific innate lymphoid cells (ILC2s) also seem to be responsible for eosinophilic inflammation in CRDs characterized by a clear T2 signature even in the absence of allergen exposure [44]. The [prop](#page-16-7)osed hypothesis suggests that ILC2s may be activated via inflammatory mediators and alarmins released by the damaged epithelium, such as thymic stromal lympho-proteins (TSLP), interleukin (IL)-25, and IL-33 [45], wh[ich](#page-16-8) also amplify the T2 response by enhancing the ability of dendritic cells to present antigens to naïve T cells. sponse by enhancing the ability of dendritic cells to present antigens to naïve T cells. ILC2s, in turn, release the same cytokine panel that is characteristic of Th2 responses, thus ILC2s, in turn, release the same cytokine panel that is characteristic of Th2 responses, thus contributing to eosinophil recruitment, mucus production, and tissue repair processes seen contributing to eosinophil recruitment, mucus production, and tissue repair processes in allergic inflamm[atio](#page-16-9)n [46] ([Fig](#page-3-0)ure 1).

Figure 1. Innate and adaptive immunity in type 2 (T2) inflammation of upper and lower airways. **Figure 1.** Innate and adaptive immunity in type 2 (T2) inflammation of upper and lower airways. Th2: T-helper 2 lymphocytes; IgE: immunoglobulins E; TSLP: thymic stromal lympho-proteins; IL: Th2: T-helper 2 lymphocytes; IgE: immunoglobulins E; TSLP: thymic stromal lympho-proteins; IL: interleukin; ILC2s: non-specific innate lymphoid cells; IL-5Rα: interleukin-5 receptor subunit α. interleukin; ILC2s: non-specific innate lymphoid cells; IL-5Rα: interleukin-5 receptor subunit α.

4. Clinical Characteristics of T2 Inflammation 4. Clinical Characteristics of T2 Inflammation

Gaining a better understanding of the pathogenic mechanisms and distinct inflam-Gaining a better understanding of the pathogenic mechanisms and distinct inflammatory endotypes in CRDs has led to a novel approach to CRDs, especially asthma and matory endotypes in CRDs has led to a novel approach to CRDs, especially asthma and COPD [\[17](#page-15-5)]. As previously mentioned, asthma has acted as a prototype for endotyping COPD [17]. As previously mentioned, asthma has acted as a prototype for endotyping CRDs, resulting in the identification of two primary disease categories. Therefore, these CRDs, resulting in the identification of two primary disease categories. Therefore, these patients are now categorized as T2-high or T2-low, the former characterized by predomi-patients are now categorized as T2-high or T2-low, the former characterized by predominant eosinophilic inflammation and the latter by either neutrophilic or paucigranulocytic nant eosinophilic inflammation and the latter by either neutrophilic or paucigranulocytic inflammatory response [\[33\]](#page-15-21). Adult asthma patients with a T2-high profile tend to report

childhood onset and higher exacerbation rates [\[47\]](#page-16-10). Most importantly, epidemiological data suggest that most patients with difficult-to-treat asthma tend to have a T2-high disease [\[48\]](#page-16-11). It is also noteworthy that T2-high patients frequently exhibit specific allergen sensitization, but intermediate phenotypes of nonallergic T2-high patients also exist [\[49\]](#page-16-12). Conversely, these phenotypes are less contrasting for children, with no differences in lung function, exacerbations, asthma control, or steroid use when comparing T2-high and T2-low signatures [\[47\]](#page-16-10). The presence of an eosinophilic endotype has also been widely accepted in a relevant proportion of COPD patients, estimated to be as high as 40%, with substantial implications in terms of prognosis and therapy [\[50\]](#page-16-13). Data from the large Copenhagen General Population Study suggest a 1.76-fold increased risk of severe exacerbations for eosinophilic COPD patients [\[51\]](#page-16-14). On the other hand, based on meta-analytical data, it can be argued that eosinophilia in COPD is associated with reduced length of hospital stay and better steroid response in terms of pulmonary function [\[52\]](#page-16-15).

In the upper airways, the role of T2 inflammation has long been recognized and even more clearly defined [\[53\]](#page-16-16). While there is abundant evidence for allergic rhinitis and CRSwNP [\[54\]](#page-16-17), the clinical evidence of T2 inflammation in nonallergic rhinitis (NAR) remains more debated. NAR includes conditions with similar clinical characteristics, such as nonallergic rhinitis with eosinophilia syndrome (NARES) and nonallergic rhinitis with eosinophils and mast cells (NARESMA) [\[55\]](#page-16-18). These disabling diseases exhibit symptoms consistent with allergic rhinitis, but allergen skin testing shows an absence of atopy while nasal cytology reveals a characteristic eosinophilic infiltration [\[56\]](#page-16-19). Epidemiological data indicate that NARES and NARESMA share several clinical characteristics with other CRDs with nonallergic T2 signatures, including late-onset, association with other T2 comorbidities, and a significant response to corticosteroids [\[57\]](#page-16-20). Eosinophilic inflammation is also involved in most cases of CRSwNP [\[54\]](#page-16-17), but it is less known that up to 49% of the patients with CRSsNP display a type 2 immune response as well [\[58\]](#page-16-21). However, compared to CRSwNP, the absolute number of infiltrating eosinophils is extensively (18-fold) lower in T2-driven CRSsNP, and this might explain why nasal polyp formation does not occur in CRSsNP despite type 2 presentation [\[58\]](#page-16-21). T2 inflammation, thus, appears to be implicated in most cases where mucosal damage in the upper airways results in polyp formation. Recently, it has been demonstrated that nasal polyposis in AERD also has type 2 characteristics and can be distinguished from CRSwNP by enhanced tissue eosinophilia with (surprisingly) no concomitant increases in conventional T2 inflammatory mediators associated with eosinophil proliferation and recruitment [\[26\]](#page-15-14).

In light of the common molecular pathways and shared clinical traits, some authors have even proposed a new approach to these respiratory conditions, suggesting that labels such as CRSwNP and asthma may be oversimplified, emphasizing the identification and treatment of the common endotypic profile [\[17\]](#page-15-5). While this perspective is (of course) provocative and might be seen as extreme, it underscores the importance of identifying biomarkers of T2 inflammation to support personalized pharmacological and rehabilitation strategies aimed at managing recurrences, clinical deterioration, and disease exacerbation [\[15\]](#page-15-3). The ambitious goal is to develop unified approaches for patients with T2 inflammatory diseases of the upper and lower airways, especially when these conditions coexist [\[53\]](#page-16-16).

5. Biomarkers of T2 Inflammation

Recently, strong evidence has endorsed a biomarker-driven approach to CRDs, targeting individuals with specific endotypic traits and some common clinical characteristics who are, therefore, more likely to respond to specific pharmacological and rehabilitation strategies [\[59\]](#page-16-22). As we discuss below, this approach has gained popularity with the growing recognition of T2-driven diseases.

5.1. Blood and Tissue Eosinophils

In this scenario, it becomes clear how identifying eosinophils in the bloodstream and respiratory tract fluids (such as sputum, nasal secretions, and bronchial lavage) could facilitate the recognition of T2 inflammatory patterns [\[60\]](#page-16-23). However, while induced sputum sampling for airway examination is the gold standard but uncommon in routine clinical practice for technical reasons [\[61\]](#page-16-24), the count of eosinophils in peripheral blood is frequently utilized as a surrogate marker for eosinophilic airway inflammation [\[62\]](#page-16-25). In fact, blood eosinophils are correlated with pulmonary eosinophil counts from sputum or lung tissue in both asthmatic and COPD patients [\[63\]](#page-17-0), although cardiac diseases in COPD and gastrointestinal reflux disease in asthma can diminish this association [\[64\]](#page-17-1). Collectively, it is important to highlight the evidence that this correlation is stronger in asthma than in COPD [\[65\]](#page-17-2), with blood eosinophils representing accurate biomarkers of asthma control and airway eosinophilia across all severity degrees of asthma [\[66\]](#page-17-3). In this regard, one should also consider that distinct phenotypes of blood and sputum eosinophils have been observed in COPD and asthma patients, with elevated levels of CD193+ and CD66b+ eosinophils reported in COPD and increased levels of CD11b+ cells in asthma [\[67\]](#page-17-4). Overall, while the evidence regarding the clinical significance of eosinophils is stronger and more intuitive in the case of asthma, with a better correlation between peripheral and airway eosinophilia [\[68\]](#page-17-5), the data on COPD are more conflicting and subject to different clinical interpretations [\[63\]](#page-17-0). In this regard, it has been shown that tissue eosinophils that are found in large numbers in COPD exacerbations and in smaller numbers in stable COPD are not accompanied by an increase in IL-5, suggesting that they may have different functions and recruiting mechanisms than eosinophils in asthma [\[69\]](#page-17-6). Accordingly, recent evidence even indicates that blood eosinophils cannot be assumed to reflect lung tissue eosinophils in COPD [\[70\]](#page-17-7). However, data from the large Copenhagen General Population Study cohort suggest that increased blood eosinophil levels are linked to a higher risk of exacerbations in COPD [\[51\]](#page-16-14). Additionally, during acute COPD exacerbations requiring hospitalization, blood eosinophilia has been shown to predict a higher short-term treatment success rate but also a higher rate of readmissions [\[71\]](#page-17-8). Similar conclusions on the risk of exacerbations are drawn from the SPIROMICS and the ECLIPSE studies [\[50](#page-16-13)[,72\]](#page-17-9). However, the SPIROMICS also found that blood eosinophilia performed worse than sputum eosinophilia in identifying a subgroup of patients with reduced forced expiratory volume in $1 s$ (FEV₁) and markers of emphysema on a computed tomography (CT) scan [\[72\]](#page-17-9). Conversely, blood eosinophilia in the ECLIPSE study was linked to higher $FEV₁$, better exercise capacity, lower symptom severity and emphysema score, and an even lower bacterial load [\[50\]](#page-16-13). Despite some conflicting findings in the literature, it is important to highlight that peripheral blood eosinophils are, however, included in the most recent Global Initiative for Chronic Obstructive Lung Disease (GOLD) statement to drive add-on therapy with steroids in COPD [\[73\]](#page-17-10).

Evidently, when discussing eosinophils as biomarkers of T2 inflammation, certain considerations must be made regarding the individual variability of blood and tissue eosinophils, as well as the confounding factors that can influence them. In this regard, it has been reported that a single measurement might not be sufficient in asthma due to the variability of blood eosinophils over time and evidence that roughly one in six patients whose average count is above 150 cells/ μ L may initially have counts below this threshold [\[74\]](#page-17-11). Moreover, in COPD as well as other clinical conditions, the threshold value appears to be important, with values below 150 cells/ μ L or above 300 cells/ μ L associated with a lower level of individual variability [\[75](#page-17-12)[,76\]](#page-17-13). Numerous other factors can influence the measurement, including smoking, seasonality, technical analytical factors [\[75\]](#page-17-12), and a range of pathological conditions such as nasal polyps, vasculitis, gastrointestinal diseases, leukemias, Hodgkin's lymphoma, and adverse drug reactions [\[41\]](#page-16-4). Given that eosinophils are used to predict therapeutic responses, it is reasonable to assume that these therapies would also impact eosinophil levels. The administration of either inhaled (ICSs) or oral corticosteroids (OCSs) results in a reduction in eosinophil count in both asthma and COPD, which is dose-dependent and correlates with the effectiveness of corticosteroid treatment and suppression of eosinophilic airway inflammation [\[77](#page-17-14)[,78\]](#page-17-15). Furthermore, in COPD, both ICSs and OCSs have been shown to significantly suppress sputum eosinophils [\[79,](#page-17-16)[80\]](#page-17-17). When a paradoxical increase in blood eosinophiles is reported following ICS administration in COPD, more frequent exacerbations and an accelerated lung function decline should be expected [\[81\]](#page-17-18).

In the upper airways, likely due to greater accessibility, the role of circulating eosinophils has been less investigated in pathological processes, with nasal cytology instead being given greater impor[tan](#page-17-19)ce [82]. Increased tissue eosinophil counts have been detected in cytology following nasal curettage or in fluids obtained from nasal lavage in CRSwNP patients, showing a close association with the degree of epithelial damage and the development of [po](#page-17-19)lyps [82]. This aligns with the evidence suggesting that the inflammatory process driving CRSwNP is primarily mediated by eosinophils, either in the context of allergic or nonallergic [rhin](#page-17-20)itis [83]. In CRSwNP, tissue eosinophilic infiltration appears to be linked with a delay in eosinophil apoptotic mechanisms, leading to an amplification of the epithelial-to-mesenchymal transition response ind[uce](#page-17-19)[d by](#page-17-21) TGF-β [82,84]. A similar process occurs in the lower airways during T2-high asthma, where eosinophilic inflammation persists [85]. Accordingly, a strong correl[atio](#page-17-22)n between nasal and sputum eosinophilia has been reported in adults with moderate-to-severe asthma, with nasal eosinophilia via quantitative cytology also linked to br[onch](#page-17-23)odilator response [86]. These findings support recent research indicating similar inflammatory changes in the tissues of both asthma and rhinitis patients, reinforcing the concept of the upper and lower airw[ays](#page-18-0) as a unified entity [87]. Therefore, assessing upper airway inflammation could offer insights into lower airway involvement, suggesting that nasal eosinophilia might serve as an alternative to sputum analysis and the nose may serve as a sort of window to the bronchial tract $[87]$. In this regard, due to its ease of execution and non-invasiveness, it has been reported that nasal lavage cytology can be considered a valid substitute for sputum cytology in the diagnosis of the different inflammatory phenotypes of asthma, with an even higher performance than blood cell count [\[88\]](#page-18-1) (Figure 2).

Figure 2. Eosinophils in nasal lavage (Panel A) and sputum cytology (Panel B). **Figure 2.** Eosinophils in nasal lavage (Panel A) and sputum cytology (Panel B).

5.2. Fractional Exhaled Nitric Oxide (FeNO)

Along with blood eosinophil count, fractional exhaled nitric oxide (FeNO) is widely accepted as another surrogate marker of airway eosinophilia [\[89\]](#page-18-2). With different functions attributed to nitric oxide (NO) in the airways (e.g., vasodilator, bronchodilator, neurotransmitter), FeNO originates from the activity of various nitric oxide synthase (NOS) enzymes, but the predominant source of elevated NO levels arises from the inducible NOS 2 (iNOS2) enzyme, expressed in epithelial and immune cells in response to inflammation [\[89\]](#page-18-2). In contrast to a sputum eosinophil assessment, which is technically challenging and time-consuming, FeNO is non-invasive and easily reproducible [\[90\]](#page-18-3) and demonstrates a strong correlation with airway eosinophilic inflammation detected in induced sputum, thus offering a noninvasive means to evaluate T2 airway inflammation in asthma [\[91\]](#page-18-4). The correlation between blood eosinophil count and FeNO as prognostic biomarkers and their predictive role in asthma exacerbation risk has garnered interest in recent times. In a large population-based study, individuals with asthma who exhibited elevated FeNO or elevated blood eosinophiles (\geq 300 cells/ μ L) had a higher risk of exacerbation (risk ratio: 1.31), but when both biomarkers were combined, the risk ratio for acute exacerbation increased to 3.67 [\[92\]](#page-18-5).

As a biomarker of T2 inflammation, there is a tendency for FeNO levels to correlate with increased total serum immunoglobulin E (IgE) or positive results in skin prick testing among individuals with allergic asthma [\[93\]](#page-18-6). Interestingly, increased FeNO lacks complete specificity for asthma, as it can also be elevated in individuals with atopy who do not display additional asthma characteristics [\[94\]](#page-18-7). Similarly, other clinical conditions, particularly those characterized by T2 inflammation, can affect FeNO values, including CRS and allergic rhinitis [\[95\]](#page-18-8). In this regard, it is important to highlight that orally exhaled gases contain NO derived from both the lower and upper airways [\[96\]](#page-18-9). However, although some promising results have been obtained on the use of FeNO in CRSwNP monitoring [\[97\]](#page-18-10), its utility in upper airway diseases is limited. Nevertheless, their simultaneous presence should always be carefully considered when utilizing this biomarker in asthma.

In 2011, the American Thoracic Society (ATS) formulated a clinical practice guideline to interpret FeNO levels, defining values above 50 parts per billion (ppb) in adults and above 35 ppb in children as elevated [\[98\]](#page-18-11). In this regard, it is important to note that these cut-offs may vary among international guidelines and that FeNO values are influenced by various confounding factors, including age, sex, smoking habits, respiratory infections, and environmental conditions [\[99\]](#page-18-12). Cigarette smoking is notably significant, potentially reducing FeNO levels by up to 50% depending on the amount smoked [\[100\]](#page-18-13). Gender differences also play a role, with women consistently showing approximately 25% lower FeNO levels compared to men [\[99\]](#page-18-12). Respiratory infections, particularly those of viral origin, are linked to increased FeNO levels [\[99\]](#page-18-12). Additionally, supported by meta-analytical evidence, the use of ICSs and OCSs has been shown to significantly lower FeNO in both asthma and COPD patients [\[101](#page-18-14)[–103\]](#page-18-15). However, numerous studies have demonstrated the ability of FeNO to predict airway inflammation, exacerbation risk, and poor asthma control [\[91\]](#page-18-4). Therefore, ATS guidelines suggest that FeNO levels exceeding the proposed cut-off values can indicate responsiveness to corticosteroids in asthma [\[98\]](#page-18-11). Recently, the potential utility of FeNO has also been proposed for monitoring T2 inflammation in clinical contexts other than asthma [\[91\]](#page-18-4). A post hoc analysis of the PREVENT study suggested an association between COPD exacerbations and elevations in FeNO [\[104\]](#page-18-16), but a recent report on stable COPD patients revealed some instability in FeNO values, which seemed unrelated to eosinophil counts but somewhat associated with body weight [\[105\]](#page-18-17). This is partially consistent with the results of another cohort study, showing that the association between FeNO and blood eosinophils in COPD, although statically significant, is very weak [\[106\]](#page-18-18). Overall, this raises questions about its diagnostic and prognostic value in this clinical setting and supports the view that frequent FeNO measurements in COPD might have clinical utility in identifying individual patterns and their variations over time [\[107\]](#page-18-19).

While FeNO is a method with numerous potential clinical applications, it cannot reveal the source of NO production. Therefore, in relatively recent times, extended NO analysis has gained traction, including measurements of NO at multiple flow rates to discern the origin of NO from alveolar or bronchial sources [\[108\]](#page-18-20). Partitioning exhaled NO has shown elevated distal NO levels in patients with asthma and COPD [\[108\]](#page-18-20), with some evidence suggesting that these levels may not change with steroid treatment in either clinical settings [\[109\]](#page-18-21). Increased alveolar NO levels have been proposed for predicting severe, frequent, and near-term exacerbations in children with asthma [\[110\]](#page-18-22) but not in adults with COPD [\[111\]](#page-19-0). In asthma, there is also emerging evidence indicating that high distal NO may be related to distal T2 inflammation and steroid-dependent asthma [\[112\]](#page-19-1), supported by positive correlations found between alveolar NO concentration and bronchoalveolar lavage eosinophil counts [\[113\]](#page-19-2). In COPD, higher alveolar NO levels have been associated with poorer physical capacity and lower oxygen saturation following physical testing in stable states [\[114\]](#page-19-3), and are linked to accelerated decline in lung function over a two-year follow-up period [\[115\]](#page-19-4). However, similar to FeNO, data on the clinical utility of partitioning exhaled NO as a biomarker of T2 inflammation in COPD are still debated.

5.3. Immunoglobulin E (IgE)

Allergen-specific IgE is integral to the pathogenesis of allergic disorders, thus representing a reliable biomarker for symptom severity in patients with allergic rhinitis [\[116\]](#page-19-5). Allergen-specific IgE has also demonstrated relatively high sensitivity in diagnosing asthma [\[117\]](#page-19-6), with approximately 80% of patients with severe asthma exhibiting spe-cific IgE levels greater than 0.35 IU/mL [\[118\]](#page-19-7). Conversely, the role of total serum IgE as a biomarker of T2 inflammation is less clear and remains debated. Elevated IgE levels have been observed in nonallergic late-onset asthma and nasal polyps [\[119\]](#page-19-8), indicating that IgE-mediated disease can occur independently of allergen exposure. This is not surprising, as IL-4 and IL-13 can promote B cell class switching and IgE production [\[39\]](#page-16-2). Accordingly, it has been proposed that high total IgE may represent an additional biomarker for clustering T2-low asthma patients, thus identifying a group of patients with a peculiar T2-low phenotype, which is very similar to the T2-high phenotype in terms of disease severity and nasal comorbidities [\[120\]](#page-19-9). However, many studies have also shown that total IgE has a lower predictive value for airway eosinophilia compared to blood eosinophil count and FeNO, regardless of the cut-off used [\[121\]](#page-19-10). Not even this should be surprising, since IgE production occurs downstream of Th2 cell activation and the release of IL-4 and IL-13; thus, it can only address one aspect of the type 2 inflammatory response. Additionally, IgE paradoxically does not predict the likelihood of treatment response to the anti-IgE therapy with omalizumab [\[122\]](#page-19-11). Therefore, the primary clinical application of total IgE is to facilitate the dosing of omalizumab when initiating therapy, and its value as a biomarker of T2 inflammation should always be interpreted in combination with allergen sensitization tests, FeNO, eosinophils, and/or other emerging biomarkers [\[93\]](#page-18-6).

Naturally, as already pointed out for other T2 inflammation biomarkers, the interpretation of specific and total IgE levels must always consider a series of modifiable and non-modifiable factors that can influence them. For example, total serum IgE concentrations are higher in males [\[123\]](#page-19-12) and tend to increase with age [\[124\]](#page-19-13), while allergen-specific IgE shows a clear trend toward reduction with aging [\[124\]](#page-19-13). Among modifiable factors, total and specific serum IgE are higher among alcohol abusers [\[125\]](#page-19-14) and smokers [\[126\]](#page-19-15). Especially during exacerbations, increased total and fungus-specific IgE concentrations have been reported in fungal colonization of the airways (e.g., *A. fumigatus*) [\[127\]](#page-19-16), including asthma with fungal sensitization (SAFS) and allergic bronchopulmonary aspergillosis (ABPA) [\[128\]](#page-19-17). Few results are available regarding the impact of OCS and ICS. Most seem to demonstrate that steroids can markedly reduce both specific and total IgE [\[129–](#page-19-18)[131\]](#page-19-19), though a temporary increase may occur after the initiation of therapy [\[129,](#page-19-18)[132\]](#page-19-20). This should not be surprising, since serum IgE levels are known to fall when exposure to an allergen is reduced, and any treatment that reduces airway inflammation and, thus, reduces airway permeability to

allergens is likely to cause a reduction in serum IgE levels [\[119\]](#page-19-8). However, some conflicting and contrary results have also been reported in this regard [\[133\]](#page-19-21).

5.4. Emerging Biomarkers

New biomarkers for distinguishing T2 inflammation have been proposed, including the most promising serum periostin, eotaxin, and thymus and activation-regulated chemokine (TARC) [\[53\]](#page-16-16). The expression of periostin in the blood is triggered via the IL-4 and IL-13 via activation of the STAT6 pathway, directly leading to the transcription of the periostin gene [\[134\]](#page-19-22). This heightened periostin expression plays a role in tissue remodeling and fibrosis, which are crucial processes in both upper and lower airway diseases, including allergic rhinitis, CRSwNP, asthma, and COPD [\[13\]](#page-15-1). TARC is instead a chemokine with a high binding affinity for type 4 C-C chemokine receptor (CCR4), thus facilitating the recruitment, activation, and development of Th2 cells that express CCR4 [\[135\]](#page-19-23). Similarly, eotaxin selectively binds CCR3, which is expressed on both eosinophils and Th2 cells, thus controlling the migration and recruitment of eosinophiles in inflammatory sites [\[136\]](#page-20-0).

The evidence on the potential clinical use of these biomarkers is scarce but holds promise to lay the foundation for future research. To date, sputum eotaxin concentration has been shown to be significantly raised in moderate-to-severe, but not in mild, asthma, thus suggesting that its contribution is more important in more severe disease [\[137\]](#page-20-1). Accordingly, elevated serum levels of eotaxin-3 have been shown to distinguish moderate-to-severe asthma from healthy individuals [\[138\]](#page-20-2). In allergic asthma, TARC levels are significantly higher than in healthy controls, with a negative correlation between TARC levels and $FEV₁$ and a positive association with total IgE [\[135\]](#page-19-23). Increased blood levels of eotaxin, periostin, and TARC have also been reported in CRS as compared to healthy controls, with higher values in CRSwNP than in CRSsNP [\[139\]](#page-20-3). Interestingly, a decrease in these biomarkers has been observed after dupilumab therapy in CRSwNP [\[140\]](#page-20-4). Further studies are awaited to clarify the cost–benefit ratio in the current clinical practice of these and other promising biomarkers of T2 inflammation.

6. Treating T2 Inflammation: Standard Therapies and Unmet Needs

Standard treatment for respiratory diseases varies based on clinical manifestations, the location of the pathological process in the airways, its underlying mechanisms, and the presence of comorbid conditions [\[141\]](#page-20-5). Nevertheless, it is important to note that certain drug categories, often used in combination, are the mainstay of pharmacological treatment for different airway conditions. These include (but are not limited to) short- (SABAs) and long-acting β2 agonists (LABAs), long-acting muscarinic antagonists (LAMAs), and leukotriene receptor antagonists (LTRAs) [\[141\]](#page-20-5). ICSs, OCSs, and intranasal corticosteroids (INCSs), in doses ranging from low to high, are, however, the cornerstone for treating conditions of the upper and lower airways with T2 response. Accordingly, while the Global Initiative for Asthma (GINA) guidelines emphasize the use of ICS-formoterol for asthma treatment across all severity levels [\[142\]](#page-20-6), with a high baseline FeNO predicting responsiveness [\[98\]](#page-18-11), the GOLD guidelines recommend adding corticosteroids to dual bronchodilator therapy when eosinophil levels exceed 300 cells/ μ L [\[73\]](#page-17-10). INCS use is instead virtually recommended for all patients with CRSwNP, reserving the decision to switch to more targeted pharmacological therapies for cases that remain uncontrolled even after surgery, or for those where comorbidities with asthma or dermatological conditions necessitate an integrated approach [\[143\]](#page-20-7).

In this regard, it has been reported that CRSwNP has a recurrence rate of up to 80%, despite medical and endoscopic surgery [\[144\]](#page-20-8). Similarly, other T2-driven conditions in the lower airways may remain uncontrolled with steroid-based standard treatments as well. According to the GINA guidelines, asthma control is often deemed unrealistic for a significant portion of patients due to steroid resistance and persistent uncontrolled symptoms [\[142\]](#page-20-6). The epidemiological data on the rate of uncontrolled asthma vary significantly depending on the geographical areas and populations considered, but the Centers for

Disease Control and Prevention (CDC) report that up to 60% of adults [\[145\]](#page-20-9) and 44% of children [\[146\]](#page-20-10) with a confirmed diagnosis have uncontrolled asthma. Disease control may also be challenging in COPD patients, where acute exacerbations significantly impact the clinical course, causing rapid and partially irreversible lung function loss and the worsening of the long-term outcome. Frequent exacerbators, defined as those with at least two treated exacerbations per year [\[73\]](#page-17-10), represent 22% of the COPD population and face increased hospital admissions, comorbidities, and mortality, making them a priority for research and treatment [\[147\]](#page-20-11). It is interesting to note that patients with higher blood eosinophil levels during stable periods tend to experience more frequent and severe exacerbations [\[148\]](#page-20-12) and, although eosinophilic exacerbations are linked to rapid symptomatic recovery [\[52\]](#page-16-15), this also presents an opportunity to consider eosinophilic inflammation as a treatable trait in COPD, paving the way for more personalized pharmacological and rehabilitation strategies in selected cases [\[53\]](#page-16-16).

7. Precision Medicine for T2 Inflammatory Endotypes

The goal of managing various inflammatory endotypes in airway diseases is to achieve symptom control, reduce the risk of relapses and exacerbations, and prevent disease progression. In recent times, the use of biologics designed to interfere with specific inflammatory pathways has become central to this approach, thus reducing the risks of prolonged steroid therapies which include (but are not limited to) immunosuppression, weight gain, diabetes, and osteoporosis [\[149\]](#page-20-13).

Thus, the need to selectively inhibit T2 inflammation and prevent airway remodeling induced by eosinophils has led to the introduction and commercialization of six biological therapies approved for T2-high asthma and other type 2 inflammatory diseases. These therapies encompass the anti-IgE medication omalizumab, two anti-IL-5 (mepolizumab and reslizumab), the anti-IL-5 receptor subunit α (anti-IL-5R α) benralizumab, the anti-IL-4 receptor subunit α (anti-IL-5R α) dupilumab, and the anti-thymic stromal lymphopoietin (TSLP) tezepelumab. Each of these therapies targets specific pathways at different levels of the T2 inflammatory cascade. Therefore, monitoring specific biomarkers of T2 inflammation may help tailor therapies to individual patients while also monitoring adherence and effectiveness [\[13\]](#page-15-1). In this regard, if asthma has served as a prototype disease in the testing of biologics, it is noteworthy how the GINA guidelines recommend considering their use for uncontrolled allergic asthma or for asthma with features of T2 inflammation, defined by baseline blood eosinophils ≥ 150 cells/ μ L, FeNO ≥ 20 ppb, or sputum eosinophils $\geq 2\%$ [\[142\]](#page-20-6).

7.1. Anti-IgE

Omalizumab is a humanized monoclonal antibody that selectively binds the highaffinity Fc epsilon receptor of IgE on eosinophils, basophils, and mast cells [\[150\]](#page-20-14). Based on the results of randomized controlled trials [\[151,](#page-20-15)[152\]](#page-20-16), it was first licensed in 2003 for children with uncontrolled allergic asthma aged 12 years or older, later receiving approval for pediatric use (\geq 6 years) due to its good safety profile in clinical trials [\[153\]](#page-20-17). According to GINA guidelines, sensitization to (an) inhaled allergen(s) on a skin prick testing or specific IgE should always be demonstrated before starting treatment [\[142\]](#page-20-6). Also used in chronic spontaneous urticaria, omalizumab has also shown effectiveness in CRSwNP with inadequate response to ICS in phase III studies [\[154\]](#page-20-18), thus being recently approved for this indication as well. However, given its mechanism of action, omalizumab could have additional clinical applications in T2-driven diseases. This biologic has shown good efficacy and a favorable safety profile in severe asthma with overlapping COPD [\[155\]](#page-20-19), and based on meta-analytical data from randomized controlled trials [\[156\]](#page-20-20), also in patients with inadequately controlled allergic rhinitis. In a prospective randomized controlled open-label trial, a single injection of 300 mg of omalizumab two weeks before the start of the pollen season demonstrated better overall control of symptoms and quality of life, with the significantly reduced use of allergy symptom-relieving medications compared to

standard therapies [\[157\]](#page-20-21). Consequently, omalizumab is also approved in Japan for treating severe Japanese cedar pollinosis, a form of seasonal allergic rhinitis that affects up to 38.8% of the Japanese population [\[158\]](#page-21-0).

Considering that IgE production occurs downstream of Th2 cell activation and the release of IL-4 and IL-13, it can be argued that omalizumab may be effective in reducing immediate hypersensitivity reactions and acute symptoms, with limited impact on chronic inflammation and lung function [\[53\]](#page-16-16). For the same reason, while little to no impact on most biomarkers of T2 inflammation aside from IgE would be expected, it is noteworthy that omalizumab paradoxically increases total serum IgE by three to five times, maybe as a consequence of the prolonged half-life of IgE/anti-IgE complexes [\[159\]](#page-21-1). This increase in total IgE levels can persist for over a year after discontinuing treatment [\[160\]](#page-21-2). Paradoxically, IgE levels do not even predict the likelihood of response to omalizumab therapy [\[122\]](#page-19-11). Therefore, if the appropriate dose and frequency of omalizumab administration are determined by baseline IgE levels [\[161\]](#page-21-3), this biomarker does not seem to be useful in identifying good responders and monitoring therapy. Conversely, based on the results of clinical trials such as the EXTRA study [\[122\]](#page-19-11), the ERS/ATS guidelines for the management of severe asthma recommend using a FeNO cut-off of \geq 19.5 ppb to identify patients with a higher probability of benefitting from anti-IgE treatment [\[162\]](#page-21-4). Contrary findings emerged from the PROSPERO study on this matter, indicating that patients responded similarly to omalizumab therapy irrespective of their initial FeNO levels [\[163\]](#page-21-5). Given the above conflicting results, a recent meta-analysis was performed, revealing that elevated IgE and blood eosinophil levels correlate with increased FeNO levels and that all these biomarkers of T2 inflammation may be useful for selecting asthma patients who may benefit more from omalizumab [\[164\]](#page-21-6).

7.2. Anti-IL-5/5Rα

The anti-IL5 monoclonal antibodies mepolizumab and reslizumab work by preventing IL-5 from binding to its receptor, whereas the anti-IL-5R α antibody benralizumab directly targets and blocks the α subunit of the receptor on eosinophils and basophils [\[53\]](#page-16-16). The results and post hoc analyses of phase III trials suggest that mepolizumab is able to enhance lung function and decrease the frequency of exacerbations in asthmatic patients with blood eosinophil levels exceeding 150 cells/ μ L [\[165,](#page-21-7)[166\]](#page-21-8). In phase III trials, benralizumab also reduced annual exacerbation rates and was generally well tolerated for patients with severe uncontrolled asthma and blood eosinophils of 300 cells/ μ L or greater [\[167](#page-21-9)[,168\]](#page-21-10), with the phase IV SHAMAL study confirming that patients controlled on benralizumab can have meaningful reductions in ICS therapy while maintaining asthma control [\[169\]](#page-21-11). Similar positive results in terms of lung function and asthma control have been reported in phase III trials with reslizumab [\[170](#page-21-12)[,171\]](#page-21-13), even in the presence of CRSwNP with or without aspirin sensitivity [\[170\]](#page-21-12). In two randomized controlled trials, mepolizumab treatment also improved symptoms and quality of life in severe CRSwNP patients [\[172](#page-21-14)[,173\]](#page-21-15), while determining an increase in $FEV₁$ after 25 weeks of treatment [\[172\]](#page-21-14). Recently, the results of the phase III OSTRO trial (NCT03401229) were published, showing promising outcomes also for benralizumab in reducing nasal obstruction, symptoms, and airway eosinophilia in CRSwNP. As of this review, no phase III trial on the use of reslizumab in CRSwNP has yet been concluded. Based on the available results and their mechanisms of action, major regulatory agencies have, to date, approved these biologics for treating severe eosinophilic asthma, with mepolizumab also being approved for use in CRSwNP.

In recent times, given the growing acknowledgment of the role of T2 inflamma-tion in COPD [\[50\]](#page-16-13), some promising findings of trials with anti-IL-5/5R α biologics in this clinical setting are noteworthy, with further insights expected from the ongoing MATINEE (NCT04133909) and SUMMER (NCT05138250) on mepolizumab or RESOLUTE (NCT04053634) on benralizumab. A pooled analysis of the METREX and METREO trials suggests that mepolizumab reduces the annual rate of moderate/severe exacerbations by 18% compared to placebo in patients with eosinophilic COPD (\geq 150 cells/ μ L at screening or \geq 300 cells/ μ L in the previous year) [\[174\]](#page-21-16). More disappointing results have been reported to date with benralizumab in phase III GALATEA (NCT02138916) and TERRA-NOVA (NCT02155660) studies involving COPD patients with frequent exacerbations and eosinophil counts ≥220 cells/µL [\[175\]](#page-21-17).

An interesting aspect from the analysis of completed and ongoing trials is that, irrespective of the clinical context in which anti-IL- $5/5R\alpha$ are tested, blood eosinophil counts are essential for patient selection and the prediction of therapeutic response, with a limited role of the other T2 biomarkers [\[165,](#page-21-7)[166\]](#page-21-8). However, while IgE levels are not typically used for guiding anti-IL5/5Rα therapy, it is noteworthy that benralizumab was able to reduce total IgE levels correlated with a reduction in basophils (but not eosinophiles) and with an improvement in asthma control [\[176\]](#page-21-18). Some sporadic but encouraging results have also been observed regarding the potential role of FeNO in predicting the clinical response to anti-IL-5 or anti-IL-5Rα biologics. In a real-world cohort, adults with severe eosinophilic asthma who had baseline FeNO levels \geq 50 ppb experienced a greater reduction in exacerbations after 12 months of these biologics compared to those with FeNO levels < 50 ppb [\[177\]](#page-21-19). However, in a large prospectively designed real-world trial, anti-IL-5 therapies did not appear to significantly reduce FeNO levels after 6 months of treatment, but only eosinophil levels, further confirming their role as the primary biomarker in predicting and monitoring response for these drugs [\[178\]](#page-22-0).

7.3. Anti-IL-4/IL-13

Dupilumab is another monoclonal antibody that targets a component of IL-4R α , shared by both IL-13 and IL-4 [\[179\]](#page-22-1). Based on the results of phase III trials in patients aged 12 years or older [\[180\]](#page-22-2) as well as in the pediatric population [\[181\]](#page-22-3), dupilumab is now authorized as an add-on maintenance treatment for adult and pediatric patients aged \geq 6 years with moderate-to-severe asthma, characterized by an eosinophilic phenotype or steroid resistance. Interestingly, its efficacy in asthma is independent of allergic status, thus suggesting that pretreatment allergy tests may be of limited predictive value [\[182\]](#page-22-4). Similarly, by reducing polyp size, sinus opacification, and symptom severity, dupilumab has demonstrated efficacy and a favorable safety profile in CRSwNP [\[140\]](#page-20-4), leading to its authorization for this condition by major medical agencies. Interestingly, when administered to patients with uncontrolled persistent asthma and comorbid perennial [\[183\]](#page-22-5) or seasonal allergic rhinitis [\[184\]](#page-22-6), dupilumab has shown the capacity to also reduce the rhinitis-associated symptom burden. More recently, there has also been growing interest in evaluating the use of dupilumab in eosinophilic COPD, based on encouraging results from the phase III BOREAS study (NCT03930732) conducted on COPD patients with elevated exacerbation risk despite the use of triple therapy and a blood eosinophil count of at least 300 cells/ μ L [\[185\]](#page-22-7). This multicenter study documented fewer exacerbations, less severe respiratory symptoms, and better lung function as well as quality of life in COPD patients receiving add-on dupilumab subcutaneously every 2 weeks.

In an attempt to adopt a biomarker-driven approach, the analysis of data from the phase III LIBERTY ASTHMA VOYAGE study (NCT02948959) confirmed the key role of circulating blood eosinophils and FeNO in identifying T2 inflammation and predicting the clinical response to dupilumab treatment [\[186\]](#page-22-8). Similar results emerged from a post hoc analysis of the phase III LIBERTY ASTHMA QUEST study (NCT02414854), which showed that an increased baseline FeNO (\geq 25 ppb) in asthma is associated with greater clinical effects of dupilumab versus placebo in terms of $FEV₁$, independently of eosinophil levels and other clinical characteristics [\[187\]](#page-22-9). Another post hoc analysis of the same study confirmed the predictive value of blood eosinophils and FeNO even when employing the GINA threshold of 20 ppb [\[182\]](#page-22-4). Therefore, it is important to emphasize how dupilumab is the only biologic drug for which FeNO is universally accepted as a prognostic and predictive biomarker, which is independent of circulating eosinophiles and, thus, capable of guiding the therapeutic choice [\[187\]](#page-22-9).

7.4. Anti-TSLP

By inhibiting TSLP, tezepelumab disrupts the T2 inflammatory cascade at an upstream point, thus reducing the production and activity of multiple inflammatory mediators. Based on the results of phase IIb PATHWAY (NCT02054130) [\[188\]](#page-22-10) and phase III NAVIGATOR trials (NCT03347279) [\[189\]](#page-22-11), tezepelumab is currently licensed for the treatment of severe asthma that is not adequately controlled despite high doses of ICSs plus another maintenance medication. Moreover, given its mechanism of action, this biologic drug is currently being evaluated in the phase IIa COURSE trial (NCT04039113) for use in moderate to severe COPD with frequent exacerbations despite triple therapy, and in the phase III WAYPOINT trial (NCT04851964) for its use in CRSwNP.

Similar to other biologics that target T2 inflammation, it is generally expected that tezepelumab would demonstrate greater efficacy in patients with a clear T2 signature. Accordingly, it has been shown that tezepelumab impacts all biomarkers of type 2 response, consistently reducing blood eosinophiles, FeNO, and total IgE levels in patients with moderate-to-severe uncontrolled asthma [\[190\]](#page-22-12). However, tezepelumab is not exclusively an anti-eosinophil drug, and it has been demonstrated that its efficacy extends beyond T2-high asthma [\[191\]](#page-22-13). In this regard, the results of the PATHWAY and NAVIGATOR trials show a decrease in the annualized exacerbation rate in asthma patients regardless of whether they have high or low levels of T2 inflammation biomarkers, including blood eosinophils and FeNO, although it should be recognized that patients with elevated levels of these biomarkers experience a more significant clinical improvement [\[191\]](#page-22-13). Accordingly, the pooled analysis of these trials suggests a reduction in exacerbation rates even in a "triple T2 low" biomarker subgroup, characterized by eosinophiles < 150 cells/ μ L, FeNO < 25 ppb, and no perennial allergy [\[191\]](#page-22-13). This should not come as much of a surprise, considering that the release of alarmins by damaged epithelium is not exclusive to T2 inflammation but rather a common defense mechanism in respiratory conditions with diverse pathogenesis [\[45\]](#page-16-8). Given the above, the decision to administer this medication to asthmatic patients is primarily driven by clinical criteria, specifically the inadequate response to steroid therapy, rather than the identification of elevated T2 inflammatory biomarkers.

7.5. Anti-IL-33/ST2

In recent times, drugs directed against IL-33 (itepekimab and tozorakimab) or its ST2 receptor (astegolimab) have held significant promise for the future of CRD treatment, with numerous trials currently registered. Among them, tozorakimab has garnered particular interest due to its dual mechanism of action, potently inhibiting IL-33 signaling on immune cells while also preventing IL-33 oxidation and subsequent epithelial dysfunction mediated by the RAGE/EGFR pathway [\[192\]](#page-22-14). Hence, promising yet unpublished results have emerged from the phase II FRONTIER-3 trial (NCT04570657) with tozorakimab in adult patients with moderate-to-severe asthma. In a phase II study, itepekimab has also demonstrated a lower incidence of exacerbations, while improving lung function in patients with moderate-to-severe asthma [\[193\]](#page-22-15). Similar results have been reported from the phase IIb ZENYATTA trial (NCT02918019), which demonstrated the efficacy of astegolimab in reducing exacerbation rates in both T2-high and T2-low uncontrolled severe asthma [\[194\]](#page-22-16). Some disappointing results have also emerged in patients with moderate-to-very severe COPD, where astegolimab did not significantly reduce the exacerbation rate [\[195\]](#page-22-17). To date, the scientific community is awaiting the results of a number of ongoing trials with anti-alarmins in COPD patients with a history of exacerbations, such as OBEORN (NCT05166889), TITANIA (NCT05158387), and its extension PROSPERO (NCT05742802) for tozorakimab, as well as the three AERIFY trials for itepekimab.

8. Future Perspectives

In the era of precision medicine, CRDs continue to be significant contributors to global morbidity and mortality and, despite much of this burden being preventable or treatable through pharmacological and rehabilitation strategies [\[196,](#page-22-18)[197\]](#page-22-19), these diseases still receive poor attention compared to other non-communicable diseases [\[9\]](#page-14-6). In this scenario, identifying common endotypes for different phenotypic manifestations is progressively paving the way for personalized treatments tailored to distinct immunological profiles of patient clusters, rather than solely addressing specific clinical manifestations [\[17\]](#page-15-5). The frequent coexistence of upper and lower airway diseases with a shared pathogenic mechanism is giving a further impulse to translational research on this matter, and a biomarker-driven approach is becoming crucial for the early detection of individuals who would benefit most from these targeted therapies [\[59\]](#page-16-22). With the growing acknowledgment of T2-driven diseases, the "one airway, one disease" paradigm is gaining traction and promises to change the future of respiratory medicine [\[198\]](#page-22-20). This approach is expected to not only improve patient health outcomes but also reduce future healthcare costs.

Author Contributions: P.A. conceived and designed the study, interpreted results, and drafted the manuscript. G.M., G.R., R.F., C.C. and F.M. draft the manuscript. G.G., E.L. and C.M. performed critical revisions. M.M. conceived and designed the study, supervised the project, drafted the manuscript, and performed critical revisions. All authors have read and agreed to the published version of the manuscript.

Funding: This research was partially supported by the "Ricerca Corrente" funding scheme of the Italian Ministry of Health.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data sharing is not applicable to this article as no new data were created or analyzed in this study.

Acknowledgments: We thank the copyright owner as parts of Figure [1](#page-3-0) were drawn using pictures from Servier Medical Art. By upgrading to CC BY 4.0, Servier Medical Art demonstrates its commitment to promoting collaboration, creativity, and knowledge-sharing within the medical and scientific community. To view the full details of this license, visit <https://creativecommons.org/licenses/by/4.0/> (accessed on 18 July 2024).

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Padron-Monedero, A.; GBD. Global burden of chronic respiratory diseases and risk factors, 1990–2019: An update from the Global Burden of Disease Study 2019. *EClinicalMedicine* **2023**, *59*, 101936. [\[CrossRef\]](https://doi.org/10.1016/j.eclinm.2023.101936)
- 2. Keil, D.C.; Stenzel, N.M.; Kuhl, K.; Vaske, I.; Mewes, R.; Rief, W.; Kenn, K. The impact of chronic obstructive pulmonary disease-related fears on disease-specific disability. *Chron. Respir. Dis.* **2014**, *11*, 31–40. [\[CrossRef\]](https://doi.org/10.1177/1479972313516881) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24431409)
- 3. Eisner, M.D.; Yelin, E.H.; Trupin, L.; Blanc, P.D. The influence of chronic respiratory conditions on health status and work disability. *Am. J. Public. Health* **2002**, *92*, 1506–1513. [\[CrossRef\]](https://doi.org/10.2105/AJPH.92.9.1506) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12197984)
- 4. Djibo, D.A.; Goldstein, J.; Ford, J.G. Prevalence of disability among adults with chronic obstructive pulmonary disease, Behavioral Risk Factor Surveillance System 2016–2017. *PLoS ONE* **2020**, *15*, e0229404. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0229404) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32106254)
- 5. Shi, X.Y.; Ren, Y.; Gu, X.M.; Jia, Y.R.; Wang, X. Impact of pulmonary rehabilitation on patients with different chronic respiratory diseases during hospitalization. *Medicine* **2024**, *103*, e37778. [\[CrossRef\]](https://doi.org/10.1097/MD.0000000000037778) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38608115)
- 6. Rochester, C.L.; Alison, J.A.; Carlin, B.; Jenkins, A.R.; Cox, N.S.; Bauldoff, G.; Bhatt, S.P.; Bourbeau, J.; Burtin, C.; Camp, P.G.; et al. Pulmonary Rehabilitation for Adults with Chronic Respiratory Disease: An Official American Thoracic Society Clinical Practice Guideline. *Am. J. Respir. Crit. Care Med.* **2023**, *208*, e7–e26. [\[CrossRef\]](https://doi.org/10.1164/rccm.202306-1066ST) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37581410)
- 7. Jenkins, A.R.; Cox, N.S.; Tappan, R.S.; Holland, A.E.; Rochester, C.L.; Alison, J.A.; Carlin, B.; Ruminjo, J.K.; Thomson, C.C. Summary for Clinicians: Clinical Practice Guideline on Pulmonary Rehabilitation for Adults with Chronic Respiratory Disease. *Ann. Am. Thorac. Soc.* **2024**, *21*, 533–537. [\[CrossRef\]](https://doi.org/10.1513/AnnalsATS.202310-909CME) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38241016)
- 8. Labaki, W.W.; Han, M.K. Chronic respiratory diseases: A global view. *Lancet Respir. Med.* **2020**, *8*, 531–533. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(20)30157-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32526184)
- 9. Collaborators, G.B.D.C.R.D. Prevalence and attributable health burden of chronic respiratory diseases, 1990–2017: A systematic analysis for the Global Burden of Disease Study 2017. *Lancet Respir. Med.* **2020**, *8*, 585–596. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(20)30105-3)
- 10. World Health Organization (WHO). Chronic Respiratory Diseases. Available online: [https://www.who.int/health-topics/](https://www.who.int/health-topics/chronic-respiratory-diseases#tab=tab_1) [chronic-respiratory-diseases#tab=tab_1](https://www.who.int/health-topics/chronic-respiratory-diseases#tab=tab_1) (accessed on 7 July 2024).
- 11. Dominguez-Ortega, J.; Mullol, J.; Alvarez Gutierrez, F.J.; Miguel-Blanco, C.; Castillo, J.A.; Olaguibel, J.M.; Blanco-Aparicio, M. The effect of biologics in lung function and quality of life of patients with united airways disease: A systematic review. *J. Allergy Clin. Immunol. Glob.* **2024**, *3*, 100174. [\[CrossRef\]](https://doi.org/10.1016/j.jacig.2023.100174)
- 12. Braunstahl, G.J. United airways concept: What does it teach us about systemic inflammation in airways disease? *Proc. Am. Thorac. Soc.* **2009**, *6*, 652–654. [\[CrossRef\]](https://doi.org/10.1513/pats.200906-052DP) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20008869)
- 13. Yii, A.C.A.; Tay, T.R.; Choo, X.N.; Koh, M.S.Y.; Tee, A.K.H.; Wang, D.Y. Precision medicine in united airways disease: A "treatable traits" approach. *Allergy* **2018**, *73*, 1964–1978. [\[CrossRef\]](https://doi.org/10.1111/all.13496)
- 14. Anderson, G.P. Endotyping asthma: New insights into key pathogenic mechanisms in a complex, heterogeneous disease. *Lancet* **2008**, *372*, 1107–1119. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(08)61452-X)
- 15. Hizawa, N. The understanding of asthma pathogenesis in the era of precision medicine. *Allergol. Int.* **2023**, *72*, 3–10. [\[CrossRef\]](https://doi.org/10.1016/j.alit.2022.09.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36195530)
- 16. Vatrella, A.; Maglio, A.; Pelaia, C.; Ciampo, L.; Pelaia, G.; Vitale, C. Eosinophilic inflammation: An Appealing Target for Pharmacologic Treatments in Severe Asthma. *Biomedicines* **2022**, *10*, 2181. [\[CrossRef\]](https://doi.org/10.3390/biomedicines10092181)
- 17. Agusti, A.; Bel, E.; Thomas, M.; Vogelmeier, C.; Brusselle, G.; Holgate, S.; Humbert, M.; Jones, P.; Gibson, P.G.; Vestbo, J.; et al. Treatable traits: Toward precision medicine of chronic airway diseases. *Eur. Respir. J.* **2016**, *47*, 410–419. [\[CrossRef\]](https://doi.org/10.1183/13993003.01359-2015)
- 18. Cho, H.J.; Ha, J.G.; Lee, S.N.; Kim, C.H.; Wang, D.Y.; Yoon, J.H. Differences and similarities between the upper and lower airway: Focusing on innate immunity. *Rhinology* **2021**, *59*, 441–450. [\[CrossRef\]](https://doi.org/10.4193/Rhin21.046)
- 19. Giavina-Bianchi, P.; Aun, M.V.; Takejima, P.; Kalil, J.; Agondi, R.C. United airway disease: Current perspectives. *J. Asthma Allergy* **2016**, *9*, 93–100. [\[CrossRef\]](https://doi.org/10.2147/JAA.S81541) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27257389)
- 20. Brozek, J.L.; Bousquet, J.; Agache, I.; Agarwal, A.; Bachert, C.; Bosnic-Anticevich, S.; Brignardello-Petersen, R.; Canonica, G.W.; Casale, T.; Chavannes, N.H.; et al. Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines-2016 revision. *J. Allergy Clin. Immunol.* **2017**, *140*, 950–958. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2017.03.050)
- 21. Leynaert, B.; Bousquet, J.; Neukirch, C.; Liard, R.; Neukirch, F. Perennial rhinitis: An independent risk factor for asthma in nonatopic subjects: Results from the European Community Respiratory Health Survey. *J. Allergy Clin. Immunol.* **1999**, *104*, 301–304. [\[CrossRef\]](https://doi.org/10.1016/S0091-6749(99)70370-2)
- 22. Hakansson, K.; Bachert, C.; Konge, L.; Thomsen, S.F.; Pedersen, A.E.; Poulsen, S.S.; Martin-Bertelsen, T.; Winther, O.; Backer, V.; von Buchwald, C. Airway Inflammation in Chronic Rhinosinusitis with Nasal Polyps and Asthma: The United Airways Concept Further Supported. *PLoS ONE* **2015**, *10*, e0127228. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0127228) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26132710)
- 23. Coumou, H.; Westerhof, G.A.; de Nijs, S.B.; Zwinderman, A.H.; Bel, E.H. Predictors of accelerated decline in lung function in adult-onset asthma. *Eur. Respir. J.* **2018**, *51*, 1701785. [\[CrossRef\]](https://doi.org/10.1183/13993003.01785-2017) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29444915)
- 24. Widal, F.; Abrami, P.; Lermoyez, J. First complete description of the aspirin idiosyncrasy-asthma-nasal polyposis syndrome (plus urticaria)--1922 (with a note on aspirin desensitization). By F. Widal, P. Abrami, J. Lermoyez. *J. Asthma* **1987**, *24*, 297–300. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3327855)
- 25. Samter, M.; Beers, R.F., Jr. Intolerance to aspirin. Clinical studies and consideration of its pathogenesis. *Ann. Intern. Med.* **1968**, *68*, 975–983. [\[CrossRef\]](https://doi.org/10.7326/0003-4819-68-5-975) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/5646829)
- 26. Stevens, W.W.; Ocampo, C.J.; Berdnikovs, S.; Sakashita, M.; Mahdavinia, M.; Suh, L.; Takabayashi, T.; Norton, J.E.; Hulse, K.E.; Conley, D.B.; et al. Cytokines in Chronic Rhinosinusitis. Role in Eosinophilia and Aspirin-exacerbated Respiratory Disease. *Am. J. Respir. Crit. Care Med.* **2015**, *192*, 682–694. [\[CrossRef\]](https://doi.org/10.1164/rccm.201412-2278OC) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26067893)
- 27. Liu, T.; Barrett, N.A.; Kanaoka, Y.; Yoshimoto, E.; Garofalo, D.; Cirka, H.; Feng, C.; Boyce, J.A. Type 2 Cysteinyl Leukotriene Receptors Drive IL-33-Dependent Type 2 Immunopathology and Aspirin Sensitivity. *J. Immunol.* **2018**, *200*, 915–927. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.1700603) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29282304)
- 28. Bachert, C.; Luong, A.U.; Gevaert, P.; Mullol, J.; Smith, S.G.; Silver, J.; Sousa, A.R.; Howarth, P.H.; Benson, V.S.; Mayer, B.; et al. The Unified Airway Hypothesis: Evidence from Specific Intervention with Anti-IL-5 Biologic Therapy. *J. Allergy Clin. Immunol. Pract.* **2023**, *11*, 2630–2641. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2023.05.011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37207831)
- 29. Tiotiu, A.; Novakova, P.; Baiardini, I.; Bikov, A.; Chong-Neto, H.; de-Sousa, J.C.; Emelyanov, A.; Heffler, E.; Fogelbach, G.G.; Kowal, K.; et al. Manifesto on united airways diseases (UAD): An Interasma (global asthma association—GAA) document. *J. Asthma* **2022**, *59*, 639–654. [\[CrossRef\]](https://doi.org/10.1080/02770903.2021.1879130)
- 30. Kuruvilla, M.E.; Lee, F.E.; Lee, G.B. Understanding Asthma Phenotypes, Endotypes, and Mechanisms of Disease. *Clin. Rev. Allergy Immunol.* **2019**, *56*, 219–233. [\[CrossRef\]](https://doi.org/10.1007/s12016-018-8712-1)
- 31. Luo, W.; Hu, J.; Xu, W.; Dong, J. Distinct spatial and temporal roles for Th1, Th2, and Th17 cells in asthma. *Front. Immunol.* **2022**, *13*, 974066. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2022.974066)
- 32. Harrington, L.E.; Mangan, P.R.; Weaver, C.T. Expanding the effector CD4 T-cell repertoire: The Th17 lineage. *Curr. Opin. Immunol.* **2006**, *18*, 349–356. [\[CrossRef\]](https://doi.org/10.1016/j.coi.2006.03.017)
- 33. Kyriakopoulos, C.; Gogali, A.; Bartziokas, K.; Kostikas, K. Identification and treatment of T2-low asthma in the era of biologics. *ERJ Open Res.* **2021**, *7*. [\[CrossRef\]](https://doi.org/10.1183/23120541.00309-2020)
- 34. Ham, J.; Kim, J.; Ko, Y.G.; Kim, H.Y. The Dynamic Contribution of Neutrophils in the Chronic Respiratory Diseases. *Allergy Asthma Immunol. Res.* **2022**, *14*, 361–378. [\[CrossRef\]](https://doi.org/10.4168/aair.2022.14.4.361) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35837821)
- 35. Eyerich, K.; Dimartino, V.; Cavani, A. IL-17 and IL-22 in immunity: Driving protection and pathology. *Eur. J. Immunol.* **2017**, *47*, 607–614. [\[CrossRef\]](https://doi.org/10.1002/eji.201646723)
- 36. Wang, Y.H.; Voo, K.S.; Liu, B.; Chen, C.Y.; Uygungil, B.; Spoede, W.; Bernstein, J.A.; Huston, D.P.; Liu, Y.J. A novel subset of CD4(+) T(H)2 memory/effector cells that produce inflammatory IL-17 cytokine and promote the exacerbation of chronic allergic asthma. *J. Exp. Med.* **2010**, *207*, 2479–2491. [\[CrossRef\]](https://doi.org/10.1084/jem.20101376)
- 37. Irvin, C.; Zafar, I.; Good, J.; Rollins, D.; Christianson, C.; Gorska, M.M.; Martin, R.J.; Alam, R. Increased frequency of dual-positive TH2/TH17 cells in bronchoalveolar lavage fluid characterizes a population of patients with severe asthma. *J. Allergy Clin. Immunol.* **2014**, *134*, 1175–1186.e7. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2014.05.038) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25042748)
- 38. Kokubo, K.; Onodera, A.; Kiuchi, M.; Tsuji, K.; Hirahara, K.; Nakayama, T. Conventional and pathogenic Th2 cells in inflammation, tissue repair, and fibrosis. *Front. Immunol.* **2022**, *13*, 945063. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2022.945063) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36016937)
- 39. Punnonen, J.; Aversa, G.; Cocks, B.G.; McKenzie, A.N.; Menon, S.; Zurawski, G.; de Waal Malefyt, R.; de Vries, J.E. Interleukin 13 induces interleukin 4-independent IgG4 and IgE synthesis and CD23 expression by human B cells. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 3730–3734. [\[CrossRef\]](https://doi.org/10.1073/pnas.90.8.3730)
- 40. Gigon, L.; Fettrelet, T.; Yousefi, S.; Simon, D.; Simon, H.U. Eosinophils from A to Z. *Allergy* **2023**, *78*, 1810–1846. [\[CrossRef\]](https://doi.org/10.1111/all.15751)
- 41. Wechsler, M.E.; Munitz, A.; Ackerman, S.J.; Drake, M.G.; Jackson, D.J.; Wardlaw, A.J.; Dougan, S.K.; Berdnikovs, S.; Schleich, F.;
- Matucci, A.; et al. Eosinophils in Health and Disease: A State-of-the-Art Review. *Mayo Clin. Proc.* **2021**, *96*, 2694–2707. [\[CrossRef\]](https://doi.org/10.1016/j.mayocp.2021.04.025) 42. Spencer, L.A.; Bonjour, K.; Melo, R.C.; Weller, P.F. Eosinophil secretion of granule-derived cytokines. *Front. Immunol.* **2014**, *5*, 496. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2014.00496)
- 43. Weller, P.F.; Spencer, L.A. Functions of tissue-resident eosinophils. *Nat. Rev. Immunol.* **2017**, *17*, 746–760. [\[CrossRef\]](https://doi.org/10.1038/nri.2017.95)
- 44. Jin, J.; Sunusi, S.; Lu, H. Group 2 innate lymphoid cells (ILC2s) are important in typical type 2 immune-mediated diseases and an essential therapeutic target. *J. Int. Med. Res.* **2022**, *50*, 3000605211053156. [\[CrossRef\]](https://doi.org/10.1177/03000605211053156) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35048721)
- 45. Stanbery, A.G.; Shuchi, S.; von Moltke, J.; Tait Wojno, E.D.; Ziegler, S.F. TSLP, IL-33, and IL-25: Not just for allergy and helminth infection. *J. Allergy Clin. Immunol.* **2022**, *150*, 1302–1313. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2022.07.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35863509)
- 46. Morita, H.; Moro, K.; Koyasu, S. Innate lymphoid cells in allergic and nonallergic inflammation. *J. Allergy Clin. Immunol.* **2016**, *138*, 1253–1264. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2016.09.011)
- 47. Maison, N.; Omony, J.; Illi, S.; Thiele, D.; Skevaki, C.; Dittrich, A.M.; Bahmer, T.; Rabe, K.F.; Weckmann, M.; Happle, C.; et al. T2-high asthma phenotypes across lifespan. *Eur. Respir. J.* **2022**, *60*, 2102288. [\[CrossRef\]](https://doi.org/10.1183/13993003.02288-2021)
- 48. Rupani, H.; Kyyaly, M.A.; Azim, A.; Abadalkareen, R.; Freeman, A.; Dennison, P.; Howarth, P.; Djukanovic, R.; Vijayanand, P.; Seumois, G.; et al. Comprehensive Characterization of Difficult-to-Treat Asthma Reveals Near Absence of T2-Low Status. *J. Allergy Clin. Immunol. Pract.* **2023**, *11*, 2812–2821.e4. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2023.05.028) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37245729)
- 49. Ricciardolo, F.L.M.; Sprio, A.E.; Baroso, A.; Gallo, F.; Riccardi, E.; Bertolini, F.; Carriero, V.; Arrigo, E.; Ciprandi, G. Characterization of T2-Low and T2-High Asthma Phenotypes in Real-Life. *Biomedicines* **2021**, *9*, 1684. [\[CrossRef\]](https://doi.org/10.3390/biomedicines9111684) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34829913)
- 50. Singh, D.; Kolsum, U.; Brightling, C.E.; Locantore, N.; Agusti, A.; Tal-Singer, R.; on behalf of the ECLIPSE investigators. Eosinophilic inflammation in COPD: Prevalence and clinical characteristics. *Eur. Respir. J.* **2014**, *44*, 1697–1700. [\[CrossRef\]](https://doi.org/10.1183/09031936.00162414)
- 51. Vedel-Krogh, S.; Nielsen, S.F.; Lange, P.; Vestbo, J.; Nordestgaard, B.G. Blood Eosinophils and Exacerbations in Chronic Obstructive Pulmonary Disease. The Copenhagen General Population Study. *Am. J. Respir. Crit. Care Med.* **2016**, *193*, 965–974. [\[CrossRef\]](https://doi.org/10.1164/rccm.201509-1869OC)
- 52. Ho, J.; He, W.; Chan, M.T.V.; Tse, G.; Liu, T.; Wong, S.H.; Leung, C.C.H.; Wong, W.T.; Tsang, S.; Zhang, L.; et al. Eosinophilia and clinical outcome of chronic obstructive pulmonary disease: A meta-analysis. *Sci. Rep.* **2017**, *7*, 13451. [\[CrossRef\]](https://doi.org/10.1038/s41598-017-13745-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29044160)
- 53. Maspero, J.; Adir, Y.; Al-Ahmad, M.; Celis-Preciado, C.A.; Colodenco, F.D.; Giavina-Bianchi, P.; Lababidi, H.; Ledanois, O.; Mahoub, B.; Perng, D.W.; et al. Type 2 inflammation in asthma and other airway diseases. *ERJ Open Res.* **2022**, *8*. [\[CrossRef\]](https://doi.org/10.1183/23120541.00576-2021) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35923421)
- 54. Tomassen, P.; Vandeplas, G.; Van Zele, T.; Cardell, L.O.; Arebro, J.; Olze, H.; Forster-Ruhrmann, U.; Kowalski, M.L.; Olszewska-Ziaber, A.; Holtappels, G.; et al. Inflammatory endotypes of chronic rhinosinusitis based on cluster analysis of biomarkers. *J. Allergy Clin. Immunol.* **2016**, *137*, 1449–1456.e4. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2015.12.1324) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26949058)
- 55. Gelardi, M.; Maselli del Giudice, A.; Fiorella, M.L.; Fiorella, R.; Russo, C.; Soleti, P.; Di Gioacchino, M.; Ciprandi, G. Non-allergic rhinitis with eosinophils and mast cells constitutes a new severe nasal disorder. *Int. J. Immunopathol. Pharmacol.* **2008**, *21*, 325–331. [\[CrossRef\]](https://doi.org/10.1177/039463200802100209) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18547476)
- 56. Ellis, A.K.; Keith, P.K. Nonallergic rhinitis with eosinophilia syndrome. *Curr. Allergy Asthma Rep.* **2006**, *6*, 215–220. [\[CrossRef\]](https://doi.org/10.1007/s11882-006-0037-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16579871)
- 57. De Corso, E.; Seccia, V.; Ottaviano, G.; Cantone, E.; Lucidi, D.; Settimi, S.; Di Cesare, T.; Galli, J. Clinical Evidence of Type 2 Inflammation in Non-allergic Rhinitis with Eosinophilia Syndrome: A Systematic Review. *Curr. Allergy Asthma Rep.* **2022**, *22*, 29–42. [\[CrossRef\]](https://doi.org/10.1007/s11882-022-01027-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35141844)
- 58. Delemarre, T.; Holtappels, G.; De Ruyck, N.; Zhang, N.; Nauwynck, H.; Bachert, C.; Gevaert, E. Type 2 inflammation in chronic rhinosinusitis without nasal polyps: Another relevant endotype. *J. Allergy Clin. Immunol.* **2020**, *146*, 337–343.e6. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2020.04.040)
- 59. Pavord, I.D. Blood Eosinophil-directed Management of Airway Disease. The Past, Present, and Future. *Am. J. Respir. Crit. Care Med.* **2020**, *202*, 637–639. [\[CrossRef\]](https://doi.org/10.1164/rccm.202004-1013ED) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32356667)
- 60. Wu, A.C.; Kiley, J.P.; Noel, P.J.; Amur, S.; Burchard, E.G.; Clancy, J.P.; Galanter, J.; Inada, M.; Jones, T.K.; Kropski, J.A.; et al. Current Status and Future Opportunities in Lung Precision Medicine Research with a Focus on Biomarkers. An American Thoracic Society/National Heart, Lung, and Blood Institute Research Statement. *Am. J. Respir. Crit. Care Med.* **2018**, *198*, e116–e136. [\[CrossRef\]](https://doi.org/10.1164/rccm.201810-1895ST)
- 61. Garcia-Marcos, L.; Brand, P.L. The utility of sputum eosinophils and exhaled nitric oxide for monitoring asthma control with special attention to childhood asthma. *Allergol. Immunopathol.* **2010**, *38*, 41–46. [\[CrossRef\]](https://doi.org/10.1016/j.aller.2009.10.006)
- 62. Wang, S.A. The Diagnostic Work-Up of Hypereosinophilia. *Pathobiology* **2019**, *86*, 39–52. [\[CrossRef\]](https://doi.org/10.1159/000489341)
- 63. Singh, D.; Agusti, A.; Martinez, F.J.; Papi, A.; Pavord, I.D.; Wedzicha, J.A.; Vogelmeier, C.F.; Halpin, D.M.G. Blood Eosinophils and Chronic Obstructive Pulmonary Disease: A Global Initiative for Chronic Obstructive Lung Disease Science Committee 2022 Review. *Am. J. Respir. Crit. Care Med.* **2022**, *206*, 17–24. [\[CrossRef\]](https://doi.org/10.1164/rccm.202201-0209PP)
- 64. Pignatti, P.; Visca, D.; Lucini, E.; Cherubino, F.; Zampogna, E.; Celesia, C.; Sotgiu, G.; Spanevello, A. Correlation between sputum and blood eosinophils in asthmatic and COPD patients with comorbidities. *Eur. Respir. J.* **2018**, *52*, PA4418. [\[CrossRef\]](https://doi.org/10.1183/13993003.congress-2018.PA4418)
- 65. Pignatti, P.; Visca, D.; Cherubino, F.; Zampogna, E.; Lucini, E.; Saderi, L.; Sotgiu, G.; Spanevello, A. Do blood eosinophils strictly reflect airway inflammation in COPD? Comparison with asthmatic patients. *Respir. Res.* **2019**, *20*, 145. [\[CrossRef\]](https://doi.org/10.1186/s12931-019-1111-1)
- 66. Wagener, A.H.; de Nijs, S.B.; Lutter, R.; Sousa, A.R.; Weersink, E.J.; Bel, E.H.; Sterk, P.J. External validation of blood eosinophils, FE(NO) and serum periostin as surrogates for sputum eosinophils in asthma. *Thorax* **2015**, *70*, 115–120. [\[CrossRef\]](https://doi.org/10.1136/thoraxjnl-2014-205634)
- 67. Mycroft, K.; Paplinska-Goryca, M.; Proboszcz, M.; Nejman-Gryz, P.; Krenke, R.; Gorska, K. Blood and Sputum Eosinophils of COPD Patients Are Differently Polarized than in Asthma. *Cells* **2023**, *12*, 1631. [\[CrossRef\]](https://doi.org/10.3390/cells12121631) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37371101)
- 68. Korevaar, D.A.; Westerhof, G.A.; Wang, J.; Cohen, J.F.; Spijker, R.; Sterk, P.J.; Bel, E.H.; Bossuyt, P.M. Diagnostic accuracy of minimally invasive markers for detection of airway eosinophilia in asthma: A systematic review and meta-analysis. *Lancet Respir. Med.* **2015**, *3*, 290–300. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(15)00050-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25801413)
- 69. Saetta, M.; Di Stefano, A.; Maestrelli, P.; Turato, G.; Ruggieri, M.P.; Roggeri, A.; Calcagni, P.; Mapp, C.E.; Ciaccia, A.; Fabbri, L.M. Airway eosinophilia in chronic bronchitis during exacerbations. *Am. J. Respir. Crit. Care Med.* **1994**, *150*, 1646–1652. [\[CrossRef\]](https://doi.org/10.1164/ajrccm.150.6.7952628) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7952628)
- 70. Turato, G.; Semenzato, U.; Bazzan, E.; Biondini, D.; Tine, M.; Torrecilla, N.; Forner, M.; Marin, J.M.; Cosio, M.G.; Saetta, M. Blood Eosinophilia Neither Reflects Tissue Eosinophils nor Worsens Clinical Outcomes in Chronic Obstructive Pulmonary Disease. *Am. J. Respir. Crit. Care Med.* **2018**, *197*, 1216–1219. [\[CrossRef\]](https://doi.org/10.1164/rccm.201708-1684LE)
- 71. Prins, H.J.; Duijkers, R.; Lutter, R.; Daniels, J.M.; van der Valk, P.; Schoorl, M.; Kerstjens, H.A.; van der Werf, T.S.; Boersma, W.G. Blood eosinophilia as a marker of early and late treatment failure in severe acute exacerbations of COPD. *Respir. Med.* **2017**, *131*, 118–124. [\[CrossRef\]](https://doi.org/10.1016/j.rmed.2017.07.064)
- 72. Hastie, A.T.; Martinez, F.J.; Curtis, J.L.; Doerschuk, C.M.; Hansel, N.N.; Christenson, S.; Putcha, N.; Ortega, V.E.; Li, X.; Barr, R.G.; et al. Association of sputum and blood eosinophil concentrations with clinical measures of COPD severity: An analysis of the SPIROMICS cohort. *Lancet Respir. Med.* **2017**, *5*, 956–967. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(17)30432-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29146301)
- 73. Global Initiative for Chronic Obstructive Lung Disease. 2024 GOLD Report. Available online: [https://goldcopd.org/2024-gold](https://goldcopd.org/2024-gold-report/)[report/](https://goldcopd.org/2024-gold-report/) (accessed on 6 June 2024).
- 74. Mathur, S.K.; Fichtinger, P.S.; Evans, M.D.; Schwantes, E.A.; Jarjour, N.N. Variability of blood eosinophil count as an asthma biomarker. *Ann. Allergy Asthma Immunol.* **2016**, *117*, 551–553. [\[CrossRef\]](https://doi.org/10.1016/j.anai.2016.08.010)
- 75. Chipps, B.E.; Jarjour, N.; Calhoun, W.J.; Iqbal, A.; Haselkorn, T.; Yang, M.; Brumm, J.; Corren, J.; Holweg, C.T.J.; Bafadhel, M. A Comprehensive Analysis of the Stability of Blood Eosinophil Levels. *Ann. Am. Thorac. Soc.* **2021**, *18*, 1978–1987. [\[CrossRef\]](https://doi.org/10.1513/AnnalsATS.202010-1249OC)
- 76. Southworth, T.; Beech, G.; Foden, P.; Kolsum, U.; Singh, D. The reproducibility of COPD blood eosinophil counts. *Eur. Respir. J.* **2018**, *52*, 1800427. [\[CrossRef\]](https://doi.org/10.1183/13993003.00427-2018)
- 77. Green, R.H.; Brightling, C.E.; McKenna, S.; Hargadon, B.; Parker, D.; Bradding, P.; Wardlaw, A.J.; Pavord, I.D. Asthma exacerbations and sputum eosinophil counts: A randomised controlled trial. *Lancet* **2002**, *360*, 1715–1721. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(02)11679-5)
- 78. Lim, S.; Jatakanon, A.; John, M.; Gilbey, T.; O'Connor, B.J.; Chung, K.F.; Barnes, P.J. Effect of inhaled budesonide on lung function and airway inflammation. Assessment by various inflammatory markers in mild asthma. *Am. J. Respir. Crit. Care Med.* **1999**, *159*, 22–30. [\[CrossRef\]](https://doi.org/10.1164/ajrccm.159.1.9706006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9872813)
- 79. Siva, R.; Green, R.H.; Brightling, C.E.; Shelley, M.; Hargadon, B.; McKenna, S.; Monteiro, W.; Berry, M.; Parker, D.; Wardlaw, A.J.; et al. Eosinophilic airway inflammation and exacerbations of COPD: A randomised controlled trial. *Eur. Respir. J.* **2007**, *29*, 906–913. [\[CrossRef\]](https://doi.org/10.1183/09031936.00146306)
- 80. Brightling, C.E.; Monteiro, W.; Ward, R.; Parker, D.; Morgan, M.D.; Wardlaw, A.J.; Pavord, I.D. Sputum eosinophilia and short-term response to prednisolone in chronic obstructive pulmonary disease: A randomised controlled trial. *Lancet* **2000**, *356*, 1480–1485. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(00)02872-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11081531)
- 81. Mathioudakis, A.G.; Bikov, A.; Foden, P.; Lahousse, L.; Brusselle, G.; Singh, D.; Vestbo, J. Change in blood eosinophils following treatment with inhaled corticosteroids may predict long-term clinical response in COPD. *Eur. Respir. J.* **2020**, *55*, 1902119. [\[CrossRef\]](https://doi.org/10.1183/13993003.02119-2019)
- 82. Siddiqui, S.; Bachert, C.; Bjermer, L.; Buchheit, K.M.; Castro, M.; Qin, Y.; Rupani, H.; Sagara, H.; Howarth, P.; Taille, C. Eosinophils and tissue remodeling: Relevance to airway disease. *J. Allergy Clin. Immunol.* **2023**, *152*, 841–857. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2023.06.005)
- 83. Gelardi, M.; Giancaspro, R.; Cintoli, G.; Minafra, S.S.; Musto, S.; Cassano, M. Proposal for a new diagnostic-therapeutic algorithm in chronic rhinosinusitis with nasal polyps. *Acta Biomed.* **2023**, *94*, e2023218. [\[CrossRef\]](https://doi.org/10.23750/abm.v94i4.14152)
- 84. Mottais, A.; Riberi, L.; Falco, A.; Soccal, S.; Gohy, S.; De Rose, V. Epithelial-Mesenchymal Transition Mechanisms in Chronic Airway Diseases: A Common Process to Target? *Int. J. Mol. Sci.* **2023**, *24*, 12412. [\[CrossRef\]](https://doi.org/10.3390/ijms241512412) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37569787)
- 85. Heijink, I.H.; Kuchibhotla, V.N.S.; Roffel, M.P.; Maes, T.; Knight, D.A.; Sayers, I.; Nawijn, M.C. Epithelial cell dysfunction, a major driver of asthma development. *Allergy* **2020**, *75*, 1902–1917. [\[CrossRef\]](https://doi.org/10.1111/all.14421) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32460363)
- 86. Amorim, M.M.; Araruna, A.; Caetano, L.B.; Cruz, A.C.; Santoro, L.L.; Fernandes, A.L. Nasal eosinophilia: An indicator of eosinophilic inflammation in asthma. *Clin. Exp. Allergy* **2010**, *40*, 867–874. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2222.2009.03439.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20100189)
- 87. Ciprandi, G.; Varricchio, A.; Tajana, G.; La Mantia, I.; Tommasino, C. Peripheral Eosinophil Counts Correlate with Nasal Eosinophil Counts in Patients with Rhinitis. *J. Investig. Allergol. Clin. Immunol.* **2018**, *28*, 428–430. [\[CrossRef\]](https://doi.org/10.18176/jiaci.0306) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30530393)
- 88. de Farias, C.F.; Amorim, M.M.; Dracoulakis, M.; Caetano, L.B.; Santoro, I.L.; Fernandes, A.L. Nasal lavage, blood or sputum: Which is best for phenotyping asthma? *Respirology* **2017**, *22*, 671–677. [\[CrossRef\]](https://doi.org/10.1111/resp.12958) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27899011)
- 89. Ambrosino, P.; Marcuccio, G.; Manzo, F.; Mancusi, C.; Merola, C.; Maniscalco, M. The Clinical Application of Established and Emerging Biomarkers for Chronic Respiratory Diseases. *J. Clin. Med.* **2023**, *12*, 6125. [\[CrossRef\]](https://doi.org/10.3390/jcm12196125)
- 90. Schneider, A.; Brunn, B.; Hapfelmeier, A.; Schultz, K.; Kellerer, C.; Jorres, R.A. Diagnostic accuracy of FeNO in asthma and predictive value for inhaled corticosteroid responsiveness: A prospective, multicentre study. *EClinicalMedicine* **2022**, *50*, 101533. [\[CrossRef\]](https://doi.org/10.1016/j.eclinm.2022.101533)
- 91. Khatri, S.B.; Iaccarino, J.M.; Barochia, A.; Soghier, I.; Akuthota, P.; Brady, A.; Covar, R.A.; Debley, J.S.; Diamant, Z.; Fitzpatrick, A.M.; et al. Use of Fractional Exhaled Nitric Oxide to Guide the Treatment of Asthma: An Official American Thoracic Society Clinical Practice Guideline. *Am. J. Respir. Crit. Care Med.* **2021**, *204*, e97–e109. [\[CrossRef\]](https://doi.org/10.1164/rccm.202109-2093ST)
- 92. Price, D.B.; Bosnic-Anticevich, S.; Pavord, I.D.; Roche, N.; Halpin, D.M.G.; Bjermer, L.; Usmani, O.S.; Brusselle, G.; Ming, S.W.Y.; Rastogi, S. Association of elevated fractional exhaled nitric oxide concentration and blood eosinophil count with severe asthma exacerbations. *Clin. Transl. Allergy* **2019**, *9*, 41. [\[CrossRef\]](https://doi.org/10.1186/s13601-019-0282-7)
- 93. Simpson, A.; Custovic, A.; Pipis, S.; Adisesh, A.; Faragher, B.; Woodcock, A. Exhaled nitric oxide, sensitization, and exposure to allergens in patients with asthma who are not taking inhaled steroids. *Am. J. Respir. Crit. Care Med.* **1999**, *160*, 45–49. [\[CrossRef\]](https://doi.org/10.1164/ajrccm.160.1.9809091) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10390378)
- 94. Scott, M.; Raza, A.; Karmaus, W.; Mitchell, F.; Grundy, J.; Kurukulaaratchy, R.J.; Arshad, S.H.; Roberts, G. Influence of atopy and asthma on exhaled nitric oxide in an unselected birth cohort study. *Thorax* **2010**, *65*, 258–262. [\[CrossRef\]](https://doi.org/10.1136/thx.2009.125443) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20335297)
- 95. Duarte, R.L.M.; Rabahi, M.F.; Oliveira, E.S.T.S.; Magalhaes-da-Silveira, F.J.; Mello, F.C.Q.; Gozal, D. Fractional Exhaled Nitric Oxide Measurements and Screening of Obstructive Sleep Apnea in a Sleep-Laboratory Setting: A Cross-Sectional Study. *Lung* **2019**, *197*, 131–137. [\[CrossRef\]](https://doi.org/10.1007/s00408-018-0190-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30600394)
- 96. Liu, D.; Huang, Z.; Huang, Y.; Yi, X.; Chen, X. Measurement of nasal and fractional exhaled nitric oxide in children with upper airway inflammatory disease: Preliminary results. *Int. J. Pediatr. Otorhinolaryngol.* **2015**, *79*, 2308–2311. [\[CrossRef\]](https://doi.org/10.1016/j.ijporl.2015.10.033) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26602553)
- 97. Park, J.A.; Cha, H.; Yang, S.K.; Ryu, H.T.; Kim, D.W.; Hong, S.N.; Yang, M.S.; Kim, D.W. The Role of Fractional Exhaled Nitric Oxide in Diagnosing Asthmatic Type 2 Chronic Rhinosinusitis with Nasal Polyps. *Am. J. Rhinol. Allergy* **2023**, *37*, 524–530. [\[CrossRef\]](https://doi.org/10.1177/19458924231173205)
- 98. Dweik, R.A.; Boggs, P.B.; Erzurum, S.C.; Irvin, C.G.; Leigh, M.W.; Lundberg, J.O.; Olin, A.C.; Plummer, A.L.; Taylor, D.R.; American Thoracic Society Committee on Interpretation of Exhaled Nitric Oxide Levels (FENO) for Clinical Applications. An official ATS clinical practice guideline: Interpretation of exhaled nitric oxide levels (FENO) for clinical applications. *Am. J. Respir. Crit. Care Med.* **2011**, *184*, 602–615. [\[CrossRef\]](https://doi.org/10.1164/rccm.9120-11ST)
- 99. Dressel, H.; de la Motte, D.; Reichert, J.; Ochmann, U.; Petru, R.; Angerer, P.; Holz, O.; Nowak, D.; Jorres, R.A. Exhaled nitric oxide: Independent effects of atopy, smoking, respiratory tract infection, gender and height. *Respir. Med.* **2008**, *102*, 962–969. [\[CrossRef\]](https://doi.org/10.1016/j.rmed.2008.02.012)
- 100. Olin, A.C.; Bake, B.; Toren, K. Fraction of exhaled nitric oxide at 50 mL/s: Reference values for adult lifelong never-smokers. *Chest* **2007**, *131*, 1852–1856. [\[CrossRef\]](https://doi.org/10.1378/chest.06-2928)
- 101. Comberiati, P.; Peroni, D.; Malka-Rais, J.; Morganti, R.; Spahn, J.D. Fractional exhaled nitric oxide response to oral corticosteroids in children with mild-to-moderate asthma: Influence of race. *Ann. Allergy Asthma Immunol.* **2020**, *125*, 440–446.e1. [\[CrossRef\]](https://doi.org/10.1016/j.anai.2020.06.036)
- 102. Lim, C.S.; Rani, F.A.; Tan, L.E. Response of exhaled nitric oxide to inhaled corticosteroids in patients with stable COPD: A systematic review and meta-analysis. *Clin. Respir. J.* **2018**, *12*, 218–226. [\[CrossRef\]](https://doi.org/10.1111/crj.12518)
- 103. Busby, J.; Khoo, E.; Pfeffer, P.E.; Mansur, A.H.; Heaney, L.G. The effects of oral corticosteroids on lung function, type-2 biomarkers and patient-reported outcomes in stable asthma: A systematic review and meta-analysis. *Respir. Med.* **2020**, *173*, 106156. [\[CrossRef\]](https://doi.org/10.1016/j.rmed.2020.106156) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32979621)
- 104. Schumann, D.M.; Papakonstantinou, E.; Kostikas, K.; Grize, L.; Tamm, M.; Stolz, D. Variability of fractional exhaled nitric oxide is associated with the risk and aetiology of COPD exacerbations. *Respirology* **2023**, *28*, 445–454. [\[CrossRef\]](https://doi.org/10.1111/resp.14439) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36571108)
- 105. Ambrosino, P.; Fuschillo, S.; Accardo, M.; Mosella, M.; Molino, A.; Spedicato, G.A.; Motta, A.; Maniscalco, M. Fractional Exhaled Nitric Oxide (FeNO) in Patients with Stable Chronic Obstructive Pulmonary Disease: Short-Term Variability and Potential Clinical Implications. *J. Pers. Med.* **2022**, *12*, 1906. [\[CrossRef\]](https://doi.org/10.3390/jpm12111906) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36422082)
- 106. Vincken, S.; Sylvia, V.; Daniel, S.; Thomas, E.; Eef, V. The role of FeNO in stable COPD patients with eosinophilic airway inflammation. *Respir. Med.* **2021**, *181*, 106377. [\[CrossRef\]](https://doi.org/10.1016/j.rmed.2021.106377) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33838525)
- 107. Flynn, C.; Brightling, C. Is FeNOtyping in COPD the path to precision medicine? *Respirology* **2023**, *28*, 421–422. [\[CrossRef\]](https://doi.org/10.1111/resp.14474) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36811260)
- 108. Hogman, M.; Holmkvist, T.; Wegener, T.; Emtner, M.; Andersson, M.; Hedenstrom, H.; Merilainen, P. Extended NO analysis applied to patients with COPD, allergic asthma and allergic rhinitis. *Respir. Med.* **2002**, *96*, 24–30. [\[CrossRef\]](https://doi.org/10.1053/rmed.2001.1204) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11863206)
- 109. Roy, K.; Borrill, Z.L.; Starkey, C.; Hazel, A.L.; Morris, J.; Vestbo, J.; Singh, D. Use of different exhaled nitric oxide multiple flow rate models in COPD. *Eur. Respir. J.* **2007**, *29*, 651–659. [\[CrossRef\]](https://doi.org/10.1183/09031936.00149706) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17301096)
- 110. Paraskakis, E.; Sarikloglou, E.; Fouzas, S.; Steiropoulos, P.; Tsalkidis, A.; Bush, A. Improved prediction of asthma exacerbations by measuring distal airway inflammation. *Eur. Respir. J.* **2022**, *60*, 2101684. [\[CrossRef\]](https://doi.org/10.1183/13993003.01684-2021) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35086836)
- 111. Lazar, Z.; Kelemen, A.; Galffy, G.; Losonczy, G.; Horvath, I.; Bikov, A. Central and peripheral airway nitric oxide in patients with stable and exacerbated chronic obstructive pulmonary disease. *J. Breath. Res.* **2018**, *12*, 036017. [\[CrossRef\]](https://doi.org/10.1088/1752-7163/aac10a)
- 112. van Veen, I.H.; Sterk, P.J.; Schot, R.; Gauw, S.A.; Rabe, K.F.; Bel, E.H. Alveolar nitric oxide versus measures of peripheral airway dysfunction in severe asthma. *Eur. Respir. J.* **2006**, *27*, 951–956. [\[CrossRef\]](https://doi.org/10.1183/09031936.06.00087905)
- 113. Berry, M.; Hargadon, B.; Morgan, A.; Shelley, M.; Richter, J.; Shaw, D.; Green, R.H.; Brightling, C.; Wardlaw, A.J.; Pavord, I.D. Alveolar nitric oxide in adults with asthma: Evidence of distal lung inflammation in refractory asthma. *Eur. Respir. J.* **2005**, *25*, 986–991. [\[CrossRef\]](https://doi.org/10.1183/09031936.05.00132404) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15929952)
- 114. Hogman, M.; Thornadtsson, A.; Broms, K.; Janson, C.; Lisspers, K.; Stallberg, B.; Hedenstrom, H.; Malinovschi, A. Higher alveolar nitric oxide in COPD is related to poorer physical capacity and lower oxygen saturation after physical testing. *Eur. Respir. J.* **2019**, *54*, 1900263. [\[CrossRef\]](https://doi.org/10.1183/13993003.00263-2019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31073082)
- 115. Hogman, M.; Palm, A.; Sulku, J.; Stallberg, B.; Lisspers, K.; Broms, K.; Janson, C.; Malinovschi, A. Alveolar Nitric Oxide in Chronic Obstructive Pulmonary Disease-A Two-Year Follow-Up. *Biomedicines* **2022**, *10*, 2212. [\[CrossRef\]](https://doi.org/10.3390/biomedicines10092212) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36140313)
- 116. Corsico, A.G.; De Amici, M.; Ronzoni, V.; Giunta, V.; Mennitti, M.C.; Viscardi, A.; Marseglia, G.L.; Ciprandi, G. Allergen-specific immunoglobulin E and allergic rhinitis severity. *Allergy Rhinol.* **2017**, *8*, 1–4. [\[CrossRef\]](https://doi.org/10.2500/ar.2017.8.0187) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28381320)
- 117. Kocks, J.W.H.; Andringa, H.J.H.; van Heijst, E.; Louis, R.; Ojanguren Arranz, I.; Riemersma, R.A.; Ryan, D.; Usmani, O.S.; Flokstra-de Blok, B.M.J. Aeroallergen sensitization for detecting asthma in primary care: A diagnostic test accuracy study. *Clin. Exp. Allergy* **2021**, *51*, 1080–1084. [\[CrossRef\]](https://doi.org/10.1111/cea.13888) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33914988)
- 118. Tran, T.N.; Zeiger, R.S.; Peters, S.P.; Colice, G.; Newbold, P.; Goldman, M.; Chipps, B.E. Overlap of atopic, eosinophilic, and TH2-high asthma phenotypes in a general population with current asthma. *Ann. Allergy Asthma Immunol.* **2016**, *116*, 37–42. [\[CrossRef\]](https://doi.org/10.1016/j.anai.2015.10.027) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26707771)
- 119. Gevaert, P.; Wong, K.; Millette, L.A.; Carr, T.F. The Role of IgE in Upper and Lower Airway Disease: More Than Just Allergy! *Clin. Rev. Allergy Immunol.* **2022**, *62*, 200–215. [\[CrossRef\]](https://doi.org/10.1007/s12016-021-08901-1)
- 120. Guida, G.; Bertolini, F.; Carriero, V.; Levra, S.; Sprio, A.E.; Sciolla, M.; Orpheu, G.; Arrigo, E.; Pizzimenti, S.; Ciprandi, G.; et al. Reliability of Total Serum IgE Levels to Define Type 2 High and Low Asthma Phenotypes. *J. Clin. Med.* **2023**, *12*, 5447. [\[CrossRef\]](https://doi.org/10.3390/jcm12175447) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37685515)
- 121. Frossing, L.; Silberbrandt, A.; Von Bulow, A.; Backer, V.; Porsbjerg, C. The Prevalence of Subtypes of Type 2 Inflammation in an Unselected Population of Patients with Severe Asthma. *J. Allergy Clin. Immunol. Pract.* **2021**, *9*, 1267–1275. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2020.09.051) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33039645)
- 122. Hanania, N.A.; Wenzel, S.; Rosen, K.; Hsieh, H.J.; Mosesova, S.; Choy, D.F.; Lal, P.; Arron, J.R.; Harris, J.M.; Busse, W. Exploring the effects of omalizumab in allergic asthma: An analysis of biomarkers in the EXTRA study. *Am. J. Respir. Crit. Care Med.* **2013**, *187*, 804–811. [\[CrossRef\]](https://doi.org/10.1164/rccm.201208-1414OC)
- 123. Omenaas, E.; Bakke, P.; Elsayed, S.; Hanoa, R.; Gulsvik, A. Total and specific serum IgE levels in adults: Relationship to sex, age and environmental factors. *Clin. Exp. Allergy* **1994**, *24*, 530–539. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2222.1994.tb00950.x)
- 124. De Amici, M.; Ciprandi, G. The Age Impact on Serum Total and Allergen-Specific IgE. *Allergy Asthma Immunol. Res.* **2013**, *5*, 170–174. [\[CrossRef\]](https://doi.org/10.4168/aair.2013.5.3.170) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23638316)
- 125. Vidal, C.; Armisen, M.; Dominguez-Santalla, M.J.; Gude, F.; Lojo, S.; Gonzalez-Quintela, A. Influence of alcohol consumption on serum immunoglobulin E levels in atopic and nonatopic adults. *Alcohol. Clin. Exp. Res.* **2002**, *26*, 59–64. [\[CrossRef\]](https://doi.org/10.1111/j.1530-0277.2002.tb02432.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11821655)
- 126. Burrows, B.; Halonen, M.; Barbee, R.A.; Lebowitz, M.D. The relationship of serum immunoglobulin E to cigarette smoking. *Am. Rev. Respir. Dis.* **1981**, *124*, 523–525. [\[CrossRef\]](https://doi.org/10.1164/arrd.1981.124.5.523) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7305105)
- 127. Woolnough, K.; Fairs, A.; Pashley, C.H.; Wardlaw, A.J. Allergic fungal airway disease: Pathophysiologic and diagnostic considerations. *Curr. Opin. Pulm. Med.* **2015**, *21*, 39–47. [\[CrossRef\]](https://doi.org/10.1097/MCP.0000000000000129) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25415407)
- 128. Knutsen, A.P.; Bush, R.K.; Demain, J.G.; Denning, D.W.; Dixit, A.; Fairs, A.; Greenberger, P.A.; Kariuki, B.; Kita, H.; Kurup, V.P.; et al. Fungi and allergic lower respiratory tract diseases. *J. Allergy Clin. Immunol.* **2012**, *129*, 280–291; quiz 292–283. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2011.12.970) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22284927)
- 129. Settipane, G.A.; Pudupakkam, R.K.; McGowan, J.H. Corticosteroid effect on immunoglobulins. *J. Allergy Clin. Immunol.* **1978**, *62*, 162–166. [\[CrossRef\]](https://doi.org/10.1016/0091-6749(78)90101-X) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/681628)
- 130. Ohrui, T.; Funayama, T.; Sekizawa, K.; Yamaya, M.; Sasaki, H. Effects of inhaled beclomethasone dipropionate on serum IgE levels and clinical symptoms in atopic asthma. *Clin. Exp. Allergy* **1999**, *29*, 357–361. [\[CrossRef\]](https://doi.org/10.1046/j.1365-2222.1999.00474.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10202343)
- 131. Murayama, N.; Doi, S.; Inoue, T.; Takamatsu, I.; Kameda, M.; Takeda, K.; Toyoshima, K. Inhaled steroid inhibits development of total and mite IgE. *Immunol. Med.* **2018**, *41*, 17–22. [\[CrossRef\]](https://doi.org/10.1080/09114300.2018.1451599)
- 132. Zieg, G.; Lack, G.; Harbeck, R.J.; Gelfand, E.W.; Leung, D.Y. In vivo effects of glucocorticoids on IgE production. *J. Allergy Clin. Immunol.* **1994**, *94*, 222–230. [\[CrossRef\]](https://doi.org/10.1053/ai.1994.v94.a54936)
- 133. Kerstjens, H.A.; Schouten, J.P.; Brand, P.L.; Schoonbrood, D.F.; Sterk, P.J.; Postma, D.S. Importance of total serum IgE for improvement in airways hyperresponsiveness with inhaled corticosteroids in asthma and chronic obstructive pulmonary disease. The Dutch CNSLD Study Group. *Am. J. Respir. Crit. Care Med.* **1995**, *151*, 360–368. [\[CrossRef\]](https://doi.org/10.1164/ajrccm.151.2.7842192) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7842192)
- 134. Sonnenberg-Riethmacher, E.; Miehe, M.; Riethmacher, D. Periostin in Allergy and Inflammation. *Front. Immunol.* **2021**, *12*, 722170. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2021.722170) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34512647)
- 135. Luu Quoc, Q.; Moon, J.Y.; Lee, D.H.; Ban, G.Y.; Kim, S.H.; Park, H.S. Role of Thymus and Activation-Regulated Chemokine in Allergic Asthma. *J. Asthma Allergy* **2022**, *15*, 157–167. [\[CrossRef\]](https://doi.org/10.2147/JAA.S351720) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35153491)
- 136. Crump, M.P.; Rajarathnam, K.; Kim, K.S.; Clark-Lewis, I.; Sykes, B.D. Solution structure of eotaxin, a chemokine that selectively recruits eosinophils in allergic inflammation. *J. Biol. Chem.* **1998**, *273*, 22471–22479. [\[CrossRef\]](https://doi.org/10.1074/jbc.273.35.22471)
- 137. Dent, G.; Hadjicharalambous, C.; Yoshikawa, T.; Handy, R.L.; Powell, J.; Anderson, I.K.; Louis, R.; Davies, D.E.; Djukanovic, R. Contribution of eotaxin-1 to eosinophil chemotactic activity of moderate and severe asthmatic sputum. *Am. J. Respir. Crit. Care Med.* **2004**, *169*, 1110–1117. [\[CrossRef\]](https://doi.org/10.1164/rccm.200306-855OC) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15001461)
- 138. Silkoff, P.E.; Laviolette, M.; Singh, D.; FitzGerald, J.M.; Kelsen, S.; Backer, V.; Porsbjerg, C.M.; Girodet, P.O.; Berger, P.; Kline, J.N.; et al. Identification of airway mucosal type 2 inflammation by using clinical biomarkers in asthmatic patients. *J. Allergy Clin. Immunol.* **2017**, *140*, 710–719. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2016.11.038) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28089872)
- 139. Gokani, S.A.; Espehana, A.; Pratas, A.C.; Luke, L.; Sharma, E.; Mattock, J.; Gavrilovic, J.; Clark, A.; Wileman, T.; Philpott, C.M. Systematic Review of Protein Biomarkers in Adult Patients with Chronic Rhinosinusitis. *Am. J. Rhinol. Allergy* **2023**, *37*, 705–729. [\[CrossRef\]](https://doi.org/10.1177/19458924231190568) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37491901)
- 140. Bachert, C.; Han, J.K.; Desrosiers, M.; Hellings, P.W.; Amin, N.; Lee, S.E.; Mullol, J.; Greos, L.S.; Bosso, J.V.; Laidlaw, T.M.; et al. Efficacy and safety of dupilumab in patients with severe chronic rhinosinusitis with nasal polyps (LIBERTY NP SINUS-24 and LIBERTY NP SINUS-52): Results from two multicentre, randomised, double-blind, placebo-controlled, parallel-group phase 3 trials. *Lancet* **2019**, *394*, 1638–1650. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(19)31881-1)
- 141. Terry, P.D.; Dhand, R. The 2023 GOLD Report: Updated Guidelines for Inhaled Pharmacological Therapy in Patients with Stable COPD. *Pulm. Ther.* **2023**, *9*, 345–357. [\[CrossRef\]](https://doi.org/10.1007/s41030-023-00233-z)
- 142. Global Strategy for Asthma Management and Prevention. 2024 GINA Main Report. Available online: [https://ginasthma.org/20](https://ginasthma.org/2024-report/) [24-report/](https://ginasthma.org/2024-report/) (accessed on 6 June 2024).
- 143. Rank, M.A.; Chu, D.K.; Bognanni, A.; Oykhman, P.; Bernstein, J.A.; Ellis, A.K.; Golden, D.B.K.; Greenhawt, M.; Horner, C.C.; Ledford, D.K.; et al. The Joint Task Force on Practice Parameters GRADE guidelines for the medical management of chronic rhinosinusitis with nasal polyposis. *J. Allergy Clin. Immunol.* **2023**, *151*, 386–398. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2022.10.026)
- 144. van der Veen, J.; Seys, S.F.; Timmermans, M.; Levie, P.; Jorissen, M.; Fokkens, W.J.; Hellings, P.W. Real-life study showing uncontrolled rhinosinusitis after sinus surgery in a tertiary referral centre. *Allergy* **2017**, *72*, 282–290. [\[CrossRef\]](https://doi.org/10.1111/all.12983) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27392210)
- 145. Centers for Disease Control and Prevention. Uncontrolled Asthma among Adults. 2019. Available online: [https:](https://archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-adults-2019.htm#:~:text=60.1%25%20of%20adults%20with%20current,%E2%80%9334%20years%20(53.3%25)) [//archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-adults-2019.htm#:~:text=60.1%25%20of%](https://archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-adults-2019.htm#:~:text=60.1%25%20of%20adults%20with%20current,%E2%80%9334%20years%20(53.3%25)) [20adults%20with%20current,%E2%80%9334%20years%20\(53.3%25\)](https://archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-adults-2019.htm#:~:text=60.1%25%20of%20adults%20with%20current,%E2%80%9334%20years%20(53.3%25)) (accessed on 6 June 2024).
- 146. Centers for Disease Control and Prevention. Uncontrolled Asthma among Children with Current Asthma, 2018–2020. Available online: [https://archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-children-2018-2020.htm#:~:](https://archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-children-2018-2020.htm#:~:text=Uncontrolled%20Asthma%20Among%20Children%20With%20Current%20Asthma,%202018%E2%80%932020,-Forty-four%20percent&text=About%2044.0%25%20of%20children%20with,%25)%20(sex%20at%20birth)) [text=Uncontrolled%20Asthma%20Among%20Children%20With%20Current%20Asthma,%202018%E2%80%932020,-Forty](https://archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-children-2018-2020.htm#:~:text=Uncontrolled%20Asthma%20Among%20Children%20With%20Current%20Asthma,%202018%E2%80%932020,-Forty-four%20percent&text=About%2044.0%25%20of%20children%20with,%25)%20(sex%20at%20birth))[four%20percent&text=About%2044.0%25%20of%20children%20with,%25\)%20\(sex%20at%20birth\)](https://archive.cdc.gov/www_cdc_gov/asthma/asthma_stats/uncontrolled-asthma-children-2018-2020.htm#:~:text=Uncontrolled%20Asthma%20Among%20Children%20With%20Current%20Asthma,%202018%E2%80%932020,-Forty-four%20percent&text=About%2044.0%25%20of%20children%20with,%25)%20(sex%20at%20birth)) (accessed on 6 June 2024).
- 147. Wilkinson, T.M.A.; Aris, E.; Bourne, S.C.; Clarke, S.C.; Peeters, M.; Pascal, T.G.; Taddei, L.; Tuck, A.C.; Kim, V.L.; Ostridge, K.K.; et al. Drivers of year-to-year variation in exacerbation frequency of COPD: Analysis of the AERIS cohort. *ERJ Open Res.* **2019**, *5*, 00248-2018. [\[CrossRef\]](https://doi.org/10.1183/23120541.00248-2018) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30815467)
- 148. David, B.; Bafadhel, M.; Koenderman, L.; De Soyza, A. Eosinophilic inflammation in COPD: From an inflammatory marker to a treatable trait. *Thorax* **2021**, *76*, 188–195. [\[CrossRef\]](https://doi.org/10.1136/thoraxjnl-2020-215167) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33122447)
- 149. Agusti, A.; Bafadhel, M.; Beasley, R.; Bel, E.H.; Faner, R.; Gibson, P.G.; Louis, R.; McDonald, V.M.; Sterk, P.J.; Thomas, M.; et al. Precision medicine in airway diseases: Moving to clinical practice. *Eur. Respir. J.* **2017**, *50*, 1701655. [\[CrossRef\]](https://doi.org/10.1183/13993003.01655-2017) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29051276)
- 150. Lowe, P.J.; Tannenbaum, S.; Gautier, A.; Jimenez, P. Relationship between omalizumab pharmacokinetics, IgE pharmacodynamics and symptoms in patients with severe persistent allergic (IgE-mediated) asthma. *Br. J. Clin. Pharmacol.* **2009**, *68*, 61–76. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2125.2009.03401.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19660004)
- 151. Soler, M.; Matz, J.; Townley, R.; Buhl, R.; O'Brien, J.; Fox, H.; Thirlwell, J.; Gupta, N.; Della Cioppa, G. The anti-IgE antibody omalizumab reduces exacerbations and steroid requirement in allergic asthmatics. *Eur. Respir. J.* **2001**, *18*, 254–261. [\[CrossRef\]](https://doi.org/10.1183/09031936.01.00092101) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11529281)
- 152. Busse, W.; Corren, J.; Lanier, B.Q.; McAlary, M.; Fowler-Taylor, A.; Cioppa, G.D.; van As, A.; Gupta, N. Omalizumab, anti-IgE recombinant humanized monoclonal antibody, for the treatment of severe allergic asthma. *J. Allergy Clin. Immunol.* **2001**, *108*, 184–190. [\[CrossRef\]](https://doi.org/10.1067/mai.2001.117880)
- 153. Milgrom, H.; Berger, W.; Nayak, A.; Gupta, N.; Pollard, S.; McAlary, M.; Taylor, A.F.; Rohane, P. Treatment of childhood asthma with anti-immunoglobulin E antibody (omalizumab). *Pediatrics* **2001**, *108*, E36. [\[CrossRef\]](https://doi.org/10.1542/peds.108.2.e36) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11483846)
- 154. Gevaert, P.; Omachi, T.A.; Corren, J.; Mullol, J.; Han, J.; Lee, S.E.; Kaufman, D.; Ligueros-Saylan, M.; Howard, M.; Zhu, R.; et al. Efficacy and safety of omalizumab in nasal polyposis: 2 randomized phase 3 trials. *J. Allergy Clin. Immunol.* **2020**, *146*, 595–605. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2020.05.032)
- 155. Maltby, S.; Gibson, P.G.; Powell, H.; McDonald, V.M. Omalizumab Treatment Response in a Population with Severe Allergic Asthma and Overlapping COPD. *Chest* **2017**, *151*, 78–89. [\[CrossRef\]](https://doi.org/10.1016/j.chest.2016.09.035)
- 156. Tsabouri, S.; Ntritsos, G.; Koskeridis, F.; Evangelou, E.; Olsson, P.; Kostikas, K. Omalizumab for the treatment of allergic rhinitis: A systematic review and meta-analysis. *Rhinology* **2021**, *59*, 501–510. [\[CrossRef\]](https://doi.org/10.4193/Rhin21.159) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34714895)
- 157. Zhang, Y.; Xi, L.; Gao, Y.; Huang, Y.; Cao, F.; Xiong, W.; Wang, C.; Zhang, L. Omalizumab is effective in the preseasonal treatment of seasonal allergic rhinitis. *Clin. Transl. Allergy* **2022**, *12*, e12094. [\[CrossRef\]](https://doi.org/10.1002/clt2.12094) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35024137)
- 158. Muller, M.; Igarashi, A.; Hashiguchi, K.; Kappel, M.; Paolini, F.; Yoshisue, H.; Funakubo, M.; Sharma, H.; Okano, M. The impact of omalizumab on paid and unpaid work productivity among severe Japanese cedar pollinosis (JCP) patients. *J. Med. Econ.* **2022**, *25*, 220–229. [\[CrossRef\]](https://doi.org/10.1080/13696998.2022.2033051)
- 159. Hamilton, R.G. Monitoring Allergic Patients on Omalizumab with Free and Total Serum IgE Measurements. *J. Allergy Clin. Immunol. Pract.* **2016**, *4*, 366–368. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2015.12.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26968963)
- 160. Cildag, S.; Senturk, T. The effect of omalizumab treatment on IgE and other immunoglobulin levels in patients with chronic spontaneous urticaria and its association with treatment response. *Postep. Dermatol. Alergol.* **2018**, *35*, 516–519. [\[CrossRef\]](https://doi.org/10.5114/ada.2017.71422) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30429712)
- 161. Menzella, F.; Just, J.; Sauerbeck, I.S.; Mailaender, C.; Saccheri, F.; Thonnelier, C.; Jaumont, X.; Mala, L. Omalizumab for the treatment of patients with severe allergic asthma with immunoglobulin E levels above >1500 IU/mL. *World Allergy Organ. J.* **2023**, *16*, 100787. [\[CrossRef\]](https://doi.org/10.1016/j.waojou.2023.100787) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37332525)
- 162. Holguin, F.; Cardet, J.C.; Chung, K.F.; Diver, S.; Ferreira, D.S.; Fitzpatrick, A.; Gaga, M.; Kellermeyer, L.; Khurana, S.; Knight, S.; et al. Management of severe asthma: A European Respiratory Society/American Thoracic Society guideline. *Eur. Respir. J.* **2020**, *55*, 1900588. [\[CrossRef\]](https://doi.org/10.1183/13993003.00588-2019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31558662)
- 163. Casale, T.B.; Luskin, A.T.; Busse, W.; Zeiger, R.S.; Trzaskoma, B.; Yang, M.; Griffin, N.M.; Chipps, B.E. Omalizumab Effectiveness by Biomarker Status in Patients with Asthma: Evidence From PROSPERO, A Prospective Real-World Study. *J. Allergy Clin. Immunol. Pract.* **2019**, *7*, 156–164.e1. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2018.04.043) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29800752)
- 164. Li, Y.; Li, X.; Zhang, B.; Yu, Q.; Lu, Y. Predictive biomarkers for response to omalizumab in patients with severe allergic asthma: A meta-analysis. *Expert Rev. Respir. Med.* **2022**, *16*, 1023–1033. [\[CrossRef\]](https://doi.org/10.1080/17476348.2022.2092100)
- 165. Khurana, S.; Brusselle, G.G.; Bel, E.H.; FitzGerald, J.M.; Masoli, M.; Korn, S.; Kato, M.; Albers, F.C.; Bradford, E.S.; Gilson, M.J.; et al. Long-term Safety and Clinical Benefit of Mepolizumab in Patients with the Most Severe Eosinophilic Asthma: The COSMEX Study. *Clin. Ther.* **2019**, *41*, 2041–2056.e2045. [\[CrossRef\]](https://doi.org/10.1016/j.clinthera.2019.07.007)
- 166. Ortega, H.G.; Yancey, S.W.; Mayer, B.; Gunsoy, N.B.; Keene, O.N.; Bleecker, E.R.; Brightling, C.E.; Pavord, I.D. Severe eosinophilic asthma treated with mepolizumab stratified by baseline eosinophil thresholds: A secondary analysis of the DREAM and MENSA studies. *Lancet Respir. Med.* **2016**, *4*, 549–556. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(16)30031-5)
- 167. Bleecker, E.R.; FitzGerald, J.M.; Chanez, P.; Papi, A.; Weinstein, S.F.; Barker, P.; Sproule, S.; Gilmartin, G.; Aurivillius, M.; Werkstrom, V.; et al. Efficacy and safety of benralizumab for patients with severe asthma uncontrolled with high-dosage inhaled corticosteroids and long-acting β² -agonists (SIROCCO): A randomised, multicentre, placebo-controlled phase 3 trial. *Lancet* **2016**, *388*, 2115–2127. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(16)31324-1)
- 168. FitzGerald, J.M.; Bleecker, E.R.; Nair, P.; Korn, S.; Ohta, K.; Lommatzsch, M.; Ferguson, G.T.; Busse, W.W.; Barker, P.; Sproule, S.; et al. Benralizumab, an anti-interleukin-5 receptor alpha monoclonal antibody, as add-on treatment for patients with severe, uncontrolled, eosinophilic asthma (CALIMA): A randomised, double-blind, placebo-controlled phase 3 trial. *Lancet* **2016**, *388*, 2128–2141. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(16)31322-8)
- 169. Jackson, D.J.; Heaney, L.G.; Humbert, M.; Kent, B.D.; Shavit, A.; Hiljemark, L.; Olinger, L.; Cohen, D.; Menzies-Gow, A.; Korn, S.; et al. Reduction of daily maintenance inhaled corticosteroids in patients with severe eosinophilic asthma treated with benralizumab (SHAMAL): A randomised, multicentre, open-label, phase 4 study. *Lancet* **2024**, *403*, 271–281. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(23)02284-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38071986)
- 170. Weinstein, S.F.; Katial, R.K.; Bardin, P.; Korn, S.; McDonald, M.; Garin, M.; Bateman, E.D.; Hoyte, F.C.L.; Germinaro, M. Effects of Reslizumab on Asthma Outcomes in a Subgroup of Eosinophilic Asthma Patients with Self-Reported Chronic Rhinosinusitis with Nasal Polyps. *J. Allergy Clin. Immunol. Pract.* **2019**, *7*, 589–596.e3. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2018.08.021) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30193936)
- 171. Castro, M.; Mathur, S.; Hargreave, F.; Boulet, L.P.; Xie, F.; Young, J.; Wilkins, H.J.; Henkel, T.; Nair, P.; Res-5- Study, G. Reslizumab for poorly controlled, eosinophilic asthma: A randomized, placebo-controlled study. *Am. J. Respir. Crit. Care Med.* **2011**, *184*, 1125–1132. [\[CrossRef\]](https://doi.org/10.1164/rccm.201103-0396OC) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21852542)
- 172. Bachert, C.; Sousa, A.R.; Lund, V.J.; Scadding, G.K.; Gevaert, P.; Nasser, S.; Durham, S.R.; Cornet, M.E.; Kariyawasam, H.H.; Gilbert, J.; et al. Reduced need for surgery in severe nasal polyposis with mepolizumab: Randomized trial. *J. Allergy Clin. Immunol.* **2017**, *140*, 1024–1031.e14. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2017.05.044) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28687232)
- 173. Han, J.K.; Bachert, C.; Fokkens, W.; Desrosiers, M.; Wagenmann, M.; Lee, S.E.; Smith, S.G.; Martin, N.; Mayer, B.; Yancey, S.W.; et al. Mepolizumab for chronic rhinosinusitis with nasal polyps (SYNAPSE): A randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Respir. Med.* **2021**, *9*, 1141–1153. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(21)00097-7)
- 174. Pavord, I.D.; Chapman, K.R.; Bafadhel, M.; Sciurba, F.C.; Bradford, E.S.; Schweiker Harris, S.; Mayer, B.; Rubin, D.B.; Yancey, S.W.; Paggiaro, P. Mepolizumab for Eosinophil-Associated COPD: Analysis of METREX and METREO. *Int. J. Chron. Obs. Pulmon Dis.* **2021**, *16*, 1755–1770. [\[CrossRef\]](https://doi.org/10.2147/COPD.S294333)
- 175. Criner, G.J.; Celli, B.R.; Brightling, C.E.; Agusti, A.; Papi, A.; Singh, D.; Sin, D.D.; Vogelmeier, C.F.; Sciurba, F.C.; Bafadhel, M.; et al. Benralizumab for the Prevention of COPD Exacerbations. *N. Engl. J. Med.* **2019**, *381*, 1023–1034. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa1905248)
- 176. Contoli, M.; Santus, P.; Menzella, F.; Rocchi, C.; Radovanovic, D.; Baraldi, F.; Martelli, C.; Casanova, S.; Barbetta, C.; Micheletto, C.; et al. Effects of anti-IL5 biological treatments on blood IgE levels in severe asthmatic patients: A real-life multicentre study (BIONIGE). *Clin. Transl. Allergy* **2022**, *12*, e12143. [\[CrossRef\]](https://doi.org/10.1002/clt2.12143)
- 177. Menigoz, C.; Dirou, S.; Chambellan, A.; Hassoun, D.; Moui, A.; Magnan, A.; Blanc, F.X. Use of FeNO to predict anti-IL-5 and IL-5R biologics efficacy in a real-world cohort of adults with severe eosinophilic asthma. *J. Asthma* **2023**, *60*, 1162–1170. [\[CrossRef\]](https://doi.org/10.1080/02770903.2022.2136526)
- 178. Jang, J.H.; Woo, S.D.; Lee, Y.; Kim, C.K.; Shin, Y.S.; Ye, Y.M.; Park, H.S. Changes in Type 2 Biomarkers After Anti-IL5 Treatment in Patients with Severe Eosinophilic Asthma. *Allergy Asthma Immunol. Res.* **2021**, *13*, 330–338. [\[CrossRef\]](https://doi.org/10.4168/aair.2021.13.2.330) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33474865)
- 179. Harb, H.; Chatila, T.A. Mechanisms of Dupilumab. *Clin. Exp. Allergy* **2020**, *50*, 5–14. [\[CrossRef\]](https://doi.org/10.1111/cea.13491)
- 180. Castro, M.; Corren, J.; Pavord, I.D.; Maspero, J.; Wenzel, S.; Rabe, K.F.; Busse, W.W.; Ford, L.; Sher, L.; FitzGerald, J.M.; et al. Dupilumab Efficacy and Safety in Moderate-to-Severe Uncontrolled Asthma. *N. Engl. J. Med.* **2018**, *378*, 2486–2496. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa1804092)
- 181. Bacharier, L.B.; Maspero, J.F.; Katelaris, C.H.; Fiocchi, A.G.; Gagnon, R.; de Mir, I.; Jain, N.; Sher, L.D.; Mao, X.; Liu, D.; et al. Dupilumab in Children with Uncontrolled Moderate-to-Severe Asthma. *N. Engl. J. Med.* **2021**, *385*, 2230–2240. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa2106567)
- 182. Rabe, K.F.; FitzGerald, J.M.; Bateman, E.D.; Castro, M.; Pavord, I.D.; Maspero, J.F.; Busse, W.W.; Izuhara, K.; Daizadeh, N.; Ortiz, B.; et al. Dupilumab Is Effective in Patients with Moderate-to-Severe Uncontrolled GINA-Defined Type 2 Asthma Irrespective of an Allergic Asthma Phenotype. *J. Allergy Clin. Immunol. Pract.* **2022**, *10*, 2916–2924.e4. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2022.06.036)
- 183. Weinstein, S.F.; Katial, R.; Jayawardena, S.; Pirozzi, G.; Staudinger, H.; Eckert, L.; Joish, V.N.; Amin, N.; Maroni, J.; Rowe, P.; et al. Efficacy and safety of dupilumab in perennial allergic rhinitis and comorbid asthma. *J. Allergy Clin. Immunol.* **2018**, *142*, 171–177.e1. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2017.11.051) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29355679)
- 184. Campion, N.J.; Doralt, A.; Lupinek, C.; Berger, M.; Poglitsch, K.; Brugger, J.; Quint, T.; Gangl, K.; Sinz, C.; Bartosik, T.; et al. Dupilumab reduces symptom burden in allergic rhinitis and suppresses allergen-specific IgE production. *Allergy* **2023**, *78*, 1687–1691. [\[CrossRef\]](https://doi.org/10.1111/all.15653) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36691369)
- 185. Bhatt, S.P.; Rabe, K.F.; Hanania, N.A.; Vogelmeier, C.F.; Cole, J.; Bafadhel, M.; Christenson, S.A.; Papi, A.; Singh, D.; Laws, E.; et al. Dupilumab for COPD with Type 2 Inflammation Indicated by Eosinophil Counts. *N. Engl. J. Med.* **2023**, *389*, 205–214. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa2303951)
- 186. Bacharier, L.B.; Guilbert, T.W.; Katelaris, C.H.; Deschildre, A.; Phipatanakul, W.; Liu, D.; Altincatal, A.; Mannent, L.P.; Amin, N.; Laws, E.; et al. Dupilumab Improves Lung Function Parameters in Pediatric Type 2 Asthma: VOYAGE Study. *J. Allergy Clin. Immunol. Pract.* **2024**, *12*, 948–959. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2023.12.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38092225)
- 187. Pavord, I.D.; Deniz, Y.; Corren, J.; Casale, T.B.; FitzGerald, J.M.; Izuhara, K.; Daizadeh, N.; Ortiz, B.; Johnson, R.R.; Harel, S.; et al. Baseline FeNO Independently Predicts the Dupilumab Response in Patients with Moderate-to-Severe Asthma. *J. Allergy Clin. Immunol. Pract.* **2023**, *11*, 1213–1220.e2. [\[CrossRef\]](https://doi.org/10.1016/j.jaip.2022.11.043)
- 188. Corren, J.; Parnes, J.R.; Wang, L.; Mo, M.; Roseti, S.L.; Griffiths, J.M.; van der Merwe, R. Tezepelumab in Adults with Uncontrolled Asthma. *N. Engl. J. Med.* **2017**, *377*, 936–946. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa1704064)
- 189. Menzies-Gow, A.; Corren, J.; Bourdin, A.; Chupp, G.; Israel, E.; Wechsler, M.E.; Brightling, C.E.; Griffiths, J.M.; Hellqvist, A.; Bowen, K.; et al. Tezepelumab in Adults and Adolescents with Severe, Uncontrolled Asthma. *N. Engl. J. Med.* **2021**, *384*, 1800–1809. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa2034975)
- 190. Diver, S.; Khalfaoui, L.; Emson, C.; Wenzel, S.E.; Menzies-Gow, A.; Wechsler, M.E.; Johnston, J.; Molfino, N.; Parnes, J.R.; Megally, A.; et al. Effect of tezepelumab on airway inflammatory cells, remodelling, and hyperresponsiveness in patients with moderate-to-severe uncontrolled asthma (CASCADE): A double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet Respir. Med.* **2021**, *9*, 1299–1312. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(21)00226-5)
- 191. Corren, J.; Brightling, C.E.; Boulet, L.P.; Porsbjerg, C.; Wechsler, M.E.; Menzies-Gow, A.; Ambrose, C.S.; Cook, B.; Martin, N.; Spahn, J.; et al. Not just an anti-eosinophil drug: Tezepelumab treatment for type 2 asthma and beyond. *Eur. Respir. J.* **2023**, *61*, 2202202. [\[CrossRef\]](https://doi.org/10.1183/13993003.02202-2022)
- 192. England, E.; Rees, D.G.; Scott, I.C.; Carmen, S.; Chan, D.T.Y.; Chaillan Huntington, C.E.; Houslay, K.F.; Erngren, T.; Penney, M.; Majithiya, J.B.; et al. Tozorakimab (MEDI3506): An anti-IL-33 antibody that inhibits IL-33 signalling via ST2 and RAGE/EGFR to reduce inflammation and epithelial dysfunction. *Sci. Rep.* **2023**, *13*, 9825. [\[CrossRef\]](https://doi.org/10.1038/s41598-023-36642-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37330528)
- 193. Wechsler, M.E.; Ruddy, M.K.; Pavord, I.D.; Israel, E.; Rabe, K.F.; Ford, L.B.; Maspero, J.F.; Abdulai, R.M.; Hu, C.C.; Martincova, R.; et al. Efficacy and Safety of Itepekimab in Patients with Moderate-to-Severe Asthma. *N. Engl. J. Med.* **2021**, *385*, 1656–1668. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa2024257)
- 194. Kelsen, S.G.; Agache, I.O.; Soong, W.; Israel, E.; Chupp, G.L.; Cheung, D.S.; Theess, W.; Yang, X.; Staton, T.L.; Choy, D.F.; et al. Astegolimab (anti-ST2) efficacy and safety in adults with severe asthma: A randomized clinical trial. *J. Allergy Clin. Immunol.* **2021**, *148*, 790–798. [\[CrossRef\]](https://doi.org/10.1016/j.jaci.2021.03.044) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33872652)
- 195. Yousuf, A.J.; Mohammed, S.; Carr, L.; Yavari Ramsheh, M.; Micieli, C.; Mistry, V.; Haldar, K.; Wright, A.; Novotny, P.; Parker, S.; et al. Astegolimab, an anti-ST2, in chronic obstructive pulmonary disease (COPD-ST2OP): A phase 2a, placebo-controlled trial. *Lancet Respir. Med.* **2022**, *10*, 469–477. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(21)00556-7)
- 196. Lee, A.L.; Burge, A.T. Adding Up the Value of Pulmonary Rehabilitation in COPD: Money, Morbidity, and Mortality. *Chest* **2024**, *165*, 231–232. [\[CrossRef\]](https://doi.org/10.1016/j.chest.2023.10.029) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38336430)
- 197. Jenkins, A.R.; Burtin, C.; Camp, P.G.; Lindenauer, P.; Carlin, B.; Alison, J.A.; Rochester, C.; Holland, A.E. Do pulmonary rehabilitation programmes improve outcomes in patients with COPD posthospital discharge for exacerbation: A systematic review and meta-analysis. *Thorax* **2024**, *79*, 438–447. [\[CrossRef\]](https://doi.org/10.1136/thorax-2023-220333) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38350731)
- 198. Grossman, J. One airway, one disease. *Chest* **1997**, *111*, 11S–16S. [\[CrossRef\]](https://doi.org/10.1378/chest.111.2_Supplement.11S) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9042022)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.